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The Tachykinin Peptide Family

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| | Abstract | 286 |
|------|---|-----|
| I. | Introduction | 286 |
| II. | Occurrence and species distribution of tachykinin-like peptides | 287 |
| | A. Invertebrate tachykinin-like peptides | |
| | B. Prevertebrate tachykinin-like peptides | |
| | 1. Amphioxus lanceolatus | |
| | 2. Tunicata (Protocordata) | |
| | C. Submammalian vertebrate tachykinins | |
| | 1. Amphibian skin tachykinins | |
| | 2. Brain and gut tachykinins | |
| | D. Mammalian tachykinins | |
| | 1. Mammalian tachykinins and their biosynthesis | |
| III. | Localization of tachykinin-like peptides | |
| | A. Non-neuronal localization | |
| | 1. Amphibian skin | |
| | 2. Invertebrate salivary glands | |
| | 3. Normal mammalian tissues | |
| | 4. Endocrine tachykinin-secreting tumors | |
| | B. Neuronal localization | |
| IV. | Relationships between structure/activity receptor selectivity | |
| | A. Residue occupying position 7 from the C terminus | |
| | B. Residue occupying position 4 from the C terminus | |
| | C. Residue occupying position 6 from the C terminus | |
| | D. Amino acid substitutions in the C-terminal tripeptide | |
| | E. Pro residue in the N-terminal sequence | |
| V. | Tachykinin-like peptides: pharmacological actions | |
| | A. Cardiovascular system | |
| | 1. Systemic arterial blood pressure | |
| | 2. Regional circulation | 299 |
| | B. Gastrointestinal tract | |
| | 1. Motility | 301 |
| | a. In vitro experiments | 301 |
| | b. In vivo experiments | |
| | 2. Secretions | |
| | C. Airways system | 305 |
| | D. Urogenital tract | |
| | E. Immune system | |
| | F. Central nervous system | |
| | G. Pain. | |

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†This is the last unfinished review written by professor Vittorio Erspamer before he died suddenly in October 1999. His collaborators are proud to present this review on his behalf and to honor his memory as an enthusiastic and intuitive researcher who enriched the knowledge of new and unimagined agents and actions all over the world.

| . 313 |
|-------|
| . 314 |
| . 314 |
| . 314 |
| . 315 |
| . 315 |
| . 316 |
| . 316 |
| . 317 |
| |

Abstract—The tachykinin peptide family certainly represents one of the largest peptide families described in the animal organism. So far, more than 40 tachykinins have been isolated from invertebrate (insects, worms, and molluscs), protochordate, and vertebrate (skin, gastrointestinal tract, peripheral and central nervous system) tissues. Substance P (SP), first identified by bioassay as early as 1931 but sequenced only in 1971, several years after the elucidation of the structure of eledoisin from molluscan tissues and of physalaemin from amphibian skin, may be considered as a prototype of the tachykinins. Hitherto, as many as 19 tachykinins have been isolated from amphibian integument, and eight additional peptides have been isolated from amphibian gut and brain. Counterparts of skin tachykinins in mammalian tissues are SP, neurokinin A, and neurokinin B. Three main receptor subtypes for the tachykinins have been identified (NK1, NK2, and NK3), but their number is probably destined to increase. It is obvious that the peripheral and central effects of the tachykinins may substantially vary depending on the activation of different receptor subtypes. Matters are further complicated by the frequent capacity of the single tachykinins to bind, although with different affinity, to more receptors. It has been recognized that tachykinins have a variety of effects in physiological and pathological conditions, and there is evidence suggesting intrinsic neuroprotective and neurodegenerative properties of these neuropeptides. This review provides an update on the current body of knowledge regarding tachykinin occurrence and distribution in the animal kingdom, from the lowest invertebrates to man, and the physiological and pharmacological actions of tachykinins outlining the pregnant importance of this large peptide family.

I. Introduction

Seventy years ago, von Euler and Gaddum described an unidentified substance present in alcoholic extracts of equine brain and intestine that in the rabbit displayed a potent stimulant action on the jejunum and a hypotensive action that was distinct from all compounds then known to stimulate the gut and that was referred to as "P" on the tracings and the protocols.

Using semipurified preparations, numerous biological studies of its activity were carried out, but many efforts have been made to isolate the active substance. After some unsuccessful attempts on horse intestine, substance P (SP) was isolated in a pure form from bovine hypothalamus, and 40 years later, its structure was established by Chang and Leeman (1970). SP was then isolated in a pure form and sequenced also from horse intestine (Studer et al., 1973). SP was one of the most extensively studied active substances during the half-century since its discovery, and for many years, it was believed to be the only mammalian tachykinin considered to be a neuropeptide. This belief was firmly put to rest only in 1983 with the discovery of neurokinin A (NKA) and neurokinin B (NKB) (Kangawa et al.,

1983; Kimura et al., 1983) that differ from SP in their pharmacological activity, both peripheral and central, and in their preference for different tachykinin receptor subtypes.

The story of the identification of SP was very similar to that leading to the discovery of nonmammalian tachykinins. In 1947, while investigating the occurrence of biogenic amines, especially serotonin in the posterior salivary glands of a Mediterranean octopod, Eledone moschata, an unidentified substance was found that again lowered blood pressure in rabbits and dogs, stimulated isolated preparations of intestinal smooth muscle, and caused profuse salivation in dogs and rats (Erspamer, 1949). The structure of this substance, first called moschatin and then eledoisin, was established in 1962 (Anastasi and Erspamer, 1962; Erspamer and Falconieri Erspamer, 1962). In the same year, it was found that extracts of the skin of the South American leptodactylid frog Physalaemus biligonigerus (formerly fuscumaculatus) also displayed eledoisin-like activity. Also the elucidation of the structure of physalaemin (Anastasi et al., 1964; Erspamer et al., 1964) recalled that of eledoisin and similarly was followed in rapid succession by the identification of a number of other related peptides in the skin, in the brain and gut of amphibians, and in brain and gut of submammalian species (from birds to agnata).

² Abbreviations: SP, substance P; NKA, neurokinin A; NKB, neurokinin B; RIA, radioimmunoassay; SP-LI, substance P-like immunoreactivity; HPLC, high-performance liquid chromatography; 5-HT, 5-hydroxytryptamine; TK, tachykinin; TAN, tonically autoactive, giant neuron.

These peptides, all called tachykinins, represent the largest known peptide family, including members occurring in different animal species from low invertebrates to mammals. The possible occurrence of authentic tachykinins in invertebrates was confirmed by the isolation of two tachykinins from the salivary glands of a mosquito (Champagne and Ribeiro, 1994) and by the occurrence in nervous structures of the insect *Locusta migratoria* of four related peptides (the locustatachykinins) having structure homology with the vertebrate tachykinins (Schoofs et al., 1990a,b).

The identification of the locustatachykinins was soon followed by the isolation of similar peptides in other insects and in crabs, echinoid worms, and molluscs. It will be shown that locustatachykinin-like peptides, the number of which is destined to grow, have full citizenship right in the tachykinin peptide family, which with more than 40 members represents one of the largest, if not the largest, family in the peptide world.

The purpose of this review is, first, to keep the reader up to date on the different occurrences, species distributions, and localizations of the numerous members of the tachykinin peptide family. Second, because the identification and study of nonmammalian tachykinins has contributed conspicuously to the explosive progress of knowledge in the field of mammalian active tachykinins, we put in evidence the extensive pharmacological studies on the nonmammalian tachykinins (TKs) (eledoisin, physalaemin, and kassinin) whose availability preceded by years that of the corresponding mammalian peptides.

II. Occurrence and Species Distribution of Tachykinin-Like Peptides

At present among the numerous families of neuropeptides, which are evolutionarily the oldest neurotransmitters, perhaps even older than acetylcholine and catecholamines, four tachykinin-like peptides seem to occupy a very important position.

A. Invertebrate Tachykinin-Like Peptides

It is possible separate the tachykinin-like peptides in invertebrates into three groups: a) tachykinins identified by RIA and/or immunohistochemistry, occasionally accompanied by HPLC separation, but not isolated or sequenced; b) tachykinin-related peptides of the locustatachykinin type, isolated and sequenced, which have a C-terminal Arg-NH₂ residue instead of the usual Met-NH₂ residue present in all of the classical vertebrate tachykinins; and c) authentic tachykinins having structure and biological activity identical with those of the vertebrate tachykinins.

a. Substance P-like immunoreactivity (SP-LI) was localized in the primitive nervous system of *Hydra* (Taban and Cathieni, 1979; Grimmelikhuijzen et al., 1981; Pierobon et al., 1989), in the cerebral ganglion of the locust (Benedeczky et al., 1982), in the central nervous

system of the cockroach Periplaneta americana (Verhaert and De Loof, 1985), in the retina and eyestalk neurones of the lobster Palinurus interruptus (Mancillas et al., 1981), in the eye stalk of the fiddler crab Uca pugilator (Fingerman et al., 1985), in the somatogastric system of the crab Cancer borealis and the lobster P. interruptus and Homarus americanus (Goldberg et al., 1988), in tissues of the earthworm Lumbricus terrestris (Aros et al., 1980; Kaloustian and Edmands, 1986), in the adult nervous system of the fly Sarcophaga bullata (Sivasubramanian, 1990), in the cricket Teleogryllus communis (Lembeck et al., 1985), in the brain and central ganglia of the bowfly Calliphora vomitoria and of Drosophila (Lundquist et al., 1994), in the brain, corpora cardiaca, and corpora allata of the insect Leucophaea madeirae (El-Salhy et al., 1983), in the central nervous system of the mollusc Limulus polyphemus (Mancillas and Selverstone, 1985), and in the nervous system of several parasitic trematode worms (Bush and Gupta. 1988) including Schistosoma mansoni (Gustafsson, 1987), Diphyllobathrium dendriticum (Gustafsson et al., 1986). Fasciola hepatica (Magee et al., 1989), and Diclidophora merlangi (Maule et al., 1989).

In a large number of invertebrate phyla from coelenterates to molluscs, in addition to the usual SP-like tachykinin, an NKA-like peptide has also been found. However, it is highly improbable that authentic SP or authentic NKA is present in invertebrates. First, because retention time of the invertebrate tachykinins in elution from reverse-phase HPLC columns never coincided with retention time of the mammalian tachykinins; and second, because authentic SP and/or NKA was never found, even in lower submammalian species (amphibian, fish, and agnata). It has also been shown that radioimmunoassay and immunohistochemical techniques are often insufficient to distinguish between structurally related peptides because of frequent lack of selectivity of the pertinent antisera. Callitachykinin II, for example, was recognized not only by an antiserum to the locustatachykinin (in both peptides the C-terminal residue is Arg-NH₂) but also by an antiserum to the amphibian kassinin having, like all other classical tachykinins, the C-terminal residue Met-NH₂ (Lundquist et al., 1994). Moreover, preincubation of locustatachykinin antibody with SP and preincubation of SP antibody with locustatachykinin blocked subsequent immunolabeling of the somatogastric nervous system in C. borealis, indicating that a member of the locustatachykinins is likely to be the source of the previously identified SP in the nervous system (Blitz et al., 1995).

b. Schoofs et al. (1990a,b) first described the occurrence in insects, more precisely in extracts of brain, corpora cardiaca-corpora allata, and suboesophageal ganglion of *Locusta migratoria* of five peptides, the locustatachykinins, which exhibited sequence homologies (up to 45%) with the vertebrate tachykinins, especially with amphibian and fish

tachykinins. The locustatachykinins were completely inactive in all bioassay preparations used for the vertebrate tachykinins but showed a myotropic action in the insect intestine, eliciting a potent contraction of the cockroach hindgut (Winther et al., 1998).

The prediction of Schoofs' group in their first paper (Scoofs et al., 1990b) that "the peptides discovered in this study may be just the first in a whole series of substances from arthropod species to be identified as tachykinin family peptides" was correct even beyond any expectation. Up to the present, as many as 20 locustatachykinin-like peptides were isolated not only from various other arthropods, but also from an echinoid worm and from molluscs (Nassel, 1999). Table 1, reporting the present, probably provisional situation, shows that invertebrate tachykinin-like peptides are linear peptides with 8 to 15 amino acid residues and that, with the exception of the Leucophaea tachykininrelated peptide LemTRP10, they have at their C terminus an amidated Arg residue instead of the amidated Met residue, which is peculiar without exception to all classical tachykinins, including the invertebrate tachykinins (eledoisin and sialokinin I and II). Lem TRP1 is present also in two elongated forms with 17 (LemTRP2) and (LemTRP3) amino acid residues, respectively (Winther et al., 1999).

In the light of appearance on the screen of the locustatachykinins and of the fact that locustatachykinin antisera may cross-react with SP, it is probable that in several and perhaps in most cases, the SP-LI described

in a variety of invertebrates must be ascribed to locustatachykinin-like peptides. Thus, the locustatachykinin-like peptides of invertebrates must be considered authentic tachykinins, being either the primitive representatives of the tachykinin peptide family from which the vertebrate tachykinins have evolved by simple substitution of the C-terminal residue Arg-NH $_2$ with Met-NH $_2$ or an evolutionary adaptation for the invertebrates of a common ancestral tachykinin prototype already possessing the C-terminal Met-NH $_2$ residue.

c. Authentic tachykinins with the classical C-terminal pentapeptide sequence Phe-(Tyr/Ile)-Gly-Leu-Met-NH₂ occur in non-neuronal, epithelial cells of the posterior salivary glands of the Mediterranean octopods *E. moschata* and *Eledone aldovrandi* (eledoisin, up to 100 nmol/g wet tissue) (Erspamer and Falconieri Erspamer, 1962) and in the salivary glands of the mosquito *Aedes aegypti* (sialokinins I and II) (Champagne and Ribeiro, 1994): eledoisin, pGlu-Pro-Ser-Lys-Asp-Ala-Phe-Ile-Gly-Leu-Met-NH₂; sialokinin I, Asn-Thr-Gly-Asp-Lys-Phe-Tyr-Gly-Leu-Met-NH₂; sialokinin II, Asp-Thr-Gly-Asp-Lys-Phe-Tyr-Gly-Leu-Met-NH₂.

These peptides display the full spectrum of activity of the mammalian tachykinins and bind to the same receptors. It is remarkable that eledoisin occurs only in *Eledone* but not in the strictly related *Octopus vulgaris*. Yet, the salivary glands of both *Eledone* and *Octopus* contain large amounts of biogenic amines: serotonin (up to 2 μ mol/g), octopamine, tyramine, and histamine.

TABLE 1
Amino acid sequence of invertebrate tachykinin-related peptides

| Source/Peptide | Primary Structure | Reference |
|----------------------|---|-------------------------|
| Locusta migratoria | | Schoofs et al., 1990a,b |
| LomTK I | Gly-Pro-Ser-Gly-Phe-Tyr-Gly-Val-Arg-NH2 | |
| LomTK II | Ala-Pro-Leu-Ser-Gly-Phe-Tyr-Gly-Val-Arg-NH2 | |
| LomTK III | Ala-Pro-Gln-Ala-Gly-Phe-Tyr-Gly-Val-Arg-NH2 | |
| LomTK IV | Ala-Pro-Ser-Leu-Gly-Phe-Tyr-Gly-Val-Arg-NH2 | |
| Culex salinarius | | Clottens et al., 1993 |
| CusTK II | Ala-Pro-Ser-Gly-Phe-Met-Gly-Met-Arg-NH2 | |
| CusTK III | Ala-Pro-Tyr-Gly-Phe-Thr-Gly-Met-Arg-NH ₂ | |
| Anodonta cygnea | | Fujisawa et al., 1993 |
| AncTK | pGlu-Tyr-Gly-Phe-His-Ala-Val-Arg-NH2 | - |
| Urechis unicintus | | Ikeda et al., 1993 |
| UruTKI | Leu-Glu-Gln-Ser-Gln-Phe-Val-Gly-Ser-Arg-NH2 | , |
| UruTKII | Ala-Ala-Gly-Met-Gly-Phe-Phe-Gly-Ala-Arg-NH2 | |
| Calliphora vomitoria | | Lundquist et al., 1994 |
| CavTKI | Ala-Pro-Thr-Ala-Phe-Tyr-Gly-Val-Arg-NH2 | • |
| CavTKII | Gly-Leu-Gly-Asn-Asn-Ala-Phe-Val-Gly-Val-Arg-NH2 | |
| Leucophea maderae | | Muren and Nassel, 1996 |
| LemTRP1 | Ala-Pro-Ser-Gly-Phe-Leu-Gly-Val-Arg-NH2 | • |
| LemTRP4 | Ala-Pro-Ser-Gly-Phe-Met-Gly-Met-Arg-NH2 | |
| LemTRP5 | Ala-Pro-Ala-Met-Gly-Phe-Gln-Gly-Val-Arg-NH2 | |
| LemTRP6 | Ala-Pro-Ala-Ala-Gly-Phe-Phe-Gly-Met-Arg-NH2 | |
| LemTRP7 | Val-Pro-Ala-Ser-Gly-Phe-Phe-Gly-Met-Arg-NH2 | |
| LemTRP8 | Gly-Pro-Ser-Met-Gly-Phe-His-Gly-Met-Arg-NH2 | |
| LemTRP9 | Ala-Pro-Ser-Met-Gly-Phe-Gln-Gly-Met-Arg-NH2 | |
| Cancer borealis | | Christie et al., 1997 |
| CabTRP1a | Ala-Pro-Ser-Gly-Phe-Leu-Gly-Met-Arg-NH2 | , |
| CabTRP1b | Ser-Gly-Phe-Leu-Gly-Met-Arg-NH2 | |
| Penaeus vannamei | | Nieto et al., 1998 |
| PevTRP | Ala-Pro-Ser-Gly-Phe-Leu-Gly-Met-Arg-NH2 | , |

L. migratoria (arthropod): brain, corpora cardiaca-corpora allata, subesophageal ganglion; C. salinarus (artropod): nervous tissue; A. cygnea (bivalve mollusc): central nervous system; Urechis uncinatus (echinoid worm): nervous tissues; C. vomitoria (artropod): nervous tissues; Leucophea maderae (artropod): brain and midgut; Cancer borealis (artropod): stomatogastric nervous system; Peneaeus vennamei (artropod): central nervous system.

B. Prevertebrate Tachykinin-Like Peptides

- 1. Amphioxus lanceolatus. Radioimmunoassay combined with HPLC suggests the occurrence of small amount of SP-LI in brain and spinal cord of this prevertebrate species (Lembeck et al., 1985).
- 2. Tunicata (Protocordata). Using immunohistochemical techniques only, the presence of an antigen related to substance P has been demonstrated in the neuronal ganglion (Fritsch et al., 1979), gill epithelium (Fritsch et al., 1980), and alimentary tract (Fritsch et al., 1982) of the ascidian Ciona intestinalis. More recently, the occurrence of tachykinins in C. intestinalis tissues was re-examined by O'Neil et al. (1987) using specific antisera for the C terminus (C) of SP and the N terminus (N) of mammalian SP and NKA, completed with immunohistochemistry and reverse-phase HPLC of the tissue extracts. It was found that only C-SP-LI (not N-SP-LI) occurs both in cells of the ganglia and in peripheral neurons, together with but separately from N-NKA-LI. Only C-SP-LI was found in endocrine cells of the pharynx. However, we conclude that already at the prevertebrate stage of chordate evolution, the tachykinin family is represented by at least two distinct members that are provided by separate cell populations, none of which was identical with either mammalian SP or mammalian NKA.

C. Submammalian Vertebrate Tachykinins

The formidable enlargement of the tachykinin peptide family is consequent to systematic studies conducted on the one side on the amphibian skin and on the other side on the brain and intestines of submammalian vertebrates, mainly in their cold-blooded classes: reptiles, amphibians, fish, and agnata. The story of the group of the amphibian skin peptides began in 1964 with isolation and structure elucidation of physalaemin (Erspamer et al., 1964), followed by the systematic screening of peptide contents in the skin of as many as 600 amphibian species from all over the world, which resulted in the discovery and isolation of numerous neuropeptides, belonging to a dozen distinct families, among which is that of the tachykinins (with 21 members).

The fruitful search for tachykinin peptides in brain and gut of submammalian vertebrates started in 1986 with the isolation and structure elucidation of scyliorhinins I and II from dogfish intestine (Conlon et al., 1986a). At the end of 1998, a list of as many as 24 novel tachykinins was available, 12 from brain and 12 from gut. Skin tachykinins and brain/gut tachykinins will be discussed separately.

1. Amphibian Skin Tachykinins. Table 2 summarizes the present situation. The great majority of amphibian skin peptides have the classical C-terminal pentapeptide sequence: Phe-X-Gly-Leu-Met-NH₂. However, important exceptions are represented by: 1) some tachykinins from the skin of the Australian frog Agalychnis callidryas, namely AC-AR1, -AR2, and -AR3 with the C-terminal pentapeptide sequence Phe-Tyr-Pro-Gly-Met-NH₂ and AC-AR4 with sequence Phe-Tyr-Pro-Val-Met-NH₂; and 2)

TABLE 2
Amino acid sequence of natural amphibian skin tachykinins

| | Amino acia sequence of natural amphibian skin tachykinins | |
|--|--|-------------------------|
| Source/Peptide | Primary Structure | Reference |
| Physalaemus biligonigerus | | Anastasi et al., 1964; |
| (fuscumaculatus) | | Erspamer et al., 1964 |
| Physalaemin (PHYS) | ${	t pGlu-Ala-Asp-Pro-Asn-Lys-Phe-Tyr-Gly-Leu-Met-NH}_2$ | |
| Uperuleia rugosa | | Nakajima et al., 1980 |
| $[Lys^5, Thr^6]PHYS$ | ${	t pGlu-Ala-Asp-Pro-Lys-Thr-Phe-Tyr-Gly-Leu-Met-NH}_2$ | |
| Uperuleia marmorata | | Anastasi et al., 1975 |
| Uperolein | pGlu-Pro-Asp-Pro-Asn-Ala-Phe-Tyr-Gly-Leu-Met-NH $_2$ | |
| Uperuleia inundata | | Bradford et al., 1996 |
| Uperin | $	t pGlu-Ala-Asp-Pro-Asn-Ala-Phe-Tyr-Gly-Leu-Met-NH_2$ | |
| Kassina (Hylambates) | | Yasuhara et al., 1981 |
| maculata | | |
| Hylambatin | ${\tt Asp-Pro-Asp-Pro-Asn-Arg-Phe-Tyr-Gly-Leu-Met-NH}_2$ | |
| [Glu ² , Pro ⁵]Kassinin | ${\tt Asp-Glu-Pro-Lys-Pro-Asp-Gln-Phe-Val-Gly-Leu-Met-NH}_2$ | |
| Kassina senegalensis | | Anastasi et al., 1976 |
| Kassinin | ${\tt Asp-Val-Pro-Lys-Ser-Asp-Gln-Phe-Val-Gly-Leu-Met-NH}_2$ | |
| Rana margaratae | | Lu et al., 1990 |
| Ranamargarin | ${\tt Asp-Asp-Ala-Ser-Asp-Arg-Ala-Lys-Lys-Phe-Tyr-Gly-Leu-Met-NH}_2$ | |
| Phyllomedusa bicolor | | Anastasi and Falconieri |
| Phyllomedusin | pGlu-Pro-Asn-Pro-Asn-Arg-Phe-Ile-Gly-Leu-Met-NH2 | Erspamer, 1970 |
| Pseudophryne guntheri | | Simmaco et al., 1990 |
| PG-SP1 | pGlu-Pro-Asn-Pro-Asp-Glu-Phe-Tyr-Gly-Leu-Met-NH $_{ m 2}$ | |
| PG-SP2 | pGlu-Pro-Asn-Pro-Asn-Glu-Phe-Tyr-Gly-Leu-Met-NH2 | |
| PG-KI | pGlu-Pro-His-Pro-Asp-Glu-Phe-Val-Gly-Leu-Met-NH2 | |
| PG-KII | pGlu-Pro-Asn-Pro-Asp-Glu-Phe-Val-Gly-Leu-Met-NH2 | |
| PG-KIII | pGlu-Pro-His-Pro-Asn-Glu-Phe-Val-Gly-Leu-Met-NH2 | |
| Agalychnis callidryas | | Mignogna et al., 1997 |
| AC-AL | ${	t Gly-Pro-Pro-Asp-Pro-Asn-Lys-Phe-Ile-Gly-Leu-Met-NH}_2$ | |
| AC-AR1 | Gly-Pro-Pro-Asp-Pro-Asp-Arg-Phe-Tyr-Pro-Gly-Met-NH2 | |
| AC-AR2 | Gly-Pro-Pro-Asp-Pro-Asp-Lys-Phe-Tyr-Pro-Gly-Met-NH2 | |
| AC-AR3 | pGlu-Pro-Asp-Pro-Asp-Lys-Phe-Tyr-Pro-Glyl-Met-NH2 | |
| AC-AR4 | ${	t Gly-Pro-Pro-Asp-Pro-Asn-Lys-Phe-Tyr-Pro-Val-Met-NH}_2^-$ | |

hylambatin from the skin of the South-African frog $Hylambates\ maculatus$ with the C-terminal pentapeptide sequence Phe-Tyr-Gly-Met-Met-NH $_2$. It is evident that in the C-terminal pentapeptide only the Phe residue at position 5 from the C terminus and Met-NH $_2$ are immutable.

All of the amphibian skin peptides have a non-neuronal origin, being synthesized in the syncytial cells dressing the wall of the granular glands. These cells are capable of cosynthesizing, costoring, and cosecreting not only peptides belonging to different families, but also amines and alkaloids belonging to various classes and families. The amphibian syncytial cells behave like some mammalian endocrine cells, e.g., the enterochromaffin cells, which may contain both biogenic amines and peptides (substance P, guanylin), and like a number of central and peripheral neurons in which amine messengers coexist with peptide messengers.

2. Brain and Gut Tachykinins. Table 3 summarizes the present situation. All of the tabulated tachykinins, with the exception of ranatachynin D, show the classical C-terminal pentapeptide Phe-X-Gly-Leu-Met-NH₂. Of considerable interest is the fact that in goldfish, cod, and trout NKA-like peptides, the usual acidic Asp residue at position 7 from the C terminus, crucial for receptor NK2/NK3 selectivity, is replaced by the neutral Asn residue.

The list of tachykinins shown in the Table 3 should be completed by authentic NKA occurring in the intestine of the chicken and of the alligator and in the brain of the python, and by authentic NKB found only in the brain of *Rana esculenta*.

Moreover, NKA is present in as many as six submammalian species also by its elongated form, the γ -neuropeptides, as shown in Table 4. From the above sequences, it is evident that none of the submammalian γ -neuropeptides is identical with the corresponding mammalian peptide. Substantial differences in the amino acid composition may be seen not only in the flanking sequence but in all examined fish, even in the NKA-like C-terminal decapeptides.

D. Mammalian Tachykinins

1. Mammalian Tachykinins and Their Biosynthesis. Until now, only three tachykinins have been isolated and sequenced from mammalian tissues: SP, NKA (neuromedin L, neurokinin, and substance k), and NKB (neurokinin and neuromedin k). NKA is present also in two elongated forms, neuropeptide K and neuropeptide- γ (Table 5).

It is hardly conceivable, but of course it is possible, that mammalian tissues contain only three members of the tachykinin family. As a matter of fact, the number of mammalian species in which tachykinin peptides have been isolated is very scanty: horse and guinea pig intestine, porcine spinal cord, and in some additional species (rat and man) preprotachykinins have been detected. RIA or immunohistochemistry, again in a limited number of species and especially in the rat, has detected all other tachykinin locations. This is all. Yet fish present five different tachy-

kinins in the brain and six in the gut, and the four examined amphibian species exhibit as many as nine different tachykinins altogether in the brain and the gut. It is evident that the occurrence even in mammalian tissues of tachykinins other than the three classical ones is likely. Lazarus group (Lazarus and Di Augustine, 1980; Lazarus et al., 1980), using an antiserum specifically recognizing the N-terminal region of physalaemin, was able to detect a physalaemin-LI in a number of tissues of three mammalian species (guinea pig, mouse, and rat) with peaks in guinea pig and mouse gastric fundus, pylorus, and duodenum (up to 18 pmol/g lyophilized tissue). Moreover, physalaemin antiserum caused a clear-cut immunostaining in a population of cells of the Brunner's gland of the guinea pig duodenum, and several other examples of the expression of physalaemin-, eledoisin-, and kassinin-LI may be found in carcinoids (see Section III.A.4.).

Mammalian tachykinins are derived from two preprotachykinin genes: the PPT-A gene, which encodes the sequences of SP, NKA, and neuropeptide K and neuropeptide- γ , and the PPT-B gene, which encodes the sequence of NKB (Nawa et al., 1983; Kotani et al., 1986; Bonner et al., 1987; Krause et al., 1987).

The precursor RNA from PPT-A is alternatively processed to yield three different mRNAs (Nawa et al., 1984). The three precursor proteins from which the mRNA codes are designated α -, β -, and γ -PPT; α -PPT, which generates SP; β -PPT, which generates SP, NKA, and neuropeptide K; and γ -PPT, which generates SP, NKA, and neuropeptide- γ . The biological significance of the alternative splicing of PPT-A is unknown. The relative proportion of α -, β -, and γ -PPT mRNAs is markedly species dependent. For example, β -PPT is the predominant form expressed in human basal ganglia (Bannon et al., 1992), whereas α -PPT prevails in the bovine brain (Nawa et al., 1984).

 α -PPT mRNA is abundant in the brain, whereas β -and γ -PPT mRNAs are found mainly in peripheral tissues (Nakanishi, 1987). PPT-B mRNA is found in the brain (hypothalamus) and intestine (Kotani et al., 1986). Tachykinins are liberated from their precursors by the action of specific processing proteases. Typical cleavage points are Lys-Arg, Arg-Arg, and Arg-Lys doublets and the cleavage is carried out by six groups of proteolytic enzymes called convertases (Chretien et al., 1989; Steiner et al., 1992; Marcinkiewicz et al., 1993). COOHterminal amidation after cleavage is generated from the precursor sequence, Gly-Leu-Met-Gly-Lys-Arg, in which Gly acts as the amide donor.

As with all known neurotransmitters, neuronal tachy-kinins are also released from the nerve ending after a calcium-dependent mechanism in response to application of physiological and nonphysiological stimuli (electrical stimulation, potassium, or capsaicin depolarization) (Maggi et al., 1993). Concerning release, two points are firmly established.

First, like that of biogenic amines, which are considered "rapid transmitters" and which under certain con-

TABLE 3
Amino acid sequence of submammalian vertebrate tachykinins

| | | capacitions and icon in aminutation to accomple man country | |
|--|----------------|--|--|
| Source/Peptide | | Primary Structure | Reference |
| Gallus domesticus | | | |
| Alligator mississinionsis | Gut | ${ m Arg-Pro-Arg-Pro-Gln-Gln-Phe-Gly-Leu-Met-NH}_2$ | Conlon et al., 1988 |
| Tring and treascast tong | Brain | Arg-Pro-Arg-Pro-Gln-Gln-Phe-Phe-Gly-Leu-Met-NH ₂ | Wang et al., 1992b |
| $Python\ molurus$ | - | | |
| Dana actochaiana | Gut | ${ m Arg-Pro-Arg-Pro-Gln-Gln-Phe-Tyr-Gly-Leu-Met-NH}_2$ | Conlon et al., 1997 |
| Ranatachykinis | A) Brain, gut | Lys-Pro-Ser-Pro-Asp-Arg-Phe-Tyr-Gly-Leu-Met-NH, | Kozawa et al., 1991 |
| • | B) Brain | Tyr-Lys-Ser-Asp-Ser-Phe-Tyr-Gly-Leu-Met-NH2 | |
| | C) Gut | His-Asn-Pro-Ala-Ser-Phe-Ile-Gly-Leu-Met-NH ₂ | |
| t | D) Gut | Lys-Pro-Asn-Pro-Glu-Arg-Phe-Tyr-Ala-Pro-Met-NH2 | 1000 |
| <i>Kana riaibunaa</i> Kanakinin | Gut | LYS-Pro-Asn-Pro-GALL-Asg-Pro-Lyr-GLY-Deu-Wec-NH2 Line Tive Ton And Con-line Tive Ton Mat Mul | U Harte et al., 1991 W_{cong} et al., 1992 |
| $Bufo\ marinus\ Bufokinin$ | Gut | has byshed asplot fire its off bed mee ming bysher byshediy bed mee ming bysher asplot byshediy-leu-Met-NH2 | Conlon et al., 1998 |
| Ampnuma triaactytum | Gut | Asp-Asn-Pro-Ser-Val-Gly-Gln-Phe-Tyr-Gly-Leu-Met-NH2 | Waugh et al., 1995a |
| Oncorhynohue mybies | cat | HIS-LYS-ASP-ALA-PRE-LIE-GLY-LEU-MET-NH $_2$ | |
| ocarifational attacks | Brain Gut | Lys-Pro-Arg-Pro-His-Gln-Phe-Phe-Gly-Leu-Met-NH, His-Arg-Ile-Asn-Ser-Phe-Val-Gly-Leu-Met-NH, | Jensen et al., 1992 Jensen et al., 1993 |
| Gadus morhua | G.: | א ביין אירי אירי אירי אירי אירי איר מאר מאר מייד בייד בייד בייד בייד בייד בייד בייד | Tongon of al. 1009 |
| Carassius auratus | brain | Lys-Fro-Arg-Fro-Gin-Gin-Fre-Lie-Gi $\sqrt{-}$ Leu-Met-NH $_2$ | Jensen et al., 1992 |
| Carassin (13–21) | Brain | ${	t His-Arg-Ile-Asn-Ser-Phe-Val-Gly-Leu-Met-NH}_2$ | Conlon et al., 1991; Lin and Peter 1997 |
| A | | ${\tt Lys-Pro-Arg-Pro-His-Gln-Phe-Ile-Gly-Leu-Met-NH}_2$ | 1001, 1001 |
| Amu caloa | Stomach | ${\tt Ser-Lys-Ser-His-Gln-Phe-Tyr-Gly-Leu-Met-NH}_2$ | Waugh et al., 1995b |
| Scupnirnyncus platornyncus | Gut | ${\tt Ser-Lys-Thr-His-Gln-Phe-Tyr-Gly-Leu-Met-NH}_2$ | Wang et al., 1999 |
| Scyttorninus canicula Scyliorhinin I Scyliorhinin I | Gut | Ala-Lys-Phe-Asp-Lys-Phe-Tyr-Gly-Leu-Met-NH2 | Conlon et al., 1986a |
| | Brain | oer-rro-oer-asm-ser-bys-cys-rro-asp-ory-rro-asp-cys-rme-var-ery-bra-mer-nm ₂ Lys-pro-Arg-pro-Gly-Gln-phe-phe-Gly-Leu-Met-NH ₂ | Waugh et al., 1993 |
| $Iorpedo\ marmorata \ Des(Ser^1, Pro^2) \ Scyliorhinin II$ | Gut | ${\tt Ser-Asn-Ser-Lys-Cys-Pro-Asp-Gly-Pro-Asp-Cys-Phe-Val-Gly-Leu-Met-NH}_2$ | Conlon and Thim, 1988 |
| Sphyrna lewinini | Gut | ${\tt Ala-Lys-Phe-Asp-Lys-Phe-Tyr-Gly-Leu-Met-NH}_2$ | Waugh et al., 1995a |
| naja runa | Brain Brain | Ala-Lys-His-Asp-Lys-Phe-Tyr-Gly-Leu-Met-NH ₂ His-Lys-Leu-Gly-Ser-Phe-Val-Gly-Leu-Met-NH ₂ | Waugh et al., 1994 |
| Lampetra fluviatilis | Brain Brain | Arg-Lys-Pro-His-Pro-Lys-Glu-Phe-Val-Gly-Leu-Met-NH2 His-Phe-Asp-Glu-Phe-Val-Gly-Leu-Met-NH2 | Waugh et al., 1995a |
| Petromyzon marınus | Brain | Arg-Lys-Pro-His-Pro-Lys-Glu-Phe-Val-Gly-Leu-Met-NH ₂ | Waugh et al., 1994 |
| | | | |

292

TABLE 4 Amino acid sequence of submammalian vertebrate γ -neuropeptides

| Source | Primary Structure | Reference |
|-------------|--|---|
| Mammals | | |
| Spinal cord | DAGHGQISHKR His-Lys-Thr-Asp-Ser-Phe-Val-Gly-Leu-Met-NH; | Kangawa et al., 1983; Kimura et al., 1983 |
| Gold fish | | |
| Brain | SPANAQITRKR His-Lys-Ile-Asn-Ser-Phe-Val-Gly-Leu-Met-NH; | Conlon et al., 1991 |
| Trout | | |
| Gut | SSANPQITRKR His-Lys-Ile-Asn-Ser-Phe-Val-Gly-Leu-Met-NH, | Jensen et al., 1993 |
| Phyton | | |
| Gut | DAGYSPLSHKR His-Lys-Thr-Asp-Ser-Phe-Val-Gly-Leu-Met-NH, | Conlon et al., 1997 |
| Alligator | | |
| Brain | DAGYGQISHKR His-Lys-Thr-Asp-Ser-Phe-Val-Gly-Leu-Met-NH, | Wang et al., 1992b |
| Shark | | |
| Gut | ASGPTQAGIVGRKR Gln-Lys-Gly-Glu-Met-Phe-Val-Gly-Leu-Met-NH; | Waugh et al., 1995a |
| Bowfin | | |
| Stomach | SGAPQTVPLGRKR His-Lys-Gly-Glu-Met-Phe-Val-Gly-Leu-Met-NH, | Waugh et al., 1995b |

TABLE 5
Amino acid sequence of mammalian tachykinins

| Sp Reference SP Arg-Pro-Lys-Pro-Gln-Gln-Phe-Phe-Gly-Leu-Met-NH2 Chang et al., 1971 Bovine hypothalamus Arg-Pro-Lys-Pro-Gln-Gln-Phe-Phe-Gly-Leu-Met-NH2 Chang et al., 1971 NKA Porcine spinal cord Asp-Met-His-Lys-Asp-Phe-Phe-Val-Gly-Leu-Met-NH2 Kimura et al., 198 NKB Asp-Ala-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 Kage et al., 198 Neuropeptide-y Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 Tatemoto et al., 15 Porcine brain Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 Tatemoto et al., 15 | | Amino acid sequence of mammalian tachykinins | |
|--|------------------------|---|-----------------------|
| Arg-Pro-Lys-Pro-Gln-Gln-Phe-Phe-Gly-Leu-Met-NH ₂ His-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH ₂ cord | Peptide/Source | Primary Structure | Reference |
| cord His-Lys-Arg-Phe-Gln-Phe-Gly-Leu-Met-NH2 cord His-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 cord Asp-Ala-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Phe-Phe-Val-Gly-Leu-Met-NH2 asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-His-Gly-Leu-Met-NH2 asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 asp-Ala-Asp-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 asp-Ala-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 asp-Ala-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 asp-Ala-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 asp-Ala-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 asp-Ala-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 asp-Ala-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 asp-Ala-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg | SP | | |
| cord His-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 Asp-Ala-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Axg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Axg-Lys-Axg-Lys-Axg-Lys-Axg-Lys-Axg-His-Lys-Axg-His-Lys-Axg-His-Lys-Axg-L | Bovine hypothalamus | Arg-Pro-Lys-Pro-Gln-Gln-Phe-Gly-Leu-Met-NH ₂ | Chang et al., 1971 |
| cord His-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH ₂ Asp-Met-His-Asp-Phe-Phe-Val-Gly-Leu-Met-NH ₂ Asp-Ala-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH ₂ Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Arg-His-Gly-Leu-Met-NH ₂ | NKA | | |
| l cord Asp-Met-His-Asp-Phe-Phe-Val-Gly-Leu-Met-NH2 Asp-Ala-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Leu-Met-NH2 | Porcine spinal cord | His-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 | Kimura et al., 1983 |
| l cord Asp-Ala-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH ₂ Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lis-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lis-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lis-Lys-Arg-Lis-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lis-Lys-Lys-Lys-Lys-Lys-Lys-Lys-Lys-Lys-Ly | NKB | | |
| ne Asp-Ala-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln-Ile-Ser-His-Gly-Gln-Ile-Ser-His-Gly-Gln-Ile-Ser-His-Gly-Gln-Ile-Ser-His-Gly-Gln-Ile-Ser-His-Gly-Gln-Ile-Ser-His-Gly-Gln-Ile-Ser-His-Gly-Leu-Met-NH2 | Porcine spinal cord | Asp-Met-His-Asp-Phe-Val-Gly-Leu-Met-NH ₂ | Kangawa et al., 1983 |
| Asp-Ala-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH ₂ Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-His-Gly-Leu-Met-NH ₂ | Neuropeptide- γ | | |
| Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln- Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH ₂ | Rabbit intestine | Asp-Ala-Gly-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH ₂ | Kage et al., 1988 |
| Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln- Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 | Neuropeptide K | | |
| Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 | Porcine brain | Asp-Ala-Asp-Ser-Ser-Ile-Glu-Lys-Gln-Val-Ala-Leu-Leu-Lys-Ala-Leu-Tyr-Gly-His-Gly-Gln- | Tatemoto et al., 1985 |
| | | Ile-Ser-His-Lys-Arg-His-Gly-Gln-Ile-Ser-His-Lys-Arg-Lys-Asp-Ser-Val-Phe-Gly-Leu-Met-NH2 | |

ditions may be released massively, release of neuropeptides, considered "slow" transmitters or modulators, is probably discrete and long lasting. Second, at the nerve terminals, especially in brain and in the autonomic nervous system, a release of a single transmitter is improbable, and, at any rate, must represent an exception. The concept of co-release of different peptides, amines, amino acids, and purines is now generally accepted after the immunohistochemical demonstration of the costorage in the granular material of single neurones of more active substances (Hokfelt et al., 1986).

Once released, the tachykinins may be attacked, cleaved, and inactivated by a number of proteolytic enzymes, which, however, act with considerably different intensity on the different tachykinins. The most vulnerable peptide seems to be SP, whereas peptides having at their N-terminal the pGlu residue seem much more resistant to enzyme attack. In the proteolytic degradation of SP, three enzymes seem to display a predominant role: dipeptidyl-amino peptidase, postproline endopeptidase, and cathepsin D (Regoli et al., 1994a).

III. Localization of Tachykinin-Like Peptides

We have previously shown that tachykinins constitute one of the largest families of peptides in the world whose members are present in all animal species, from lower invertebrates to mammals. Tachykinins possess a widespread distribution in the central and peripheral nervous system that is undoubtedly the major source of these peptides. However, tachykinins have also, like numerous other peptides and like all biogenic amines, a limited and species-dependent, but not negligible, distribution in nonneuronal structures represented by the irregular and sparse localizations in which they display known and unknown functions. In the first localization (neuronal cells), the active compounds act as neurotransmitters/neuromodulators, in the second (non neuronal cells) as autocrine, paracrine, or endocrine regulators.

A. Non-Neuronal Localization

1. Amphibian Skin. The most complex and rich localization of non-neuronal tachykinins is certainly the amphibian skin, that may be considered a huge factory and storehouse of a variety of tachykinins. However, an important characteristic of the skin peptides is their extreme irregularity in occurrence and richness: some amphibian species are extremely rich in active peptides, others, even closely related species, are lacking of active peptides. These findings contribute to the incomprehension of the physiological significance of the skin tachykinins and of that of the skin biogenic amines, especially indolealkylamines that are similarly present, sometimes in enormous amounts in the skin. The occurrence in the skin of a such variety of neuropeptides and amines may perhaps be explained by the common embryogenic origin

of the integument and the nervous system from the primitive ectoderm.

- 2. Invertebrate Salivary Glands. The significance of eledoisin, as with that of the large amounts of biogenic amines, occurring in the posterior salivary glands of *E. moschata* but not in the glands of any related Octopus species is again completely obscure. On the contrary, the sialokinins occurring in the salivary glands of *A. aegypti* may be interpreted as an evolutionary selection of the peptides as vasodilators, in connection with the habit of feeding in blood of this insect (Champagne and Ribeiro, 1994).
- 3. Normal Mammalian Tissues. All of the non-neuronal localizations of the tachykinins concern mainly SP, and the occurrence of the peptide was established only by immunohistochemistry, using selective SP-antisera. SP occurring (together with serotonin) in populations of argentaffin/chromaffin and argyrophil/acidophil (with no serotonin) cells of the mammalian and probably also of lower vertebrate (ascidian and fish) intestine may act either as paracrine hormone or as a true hormone after release into the blood stream. It is possible that SP found in mammalian blood originates prevalently from the SP-containing cells of the intestinal mucosa. More than 60 years ago. Vialli and Erspamer (1933) described the so-called "acidophil basigranular cells" of the dog and cat large intestine, and it would be worthwhile checking to see if they are, like the argyrophil cells of the human large intestine, SP-producing cells. The acidophil cells are, in fact, neither argentaffin nor chromaffin, that is they do not contain serotonin. Little is known as to whether and under which conditions SP is released from the endocrine cells of the gastrointestinal mucosa. For example, release of SP from the enterochromaffin cells of the rat caecum mucosa seems to be inhibited by serotonin and calcium-free medium (Simon et al., 1992). Moreover, that SP may be released from the gut endocrine cells into the blood stream is strongly suggested by the evidence that legation of all intestinal blood vessels and evisceration in the cat significantly lowered SP plasma levels (Gamse et al., 1978) and by the fact that portal venous blood contains about 4 times more SP than peripheral blood (Pernow, 1983).

At any rate, it is obvious that blood SP must display some function. The most acceptable suggestion is that the peptide acts on the blood vessels either indirectly through release of vasodilator agents from the endothelium (vasodilation) or directly on the vascular smooth muscle, causing generally constriction or even potent stimulation of phasic movements in particular vessels (e.g., rat portal vein). There are sharp species differences in the response of the vessels to tachykinins, depending not only on the vascular beds but also on animal species (see *Section V.I.*).

4. Endocrine Tachykinin-Secreting Tumors. Carcinoids. Carcinoids are tumors of the diffuse endocrine system characterized by a typical growth pattern, silver affinity, and positive immunohistochemical reaction with

specific markers. They can express different biogenic amines (serotonin and histamine), peptides (tachykinins, bradykinin, and enteroglucagon), and prostaglandins (Creutzfeldt, 1996). Among the amines, serotonin is always present in argentaffin/chromaffin carcinoids, originating from the malignant growth of the enterochromaffin cells. They are prevailingly present in the midgut and appendix. Serotonin, however, is lacking in argyrophil nonchromaffin carcinoids, which may be present in the foregut, hindgut, lung (mostly bronchial carcinoids) and various other organs (Creutzfeldt and Stockmann, 1987). Argyrophil carcinoids may originate in the colon and probably in other sites as well, from the population of argyrophil, serotoninlacking cells described by Sokolski and Lechago (1984). Peptides (bradykinin, enteroglucagon, and especially tachykinins) may occur both in chromaffin (together with serotonin) and in argyrophil nonchromaffin carcinoids (Wilander et al., 1977, 1979).

Because in normal endocrine cells of the gut and other organs only SP has been detected by immunochemistry, as expected, the tachykinin most frequently identified not only in the primary carcinoid tumor but also in its metastases was SP, together with its fragment SP(5–11), both in normal and oxidized form (Gamse et al., 1981; Conlon et al., 1985; Roth et al., 1985; Theodorsson-Norheim et al., 1985; Bishop et al., 1989). However, NKA, together with its fragments NKA(4-10) and NKA(5-10) and its extended form neuropeptide K, may also frequently be present in carcinoids (Roth et al., 1985; Theodorsson-Norheim et al., 1985; Conlon et al., 1986b; Bishop et al., 1989); and, more rarely, an eledoisin-like immunoreactivity (eledoisin-LI) was observed together with a neurokinin B-LI (Theodorsson-Norheim et al., 1985) and an oxidized physalaemin-LI (Conlon et al., 1985).

SP-LI was also found, together with NKA-LI, in bronchial carcinoids (Creutzfeldt and Stockmann, 1987; Bishop et al., 1989), in ovarian carcinoids (Skrabanek et al., 1980; Strodel et al., 1984), and in a medullary carcinoma of the thyroid (Skrabanek et al., 1979).

Tachykinins (NKA-, neuropeptide K-, and eledoisinlike peptides) were produced also by carcinoid tumors in culture (Norheim et al., 1987) and both 5-HT- and SP-LI were found in cytoplasmatic granules isolated from an intestinal argentaffin carcinoid, supporting the view that in this case SP is costored with 5-HT in the granules of the enterochromaffin cells (Alumets et al., 1977).

Pheocromocytomas. Two pheocromocytomas showed SP-LI and SP sulfoxide-LI (Gamse et al., 1981), and another pheocromocytoma showed NKB-LI (Conlon et al., 1985). After subcellular fractionation, SP-LI and catecholamines were enriched in the chromaffin granular fraction, making it unlikely that SP-LI originates from nerve terminals (Gamse et al., 1981).

Lung carcinoma. Only a few data on the content of tachykinins in these carcinoid tumors are available: up to 2 ng/g fresh tissue of SP-like peptides, 1.2 ng/g being represented by authentic SP (Gamse et al., 1981). This

content is surprisingly low, compared with the content of serotonin (from 1 μ g to 2.5 mg per g fresh tissue) in argentaffin carcinoids (Stacey, 1966). Moreover, Lazarus et al. (1983) have presented evidence that a human small cell carcinoma may contain a tachykinin peptide (1–1.6 ng/g) that has structural and biological activity similar to that of the amphibian physalaemin.

Because carcinoid tumors and their metastases are authentic endocrine glands, releasing into the blood stream biogenic amines and tachykinins, levels of these compounds may increase in plasma and even in urine. TK-LI was found in 75% of the 65 carcinoid patients examined (Norheim et al., 1984). The major component in plasma eluted in ion-exchange chromatography was in a different position from that of the usual tachykinins. Similar results were obtained by Conlon et al. (1986b) in three of four carcinoid patients, with NKA-LI up to 1 nmol/ml plasma and SP-LI up to 345 fmol/ml. Moreover, in urine samples of 79% of the 48 carcinoid patients examined in another study, the TK-LI material was 8 times more elevated than in healthy subjects. The immunoreactive material was heterogeneous, with some components coeluting with oxidized NKA and neuropeptide K (Bergstrom et al., 1995). In the urine of patients with argentaffin carcinoids, the concentration of 5-hydroxyindolacetic acid, the main metabolite of serotonin, was 100 times more elevated than in healthy subjects (Stacey, 1966).

All of these findings on carcinoid tumors demonstrate that tumoral epithelial, non-neuronal cells of the mammalian intestine and other organs are capable of expressing and storing not only SP, as expected, but several other tachykinins as well, certainly NKA and its extended form neuropeptide K, but also NKB and kassinin-, eledoisin-, and physalaemin-like peptides, i.e., peptides occurring in normal tissues only in submammalian species.

Patients with argentaffin carcinoid tumors and their metastases very often exhibit a typical syndrome characterized by flushing, diarrhea, asthma, cyanosis, and right-side valvular disease. Creutzfeldt and Stockmann (1987) have considered tachykinins to be coresponsible only for vasodilation (flushing) and not of the other symptoms. Secretory diarrhea and enhanced motility, important features of the carcinoid syndrome, do not seem to be attributable to SP, but instead to NKA and to eledoisin-like peptides (Makridis et al., 1999).

We can conclude that carcinoid tumors are not pure tachykinin-secreting tumors and do not contribute, unlike other endocrine tumors (gastrinomas and vipomas), to the understanding of the physiological significance and function of the tachykinins.

B. Neuronal Localization

Nervous tissue represents by far the most important localization of the tachykinins in invertebrates and vertebrates. A more or less dense network of tachykininergic fibers, which release their content upon adequate stimulation, permeates all vertebrate tissues close upon

a very rich population of different receptors located on the membrane of neuronal and non-neuronal cells. In certain cerebral areas, the concentration of tachykinins may be on the order of nanomoles.

Data on the distribution and localization of neuronal tachykinins in the CNS and periphery have been obtained by a combination of HPLC with radioimmunoassay and/or by immunohistochemistry (Hokfelt et al., 1975, 1977; Pernow, 1983; Maggio, 1985).

Regional specific antisera directed against the C-terminal region of the tachykinins have been generally used, a condition that is unfortunate for discrimination between the various tachykinins (Maggio, 1988). The low specificity of several antisera and the different tissue extraction methods (Lindefors et al., 1985; Brodin et al., 1986) may explain some differences encountered in the literature on the localization of the three mammalian tachykinins.

Central nervous system. Distribution of the tachykinins in the CNS has been extensively studied only in the rat (Otsuka and Yoshioka, 1993). Data on other species are scanty. As expected, SP is generally cosynthesized, colocalized, and cosecreted with NKA. The values of immunoreactive SP in various areas of the rat brain are: olfactory tubercle, 300 pmol/g of wet tissue; amygdala, 383 pmol/g of wet tissue; nucleus caudatus, 247 pmol/g of wet tissue; globus pallidus, 332 pmol/g of wet tissue; septum, 116 to 405 pmol/g of wet tissue; hypothalamus, 208 to 626 pmol/g of wet tissue; habenula, 377 pmol/g of wet tissue; posterior pituitary, 489 pmol/g of wet tissue; thalamic nucleus, 25 pmol/g of wet tissue; globus pallidus, 332 pmol/g of wet tissue; substantia nigra, 1725 to 2580 pmol/g of wet tissue; periaqueoductal central gray, 590 to 994 pmol/g of wet tissue; locus caeruleus, 332 pmol/g of wet tissue; nuclei parabranchiales, 546 pmol/g of wet tissue; medulla oblungata, 95 to 436 pmol/g of wet tissue; dorsal horn of the spinal cord, 1070 pmol/g of wet tissue; and ventral horn, 134 pmol/g of wet tissue (Kanazawa and Jessell, 1976; Douglas et al., 1982). The concentration of SP may reach values as high as 2 to 3 μ g/g of wet tissue.

In rats, both the density and the distribution of SPcontaining neurons is changing significantly during the period just before birth, the days after birth, and into the adult period. SP-staminal cells and fibers reached maximum levels between postnatal days 5 and 15. Then density generally decreased (Inagaki et al., 1982; Sakanaka et al., 1982). The distribution of NKA is less known and in the rat brain, seems to be similar to that of SP, with clearly different locations, however, in several regions. As revealed by immunohistochemistry, NKB-containing perikarya were detected in the main and accessory olfactory bulb, some cortical regions, the olfactory tubercule, the n. accumbens, the septum, the neostriatum, several hypothalamic nuclei, the superior colliculus, the substantia nigra, the medullary reticular formation, and the external caudate nucleus (Kanazawa et al., 1984; Merchenthaler et al., 1992). NKB is also located in the spinal cord, predominantly in the dorsal horn, while it is present in negligible amounts in dorsal root ganglia and dorsal roots (Ogawa et al., 1985).

In the cat brain, kassinin-LI (NKA-LI) has a wide-spread distribution with the highest concentration present in substantia nigra, hypothalamus, and caudate n.; moderate levels in thalamus, brain stem, and spinal cord; and low levels in the cortex and cerebellum. Distribution of NKA-LI paralleled that of SP, although the ratio between the two peptides varied throughout the different areas (Hunter et al., 1985).

In human brain, the areas most rich in immunoreactive SP were: amygdala, 25 to 340 pmol/g of wet tissue; nucleus caudatus, 113 to 370 pmol/g of wet tissue; putamen, 81 to 380 pmol/g of wet tissue; globus pallidus, 518 to 1800 pmol/g of wet tissue; hypothalamus, 125 to 135 pmol/g of wet tissue; substantia nigra, 1264 to 4720 pmol/g of wet tissue; and locus caeruleus, 199 pmol/g of wet tissue (Gale et al., 1978; Emson et al., 1980; Cooper et al., 1981).

Data on the occurrence of SP and SP-like peptides in the frog and fish brain are presented by Inagaki et al. (1981).

Gut. The main sources of the neuronal tachykinins in the gut are: a) the intrinsic enteric neurons of the myenteric plexus, b) the intrinsic enteric neurons of the submucosal plexus, and c) the extrinsic primary afferent fibers. The most quantitatively important source of tachykinins in the gut is the enteric nervous system, which has its cells in the wall of the intestine and supplies all gastrointestinal effector systems. The mammalian gastrointestinal tract contains both SP and NKA and various extended forms of these tachykinins.

Neurons that contain only SP (alone or with other non-tachykinin transmitters) are considered to be intrinsic sensory neurons (Holzer and Holzer-Petsche, 1997a, 1997b). In addition to these neurons, extrinsic efferent nerve fibers also display a small, but distinct contribution to the SP/NKA immunoreactivity in the gut. These fibers originate from dorsal root ganglia and reach the periphery via sympathetic or parasympathetic nerves, passing through prevertebral ganglia. The extrinsic efferent nerves project predominantly to the vessels in the intestinal wall but they also supply the lamina propria of the gastrointestinal mucosa. There are considerable species-dependent quantitative differences in the location of the tachykinins in the various gut segments, in the concentration of the peptides and in the density of SP/NKA-containing fibers.

In most species, the highest concentrations of tachykinins in the gut are found in pylorus, gastric fundus, duodenum, and jejunum (Pearse and Polak, 1975; Lazarus et al., 1980; Hunter et al., 1985; Gates et al., 1989). In the guinea pig small intestine, the bulk of SP and NKA, which are stored in the same synaptic vesicles, is associated with the myenteric plexus in longitudinal muscle.

Concerning NKB, the peptide is generally considered to be absent from human, porcine, guinea pig, and rat

intestine, which is consistent with the absence of PPT-B expression in the enteric nervous system of the rat but is in contrast with other results, showing that human and rat intestine contains minute amounts of NKB (Holzer and Holzer-Petsche, 1997a) and even more so with data showing that highly specific antiserum to NK3 receptors detected them in nervous myenteric and submucosal neurons (Grady et al., 1996). SP- but not NKA- and NKB-immunoreactivity is present also in the gall bladder and bile duct and in the pancreas, both around blood vessels and in the acini and the islets (Otsuka and Yoshioka, 1993).

Respiratory tract. RIA and immunohistochemistry have demonstrated the presence of SP and NKA in the respiratory tract of various mammalian fibers. In the trachea and bronchi, SP-immunoreactive fibers have been found in the smooth muscle layer and around local ganglion cells. In the bronchial tree, most of the SP-positive fibers are of vagal origin; but in the lung, the fibers are both of vagal and thoracic spinal origin. (Nilsson et al., 1977; Lundberg et al., 1983; Saria et al., 1985; Manzini et al., 1989).

Blood vessels. Data on the occurrence of SP-containing fibers are rather scanty and old. SP-like immunoreactivity has been observed in fibers of the adventitia and media in various blood vessels, such as feline cerebral arteries (Liu Chen et al., 1986), guinea pig intestinal vasculature (Furness et al., 1982), and rat portal vein (Barja and Mathison, 1982). The majority of SP-containing perivascular fibers are of sensory, capsaicin-sensitive origin.

Urinary system. The distribution of SP and NKA has been extensively studied in renal pelvis and ureter and especially in the urinary bladder of several species (Sharkey et al., 1983; Gibbins et al., 1985; Maggi et al., 1987). Capsaicin treatment results in an almost complete disappearance of the tachykinin-immunoreactive fibers, suggesting that the major sources of tachykinins in the urinary bladder are sensory fibers (Maggi and Meli, 1988; Maggi et al., 1988).

Skin. In human digital skin, SP- and NKA-immunoreactivity is present in free nerve endings in dermal papillae and epidermis (Dalsgaard et al., 1985; Bjorklund et al., 1986). SP-like immunoreactive fibers are also found in the skin of the rat and cat (Hokfelt et al., 1977). Treatment with capsaicin in rats caused a 70% depletion of SP-like immunoreactivity in various skin areas, suggesting that SP is present mainly in primary afferent C-fibers (Holzer, 1991).

Immune system. Tachykinin-containing primary capsaicin-sensitive afferent nerves are present in lymphoid organs, such as thymus, spleen, lymph nodes, and lymphoid aggregates in the lung and nasal mucosa. Their distribution is prevalently perivascular, but some fibers penetrate within the follicles. Both SP and NKA have been detected in rat thymus, spleen, and lymph nodes by radioimmunoassay. In addition to NKA, neu-

ropeptide K and an eledoisin-like peptide also occur in the guinea pig thymus (Geppetti et al., 1987). However, non-neuronal sources of tachykinins are also present in the immune system. Using an anti-NKA antiserum, positive immunostaining was observed in staminal cells throughout the thymic parenchyma of the rat with predominance in the medullary area (Ericsson et al., 1990).

Because endothelial cells express SP-LI (Linnik and Moskowitz, 1989; Ralevic et al., 1990), the vascular endothelium of lymphoid organs may be the source of nonneuronal tachykinins at this level. Moreover, there is evidence that certain immune cells such as eosinophils and macrophages synthesize and release SP (for review, cf. Maggi, 1997).

Blood. Quantitative data on the concentration of immunoreactive SP in blood plasma are very variable with a wide range of values obtained by the different authors, indicating that nonspecific factors are probably interfering with the assay of immunoreactive SP. This is particularly true when unextracted plasma was used. As previously stated, the major part of circulating SP evidently originates from the intestine (Pernow, 1983). Values were as follows: man 70 to 300 and 50 to 620 fmol/ml; dog, 40 to 50 fmol/ml; and calf, 165 and 18 fmol/ml for unextracted and extracted plasma, respectively.

IV. Relationships between Structure/Activity Receptor Selectivity

Tachykinins, yet defined as peptides having the characteristic C-terminal pentapeptide Phe5-Xaa4-Gly-Leu-Met-NH₂, are identified as "aromatic tachykinins" when Xaa is an aromatic amino acid residue (Phe or Tyr) and "aliphatic tachykinins" when Xaa is an aliphatic amino acid residue (Val or Ile). All natural tachykinins are amidated at their C terminus, and this function is crucial for biological activity. Deamidated peptides are virtually inactive (Erspamer, 1994).

Structure/activity relationship studies established that the C-terminal pentapeptide was essential but not sufficient for the biological activity of the tachykinins. In fact, the C-terminal pentapeptide of eledoisin and physalaemin (Bernardi et al., 1964; Regoli et al., 1994b) like that of all other examined tachykinins was virtually inactive. The minimum chain length required for activity was six residues. These studies also recognized the Phe residue at position 5 from the C terminus and the amidation at the C terminus to be crucial for biological activity, both occurring in all vertebrate and invertebrate tachykinins, as well as the presence of the C-terminal Arg-NH₂ in the locustatachykinin-like peptides. The biological activity of the tachykinins depends on their interaction with three G protein-coupled receptors—NK1, NK2, and NK3 which share considerable structural homology, reflecting their common mechanism of action.

Receptors are small proteins of 350 to 500 amino acid residues, belonging to the family of rhodopsin-like mem-

brane structures. The tachykinin receptor displaying higher affinity for SP was termed NK1, the receptor showing higher affinity for NKA was termed NK2, and the receptor showing higher affinity for NKB was termed NK3. It should be emphasized that, up to date, all naturally occurring tachykinins may act as agonists on all three receptor types, although sometimes with considerably different affinities (Regoli et al., 1987, 1994a; Maggi et al., 1993).

Parallel bioassay on a number of isolated and in situ test systems using the natural tachykinins and selective synthetic analogs, radioligand binding studies, and the use of antagonists with increasing potency and selectivity have led to the conclusion that all of the three main tachykinin receptors are heterogeneous entities, with NK1, NK2, and NK3 subtypes (Maggi et al., 1993; Quartara and Maggi, 1997, 1998). The main second messenger system coupled to activate the three known receptor subtypes is the stimulation of phospholipase C, leading to phosphoinositol breakdown and elevation of intracellular calcium (Guard and Watson, 1991). At high tachykinin concentrations, an adenylate cyclase stimulation and cAMP formation may also come into play (Nakajima et al., 1992).

The extracellular loops of these G-protein coupled receptors probably have the specific function of selecting a ligand, whereas the interaction of the ligand with transmembrane domains is responsible for receptor activation. Tachykinin peptides, therefore, presumably contain a sequence that interacts with the extracellular loops of the receptor and a sequence that interacts with transmembrane domains. Recent findings conclusively allowed clarifying the crucial importance for and the influence on receptor selectivity and activity of some key amino acids in the tachykinin sequence (Severini et al., 2000).

A. Residue Occupying Position 7 from the C Terminus

The amino acid in position seven from the C-terminal of tachykinins seems to address the peptide ligand toward the receptor. SP and tachykinins with a neutral or basic residue in this position have a preference for the NK1 receptor. Neutral residues are generally hydrophilic, and proline in position eight from the C-terminal can increase affinity for the NK1 receptor. Tachykinins with an acidic or a couple of acidic residues in position 7 or 6 and 7 from the C-terminal addressed the peptides toward the NK2 and NK3 receptors. Interestingly, the second extracellular loop has four acidic and four basic residues in the rat NK1 receptor, three acidic and two basic residues in the NK2 receptor, and one acidic and five basic residues in the NK3 receptor.

B. Residue Occupying Position 4 from the C Terminus

In all natural tachykinins, position 4 from the C terminus is occupied either by an aromatic amino acid residue (Phe, Tyr) in the aromatic tachykinins or by an aliphatic, branched amino acid residue (Val, Ile) in the aliphatic

tachykinins. The presence of an aromatic residue invariably determines selectivity or increases the selectivity of the peptide for the NK1 receptor. This is true not only when a neutral or basic amino acid residue occupies position 7 from the C terminus but also when an acidic residue occupies position 7. The couple of aromatic residues (Phe-Tyr or Phe-Phe) present in the "message domain" of the tachykinins provide specific binding interactions with transmembrane domains of NK1 receptor.

C. Residue Occupying Position 6 from the C Terminus

The presence of a Pro residue in position 6 from the C terminus causes a profound decay of biological activity. The negative contribution of Pro6 could be related to a distortion in the interaction of the C-terminal sequence of the peptide (Phe-Xaa-Gly-Leu-Met-NH2) with all tachykinin receptors. In the *Pseudophryne güntheri* tachykinins, a Glu residue occupies position 6 from the C terminus. The couple of acidic residues Asp7-Glu6 present in PG-SP1 and PG-KII could, therefore, be responsible for the marked shift of receptor selectivity toward the NK3 receptor. This shift is quite evident for PG-KII (an aliphatic tachykinin) and much less evident for PG-SP1 (an aromatic tachykinin) in which the Phe-Tyr sequence induces NK1 receptor selectivity.

D. Amino Acid Substitutions in the C-Terminal Tripeptide

To date, six natural peptides have single or double amino acid substitutions in the C-terminal tripeptide Gly-Leu-Met-NH₂: Pro (AC-AR2, AC-AR4) or Ala (ranatachy-kinin D) for Gly; and Val (AC-AR4) or Pro (ranatachy-kinin D) or Gly (AC-AR2) or Met (hylambatin) for Leu. None of these substitutions affected the peptides' receptor selectivity, only their receptor affinity or potency.

E. Pro Residue in the N-Terminal Sequence

The Pro residue is a well represented residue in the natural tachykinins. It is nearly always located in the N-terminal moiety of the peptide sequence and has a clearcut preference for positions 8 and 10 from the C terminus. In the majority of natural NK1 receptor-preferring tachykinins, a Pro residue is present at position 8, adjacent to the crucial neutral or basic residue occupying position 7. Proline in this position could modify the conformation of the C-terminal sequence of the tachykinin peptides and helps to increase their affinity and selectivity for the NK1 receptor. Cascieri et al. (1992) have suggested that all tachykinins containing Pro at position 8 from the C terminus, for example SP, have greatly reduced affinity for NK2 and NK3 receptors, and they have attributed this behavior to the preferred conformation of the Pro-containing peptides for the NK1 receptor and unfavorable for NK2 and NK3 receptors.

V. Tachykinin-Like Peptides: Pharmacological Actions

The TKs display a number of potent pharmacological actions in the periphery and in the central nervous system. In the present chapter, analysis is limited essentially to the pharmacological actions of the nonmammalian TKs (eledoisin, physalaemin, and even kassinin) available in pure form several years before the structures of SP, NKA, and NKB were elucidated. As a consequence, the pharmacology of the TKs is based largely on the study of amphibian physalaemin, kassinin, and on molluscan eledoisin.

Whereas results obtained with eledoisin and kassinin, multireceptor agonists, do not exactly mimic results obtained with either NKA or NKB, results obtained with physalaemin, a selective NK1 agonist, are perfectly superimposable, with negligible quantitative differences, on those later obtained with SP, the mammalian selective NK1 receptor agonist.

A. Cardiovascular System

1. Systemic Arterial Blood Pressure Tachykinins administered to the anesthetized dog by the parenteral route are the most potent among all known hypotensive agents. On the dog blood pressure, physalaemin was 2 to 2.5 times less potent than SP, 10 to 20 times more potent than kassinin, and 3 to 4 times more potent than eledoisin (Erspamer, 1981). When administered by rapid intravenous injection, physalaemin was 200 to 1000 times more potent than bradykinin, and 600 to 2000 times more potent than histamine (Bertaccini et al., 1965). Physalaemin was very effective in antagonizing the pressor effects of noradrenaline and angiotensin II given at doses 100 and 10 times higher, respectively.

The rabbit was again extremely sensitive to physalaemin and even more so to SP. Eledoisin was approximately 10 times less active than SP; kassinin, 20 times less active; NKA and NKB, 200 and 2000 times less active, respectively (Bianchi Porro et al., 1965; Holzer-Petsche et al., 1985).

Intravenously injected in the sheep, eledoisin showed a hypertensive response of slow onset, probably attributable to some arousal of the animals (Ormas et al., 1975).

In cat and in rat, the effect of physalaemin was considerably less intense, with high variability and tachyphylaxis. Finally, in the decapitated chicken, physalaemin regularly elicited a biphasic response consisting of a brief hypotensive phase followed by a more intense and sustained dose-related pressure increase (Bertaccini et al., 1965). The rise in pressure observed in sheep and chicken was blocked by sympatholytic drugs and by pretreatment with reserpine, thus, indicating a release of cathecolamines from the adrenal medulla and/or other stores.

Conversely, the hypotensive effect of the tachykinins was not modified by any of the usual autonomic blocking

agents, thus suggesting a direct effect on the vascular smooth muscle.

Physalaemin, ranakinin, SP, and NKB produced a dose-dependent decrease in arterial blood pressure in the toad, *Bufo marinus*. A selective NK1 antagonist had no effect on the blood pressure fall elicited by ranakinin and SP, suggesting the existence of an NK1 receptor subtype different from mammalian NK1 receptor (Courtice et al., 1993).

In the bowfin Amia calva, a teleost fish, the bolus injection into the bulbus arteriosus of 0.1 to 10 nmol/kg of the bowfin SP resulted in a significant and dosedependent rise in vascular resistance and blood pressure and a fall in cardiac output without changes in heart rate. Those effects lasted 5 to 10 min (Waugh et al., 1995b). Similarly, in the teleost fish rainbow trout, both the trout SP and the trout NKA at intraaortic doses of 1 nmol/kg increased systemic and celiac vascular resistance leading to hypertension, bradycardia, and decrease of cardiac output. After in vitro perfusion of the aortic and celiac mesenteric vascular bed, the peptides dose dependently increased the vascular resistance. It may be concluded that in teleost fish, the fish tachykinins are potent vasoconstrictor agents (Kagstrom et al., 1996).

In the conscious, unanaesthetized dogfish *Scyliorhinus canicula*, intravenous injection of either dogfish SP or scyliorhinin I (up to 5 nmol) produced no change in arterial blood pressure, pulse amplitude, and heart rate. Injection of greater amounts of the peptides (10–50 nmol) produced a slight increase in blood pressure (Waugh et al., 1993). However, in the unrestrained spiny dogfish *Squalus acanthia*, the intravenous injection of scyliorhinin I and NKA caused hypotension, due to a general vasodilation, with transient increase in mesenteric blood flow and a prolonged increase in celiac blood flow. The peptides did not increase heart rate (Kagstrom et al., 1996).

In human volunteers, eledoisin given by rapid intravenous injection (threshold 15–20 pmol/kg) decreased blood pressure, caused spinal fluid hypertension, increased the rate of respiration and caused skin vasodilation, particularly in the head. Rise in blood pressure produced by 3 to 5 nmol/kg angiotensin or 40 to 60 nmol/kg noradrenaline was inhibited or reversed by 1 to 2 nmol/kg eledoisin injected 5 s previously (Sicuteri et al., 1963).

In other experiments, the intravenous infusion of 0.6 nmol/kg/min eledoisin or 0.2 nmol/kg/min physalaemin produced only a 20 mm Hg pressure fall that lasted 5 min. Basal levels of pressure returned despite the continued infusion (De Caro et al., 1966).

SP also decreased blood pressure. A significant difference from the basal level was found at an infusion rate of 200 pmol/kg/min or higher (Eklund et al., 1977).

These results were substantially confirmed by Evans et al. (1988), who found that both SP (3 pmol/kg/min)

and NKA (64 pmol/kg/min) did not change systolic blood pressure, whereas diastolic pressure fell significantly only after SP infusion. Moreover both peptides increased heart rate and body temperature, with skin flushing. SP was 6 to 20 times more potent than NKA.

Heart. Electrocardiogram tracings recorded from anesthetized dogs given an intravenous infusion or a subcutaneous injection of physalaemin, at doses approximately 1000 higher than the threshold hypotensive dose, produced only moderate electrocardiographic changes mainly attributable to hypotension (Bertaccini et al., 1965). In a detailed study, the following percentage changes in a number of cardiovascular parameters have been observed in dog after intravenous injection of 4 pmol/kg physalaemin: heart rate, +17.8; mean systemic arterial pressure, -22.3; mean pulmonary arterial pressure, +1.8; mean left atrial pressure, -1; mean right atrial pressure. +0.5: myocardial contractile force. +17.5; cardiac output, +52; total peripheral resistance, -65.2; and pulmonary vascular resistance, -35.4 (Nakano et al., 1968). Similar results were obtained with eledoisin (Nakano, 1964, 1965).

The effect of SP was substantially the same as that observed with physalaemin. At infusion rates ranging from 3 to 450 pmol/kg/min, SP invariably induced a dose-dependent increase of cardiac output mostly due to a larger stroke volume. SP at concentrations up to 50 pmol/ml had no effect either on the isolated guinea pig auricles or the perfused rabbit heart, suggesting that SP has no direct effect on the heart (Burcher et al., 1977).

2. Regional Circulation. Coronary bed. Physalaemin and, to a considerably lesser extent eledoisin and SP (Losay et al., 1977) displayed a very potent vasodilator action on the dog coronary vascular bed not only when given by intracoronary administration, but also when given by intravenous infusion. A transient, 50% increase in coronary flow was obtained by rapid intracoronary injection with 0.1 pmol/kg and a 100% increase with 1 pmol/kg of physalaemin. Eledoisin was 200 times less active and nitroglycerin, 10,000 times less active.

Eledoisin infused intracoronarily at a rate of 6 pmol/kg/min increased sinus coronary outflow by 20%, coronary sinus oxygen tension by 10%, and similarly increased stroke flow and cardiac oxygen consumption, without affecting mean arterial pressure and heart rate (Lochner and Parratt, 1966). Increase in coronary flow and decrease in coronary vascular resistance also was observed after intravenous infusion of eledoisin (Beretta Anguissola et al., 1966).

Skeletal muscle. The vessels of the skeletal musculature of the hindlimbs of dogs were by far the most sensitive to tachykinins of any vascular bed. Doses of eledoisin as low as 10 fmol injected into the peronal artery caused an increase in blood flow, both in the intact and denervated gastrocnemius plantaris muscle. Denervation enhanced the potency of eledoisin (Bergamaschi and Glasser, 1963, 1964). Physalaemin was 50

times more potent than eledoisin and 50,000 times more potent than nitroglycerin (Bergamaschi et al., 1966; Fregnan and Glasser, 1968). In other experiments, close arterial injection of SP caused a dose-related vasodilation in adipose tissue and skeletal muscle of the dog only with doses starting from 10 nmol (Pernow and Rosell, 1975).

SP was also a potent vasodilator in humans. Infusion of 0.7 pmol/kg/min into the brachial artery significantly increased the forearm blood flow, with increases of oxygen consumption in both cutaneous and muscle blood. At the infusion rate of 70 pmol/kg/min, there was a bright red flushing of the skin, particularly in the neck and head, with a subjective feeling of warmth in the same regions, accompanied by tachycardia (Eklund et al., 1977). No effect of SP could be seen on internal carotid blood flow (Samnegard et al., 1978).

Liver. Portal or femoral infusions of SP, eledoisin. and physalaemin (2-20 pmol/kg/min) increased blood flow in the hepatic artery and vein of the dog. Portal infusions were less effective, thus, indicating a highly inactivating capacity of the liver. Hepatic arterial and venous pressures decreased, whereas sinusoid and portal pressure increased during peptide infusion. As a consequence, hepatic arterial and outflow resistances decreased. SP was the most potent peptide, followed by physalaemin (38%) and eledoisin (10%). When given by close arterial infusion, the peptides also consistently increased blood flow in the hepatic artery (Melchiorri et al., 1977). These observations were confirmed by Takaori et al. (1989), who found that intravenous physalaemin (5 pmol/kg) caused dose-dependent increases in mesenteric arterial blood flow (70%) and portal venous blood flow (77%) in the dog.

Lung. Intravenous eledoisin (0.1–1 nmol/kg) did not increase, and sometimes slightly reduced, pulmonary arterial pressure in the guinea pig; it always increased pressure in the rabbit. In the isolated, bloodperfused rabbit lung preparation, eledoisin produced a potent vasoconstriction from threshold doses of 0.01 to 0.1 pmol/kg. Tachyphylaxis was obvious. Bradykinin was 1000 times less effective (Hauge et al., 1966). In the dog NKA was much more potent than SP in decreasing tracheal vascular resistance (Salonen et al., 1988).

Skin. The skin vasculature of the dog was far less sensitive than the vessels of the musculature. In man, injection of 0.2 nmol eledoisin into the brachial artery produced digital vascular responses consisting of an increase in the skin temperature and a consistent increase in total digital volume, despite a decrease in inflow volume. Responses seem to indicate a closure of the arteriovenous anastomoses (De Pasquale and Burch, 1966).

Infusion of SP into the rat femoral artery dose dependently produced vasodilation (threshold 0.1 pmol/kg/min) that was inhibited by mepyramine (Lembeck and Holzer, 1979).

Brain. Eledoisin infused intravenously in the dog at 0.01 nmol/kg/min decreased cerebral blood flow (-22%), with an increase (+20%) in vascular resistance (Beretta Anguissola et al., 1966). In human subjects, the intravenous infusion of eledoisin (1–15 pmol/kg/min) influenced neither the cerebral blood flow and vascular resistance nor the cerebral metabolic rate of oxygen and glucose (Bianchi Porro et al., 1965).

Mechanism of vasodilation and hypotension. All of these findings demonstrate that in some mammalian species, exogenous tachykining display a potent dilation of regional musculature accompanied by the fall of systemic blood pressure, in other mammalian and nonmammalian species the peptides display inconstant and variable effects: hypotensive/hypertensive or even frank hypertensive responses. Thus, the intervention of endogenous tachykinins in the regulation of blood pressure and regional circulation is certainly possible but irregular and unpredictable. At any rate, it is hardly conceivable that the tachykinins display a significant role in the cardiovascular system similar to that of noradrenaline, serotonin, angiotensin, prostacyclins, etc. This does not exclude that in man the tachykinins may contribute to the control of vascular tone of the cutaneous vessels of some areas. So, it has been suggested that tachykinin (SP and NKA) release is co-involved in the pathogenesis of flushing episodes (not accompanied by edema!) occurring in the carcinoid disease. To our knowledge, tachykinin antagonists have never been used in this disease. The trial could be rewarding from both a pathogenetical and a therapeutical point of view.

The striking hypotensive effects of the tachykinins observed in some animal species, as a consequence of intense vasodilation in several peripheral vascular beds, must be considered a direct effect of the peptides on the blood vessel wall. However, Regoli et al. (1987), D'Orléans-Juste et al. (1985, 1986), and Mastrangelo et al. (1987), in agreement with previous observations (Furchgott, 1983, 1984) on other vasodilators, found that in isolated strips of arteries and veins that were maximally contracted by noradrenaline, the relaxing effect of the tachykinins could be obtained only when the endothelium was intact. This clearly indicated that the site of action of the tachykinins (like that of acetylcholine, bradykinin, neurotensin, and bombesin) was not the smooth muscle cell, as formerly believed, but the endothelium. Thus, the tachykinins may act to promote the release of endogenous factors (prostacyclins, endothelium-delivered relaxing factors, and nitrous oxide) from the endothelium that is able to reduce the tone of the arterial smooth muscle fibers. However, whereas dog carotid artery without endothelium is insensitive to all tachykinins, thus, suggesting an exclusive location of its receptors in the endothelium, this is not true for the other vascular preparations. The rabbit pulmonary artery, for example, may be relaxed or contracted by tachykinins, depending on its basal tone. At high tone levels (premedication with noradrenaline), relaxation was predominant; whereas at low tone levels, contraction occurred. Relaxation may be due to the activation of NK1 receptors (SP, physalaemin) located in the endothelium, and contraction may be caused by activation of NK2 receptors (NKA, kassinin) located on the arterial smooth muscle. In the rabbit pulmonary artery possessing endothelium, the relaxing potency of SP was 3 to 4 times higher than that of NKA or kassinin; in the artery without endothelium, the contracting potency of SP was 30 to 120 times lower than that of NKA or kassinin (D'Orléans-Juste et al., 1986). Similarly, in the rat portal vein (with intact endothelium and not pretreated with noradrenaline), contraction elicited by NKB and kassinin was brought about by NK3 receptors, presumably located on the smooth muscle membrane (Mastrangelo et al., 1987).

In the intact animal, response of the vasculature to tachykinins is complex, depending on the animal species, density in the smooth muscle cells, and the endothelium of the different receptor types, as well as the kind of tachykinins administered or released. Whereas in dogs and rabbits, the NK1-preferring tachykinins (SP, physalaemin) regularly cause a dose-dependent intense hypotension with no sign of tachyphylaxis, the same tachykinins in cats, sheeps, rats, and pigeons may produce moderate hypotension with obvious tachyphylaxis, hypotension/hypertension, or frank hypertension, thus, indicating a complex activation of different receptor types, and perhaps a different availability of relaxing factors in the endothelium (tachyphylaxis).

Capillary permeability. The capillary permeability was enhanced by intradermal injection of eledoisin and physalaemin in the rat, guinea pig, and man. Eledoisin was 2 to 10 times more potent than histamine in all animal species and more potent than bradykinin in man (De Caro, 1963). Intradermal injection of eledoisin at doses exceeding 1 pmol in man caused pain, local edema, and erythema; whereas when injected intraperitoneally, intramuscolarly, or subcutaneously into human subjects at doses up to 17 to 50 nmol, the peptide failed to elicit pain (Kantor et al., 1967). Similarly, intradermal injection of 0.1 to 10 μ M SP in man induced flare, wheal, and itching, which was similar to the response induced by histamine and, like histamine, was blocked by antihistaminic drugs (Hagermark et al., 1978).

Introduced in the mouse pleural cavity, SP (ED $_{50}$ = 14 nM) caused a long-lasting recruitment of leukocytes and a small but evident exudation. These effects were partially inhibited by NK1 receptor antagonists and were mediated by nitric oxide (Frode-Saleh et al., 1999). Finally, close-arterial infusion of SP, from doses as low as 0.1–0.5 pmol/min in the rat skin, induced vasodilation and plasma extravasation (Lembeck and Holzer, 1979).

Of great interest is that intradermal injection of SP (30–300 pmol) induced a dose-dependent edema on wild-

type mice, whereas in NK1 receptor knockout mice, the peptide was inactive. The reaction in wild mice was reduced by the histamine antagonist mepyramine, indicating that edema induced by the tachykinin, although totally dependent on NK1 receptor-mediated mechanism, contains a mast cell-dependent component (Cao et al., 1998).

Intravenously injected in the rat, NKA induced a dose-dependent extravasation of Evans blue in stomach, duodenum, jejunum, caecum, and colon but had no effect in ileum. NKB equipotently produced extravasation in the stomach but had no effect in other parts of the gut. SP was ineffective (Lordal et al., 1996).

NKA (100–400 pmol/kg/min) given by intravenous infusion in an in situ perfusion model of the anesthetized rat stimulated duodenal motility and increased duodenal mucosal bicarbonate secretion, fluid output, and mucosal permeability. Effects were dependent on NK2 receptor activation (Hallgren et al., 1997). Plasma extravasation was induced by SP and bradykinin also in mouse gastrointestinal tract and pancreas (Figini et al., 1997). On the contrary, neither capsaicin nor SP (0.1–1 mg/ml) nebulized or given intravenously (0.75 nmol/kg) in rabbits produced significant increases in tracheal or bronchial Evans blue concentration, indicating that SP does not activate the microvascular leakage in the major airways of the rabbit (Matheson et al., 1997).

B. Gastrointestinal Tract

1. Motility. The stimulant action on intestinal motility of crude extracts, together with their hypotensive action, was at the root of the discovery first of SP and then of eledoisin and physalaemin.

In mammalian and submammalian vertebrates, the tachykinins provoke, with few exceptions, a contractile response by the gut. The excitatory motor effects were evident in all sections of the gut, from esophagus to the rectum, and in all muscular layers, including the longitudinal muscle, the circular muscle, and the muscularis mucosae. However, motor effects may be sharply different depending on the animal species, the gut sections, the different receptor types activated, and the mechanisms involved in the motor response (direct effect on the smooth muscle and indirect effect, through implication of the many neurotransmitters and hormones that are active on the gut motility) (Holzer-Petsche, 1995; Holzer and Holzer-Petsche, 1997a, 1997b; Maggi et al., 1997).

a. In Vitro Experiments. As many as 16 isolated gastrointestinal preparations from eight animal species were assayed with eledoisin and physalaemin (Erspamer and Falconieri Erspamer, 1962; Bertaccini et al., 1965). Response was always stimulation, but intensity and reproducibility of response varied conspicuously dependent on the animal species and the various segments of the intestinal tract. After synthetic SP and, later on, NKA and NKB became available, an enormous amount

of work appeared on the action of mammalian tachykinins on isolated preparations of intestinal muscle and the receptors herein involved (Holzer and Holzer-Petsche, 1997a).

b. In Vivo Experiments. Dog. The first phenomenon observed a few minutes after a subcutaneous administration of 25 to 200 nmol/kg eledoisin was vomiting accompanied by profuse salivation. Vomiting was at first alimentary, and then the emesis episodes of increasing severity resulted in the ejection of masses of mucus, sometimes spotted with blood. Shortly after commencement of vomiting, there was a discharge of formed stools, which was soon followed by evacuation of watery stools containing mucus and blood accompanied by violent tenesmus. The tremendous gastrointestinal stimulation and profound depression of the animal lasted unchanged for 1 h and then gradually decreased. During the night, there was complete recovery, and for a few days afterward, the dog showed unusual voracity. In dogs given a subcutaneous dose of 0.15 mg/kg atropine sulfate 30 min before the injection of 20 nmol/kg eledoisin, the alkaloid failed to block stimulation of either salivary glands or gastrointestinal smooth muscle (Erspamer and Glasser, 1963).

Subcutaneous injections of 100 to 250 nmol/kg of physalaemin elicited phenomena similar to those provoked by 25 to 100 nmol/kg eledoisin, but vomiting and diarrhea accompanied by profuse salivation were less severe and recovery was rapid. Doses of 25 to 50 nmol/kg physalaemin caused moderate salivation, moderate evacuation of formed stools, and inconstant vomiting. No consistent gastrointestinal effects, except slight salivation, appeared after doses of 10 nmol/kg physalaemin (Bertaccini et al., 1965).

The general effects produced by rapid intravenous injection of physalaemin in dogs were studied by Bertazzoli and Cheli (personal communication). No obvious effects were elicited by 0.1 nmol/kg of physalaemin; after 0.3 nmol/kg, the only appreciable effect was that the dog had difficulty in sitting (tenesmus?), and this lasted only for a few minutes; 0.6 nmol/kg produced the same effect and, in addition, one or more evacuations of the bowel during the first 5 min; 2.5 nmol/kg caused not only diarrhea but also salivation, lasting 5 to 10 min. Increased salivation and diarrhea were accompanied by vomiting in dogs treated with 12 nmol/kg of physalaemin. All symptoms disappeared within 15 to 20 min. Similar doses of physalaemin (2.5 nmol/kg) given intravenously every day for 10 successive days elicited the same phenomena.

In the anesthetized dog, intravenous infusion of eledoisin at a rate of 40 pmol/kg/min produced neither salivation nor evacuation of the bowel. Infusion rates of 80 to 250 pmol/kg/min, on the other hand, caused salivation and evacuation of liquid stools. Stimulation of both salivary glands and gastrointestinal smooth muscle was atropine-resistant. High infusion rates produced a

more or less evident cutaneous vasodilation (Erspamer and Glasser, 1963).

In conscious dogs, eledoisin and physalaemin displayed a potent stimulating effect on the mechanical and electrical activity of the gut. Low doses of the peptides (2.5-15 ng/kg/min) caused an increase in frequency and duration of the interdigestive myoelectric complexes, and an increase in coordinated mechanical activity. High doses provoked the appearance of a diffuse spike activity, accompanied by intense local motor activity. Pacesetter potentials were not affected (Caprilli et al... 1975). After intravenous injection of 1 nmol/kg physalaemin in anesthetized dogs with a gastric pouch a striking increase in tonus of the gastric pouch was observed, accompanied by stimulation of rhythmic contractions, lasting 10 min. In the ileum a slight initial increase in tonus was followed by intense stimulation of rhythmic activity, lasting 10 to 15 min (Bertaccini, 1982).

In other experiments on the in situ jejuneal loop of the anesthetized dog, physalaemin was twice as potent as CCK in stimulating motility, 15 times as potent as human gastrin I, 50 to 100 times as potent as either bradykinin or carbachol, and more than 300 times as potent as acetylcholine, histamine, and serotonin. Only caerulein overcame physalaemin (by 3 times) in its stimulating effect (Mantovani et al., 1969).

In keeping with these results, the tachykinins administered by close intra-arterial injection increased the motility of gastric antrum and proximal duodenum. Physalaemin and eledoisin (5 pmol/ml) were the most potent peptides, followed by SP (50–60%) and neurokinins A and B (10–12%) (Kuwahara and Yanaihara, 1987). The effect of SP on gastric motility was found to differ depending on the sites and vagal innervation of conscious dogs (Shibata et al., 1994).

Cat. Physalaemin, eledoisin, and SP administered via the splanchnic artery produced both motor and mechanical effects in the stomach of anesthetized cats, characterized by successive phases of initial distension, sustained contraction, and late distension. At low doses, distension was the dominant effect. The sustained contraction and late distension phase were accompanied by phasic contractions. Atropine abolished the sustained contractions but had no effect on phasic contractions and distension phases (Lidberg et al., 1983; Barber et al., 1987).

Guinea pig. Tachykinins influenced peristaltic motor activity in isolated segments of the guinea pig small intestine. NK2 and particularly NK3 agonists facilitated intestinal peristalsis, whereas SP first stimulated and then inhibited peristalsis. The facilitatory effect of SP was prevented by atropine and seems to involve NK2 receptors, whereas the secondary inhibitory effect is due to NK1 stimulation (Holzer-Petsche, 1995).

Rat. The tachykinins displayed a potent spasmogenic action on the rat stomach, as measured from the

bulk of duodenal effluent. In terms of threshold doses (0.1–1 nmol/kg intravenous), eledoisin was the most potent peptide, SP was the least potent, and phyllomedusin was by far the most effective with regards to the duration of effect (Bertaccini and Coruzzi, 1977; Bertaccini, 1980). After infusion into the celiac artery at doses of 0.06 to 20 nmol/min, both SP and NKA caused contraction of the stomach, NKA being 10 times more potent than SP (Holzer-Petsche et al., 1987).

The complexity of the effects of the tachykinins on the gastrointestinal propulsion was evident after the intraperitoneal injection of the peptides. It was shown that 3 min after a test meal, both SP (> 0.75 nmol/kg) and NKA (> 8.8 nmol/kg) inhibited both gastric emptying and intestinal transit. The inhibitory effect was reversed to a stimulant effect by pretreatment with atropine. After 10 min, SP dose dependently enhanced intestinal propulsion, an effect that was atropine resistant. From the above experiments, it clearly seems that the gastrointestinal propulsion was dose- and time-dependent, with variable involvement of the autonomic nervous system (Holzer, 1985).

Sheep. Intravenous eledoisin (50–250 pmol/kg) stimulated both in anesthetized and in awake and standing animals the motility of all sections of the stomach producing an increase in tone and in rhythmic movements. The effect was immediate, atropine resistant, and lasted from 5 to 30 min, depending on the dose. The omasum was the most sensitive section. SP was at least 10 times less active than eledoisin (Ormas et al., 1977).

Toad. On the B. marinus intestine, bufokinin was the most potent agonist ($EC_{50}=0.35~\mu\mathrm{M}$), producing a long-lasting contraction similar to that evoked by physalaemin, SP, and kassinin. Surprisingly, these effects were not inhibited by highly selective NK1 receptor antagonists (Liu et al., 1999b).

Fish. The stimulant effect of the tachykinins was also evident in the fish gut. Both SP and NKA produced contraction of the vascularly preferred cod stomach; SP (pD2 7.05) was almost 6 times more potent than NKA (Jensen, 1997). The same peptides displayed an excitatory effect on the circular muscle of the cod intestine, suggesting that tachykininergic neurons are involved in the ascending excitatory reflex of peristalsis (Karila et al., 1998).

2. Secretions. Salivary secretion. The potent sialagogic effect of substance P (Haefeli and Hurlimann, 1962) and of amphibian tachykinins (Bertaccini and De Caro, 1965; Emmelin and Lenninger, 1967) was recognized several years before the sialagogic principle in a bovine hypothalamic extract was identified as substance P by Chang and Leeman (1970).

Physalaemin displayed a powerful sialagogic effect in dogs and rats. In dogs (with cannulated submaxillary gland), the threshold dose of the peptide was 0.5 nmol/kg when injected into the femoral vein and 0.1 nmol/kg when injected through the ipsilateral carotid artery. The

threshold dose by intravenous infusion was 0.1 to 0.4 nmol/kg/min. Salivary flow never lasted more than 5 to 8 min after the injection. Response was dose dependent and tachyphylaxis was lacking. The effect was appreciable even at a very low systemic blood pressure. High amounts of parasympathetic and sympathetic blocking drugs failed to block salivation induced by physalaemin, thus, indicating a direct point of action of the peptide on the acinar cells. The sialagogic activity of physalaemin exceeded eledoisin by three times, carbachol by seven times, and other known sialagogic agents by more than 100 times (Bertaccini and De Caro, 1965).

These results were confirmed by Emmelin et al. (1969), who found that physalaemin stimulated secretion from the dog parotid and submaxillary glands (threshold doses: 0.5 and 12 nmol/kg, respectively). Considerably lower doses (threshold: 5–10 pmol/kg) elicited a pressure increase in both submaxillary and parotid ducts. The contraction of the myoepithelial cells of the ducts was due to a direct effect of the peptide, because the usual autonomic blocking agents did not abolish it.

In rats, the threshold sialagogic subcutaneous dose of physalaemin was 0.1 to 0.3 nmol/kg. There was a satisfactory dose-response relationship, and the stimulatory effect could be repeated for 2 to 3 h. By intravenous infusion (threshold: 0.2 nmol/kg), salivary stimulation lasted as long as the infusion was continued (Bertaccini and De Caro, 1965). Increase in salivary flow produced both by intravenous injection (0.1–1 nmol/kg) and infusion (1 nmol/kg/min) was associated with an increase in amylase and electrolyte secretion. The flow rate of the submaxillary gland was maintained throughout the period of infusion, whereas the parotid flow rate tended to decrease, finally ceasing entirely. Again, autonomic blocking drugs failed to modify the response to physalaemin (Schneyer and Hall, 1968).

By close intra-arterial injection, the threshold sialagogic dose of physalaemin was much lower (1.5 pmol) for the submaxillary gland than for the parotid gland (62 pmol). An increase in flow of saliva occurred along with a pressure rise in the duct of the submaxillary gland (Thulin, 1976).

Chronic intravenous administration of physalaemin (10 nmol/kg twice daily for 15 days) caused a moderate enlargement of the parotid and submaxillary glands, with a 50% increase in weight. Eledoisin was inactive at doses up to 500 nmol/kg (Bertaccini et al., 1966; Cantalamessa et al., 1975). Eledoisin and physalaemin also caused salivary secretion in the hen at intravenous doses of 0.3 to 0.5 nmol/kg. Physalaemin was 8 to 10 times more potent than eledoisin (Lembeck and Starke, 1968).

As far as mammalian tachykinins are concerned, results obtained mainly with SP confirmed and extended results obtained with physalaemin. In summarizing, it was established that: a) SP potently stimulated salivation in the dog, ferret, rat, and guinea pig; whereas in

humans, cat, rabbit, mouse, and hamster, it was virtually inactive (Lembeck and Starke, 1968; Leubeck et al., 1968; Iwabuchi et al., 1992; Tobin and Ekstrom, 1992). b) The principal effect of SP and NKA was to enhance the flow of salivary fluid, which is poor in protein, through a predominant action on the secretory structures of the acini. c) The secretion of fluid was stimulated in all salivary glands of the rat, but the submaxillar glands were most sensitive to SP, whereas the parotid glands and particularly the sublingual glands were less sensitive (Ekstrom et al., 1983), d) SP also produced changes in the output of salivary components, as shown by the increase in the secretion of K⁺ and Cl⁻ ions, in discharge of proteins, glycoproteins, proteolytic enzymes, amylase, kallikrein, and mucus (Holzer and Holzer-Petsche, 1997b). e) The rank order of potency in causing salivary secretion in rats after intravenous iniection was: physalaemin = uperolein > eledoisin > SP = kassinin > NKA >> NKB (Holzer-Petsche et al., 1985).

The greater potency of physalaemin and uperolein in comparison with SP may be explained by the considerably higher resistance to enzyme attack offered in the two first peptides by their N-terminal pGlu residue (Yanaihara et al., 1977). It is generally accepted that the secretory action of the tachykinins in the rat salivary glands is mediated by the NK1 receptor (Holzer and Holzer-Petsche, 1997b).

In slices of rat parotid glands, the tachykinins caused a rapid, concentration-dependent (threshold 10-15 pmol/ml) increase in K^+ efflux and amylase release, independent of any effect on tissue concentrations of cyclic AMP or cyclic GMP levels. Simultaneously, there was a stimulation of phosphatidylinositol turnover (Rudlich and Butcher, 1976; Hanley et al., 1980). Physalaemin and substance P were emulative, eledoisin was one-half as potent, and kassinin was 10 times less potent (Brown and Hanley, 1981).

Gastric acid and biliary secretions. Physalaemin did not influence the gastric acid secretion in fasting dogs at the maximum tolerated intravenous dose (30 nmol/kg). Gastric acid secretion of the rat was stimulated very little, if at all. In the cod, a teleost fish, physalaemin and eledoisin potently stimulated gastric pepsin secretion (ED₅₀ = 1 mg/kg/min); SP was 1000 times less active (Holstein and Cederberg, 1986). In the isolated porcine nonantral stomach preparation, 0.1 µM SP induced a 2-fold increase in acid secretion and a 3- to 4-fold increase in pepsinogen secretion. Similar effects were displayed by NKA, whereas capsaicin was inactive (Schmidt et al., 1999). This result may indicate that release of endogenous SP/NKA is insufficient to affect the above parameters. In the dog, the peptide caused changes in bile flow that were associated with contraction of the gall bladder and not with a stimulated secretory activity. In fact, when the hepatic and cystic ducts were cannulated separately, physalaemin increased the

flow in the cystic duct but decreased the flow in the hepatic duct. In the rat, the peptide was completely ineffective (Bertaccini et al., 1967).

In agreement with the above results it was found that SP also attenuated basal and hormone-stimulated (cholecystokinin and vasoactive intestinal peptide) overall bile flow and output of bile acids, Na⁺, K⁺, Cl⁻, and bicarbonate (Starke et al., 1968; Holm et al., 1978; Konturek et al., 1981).

Intestinal secretion. That physalaemin and eledoisin stimulate intestinal secretion is shown by the liquid discharges produced after subcutaneous administration of these peptides. In dogs with a cannulated 25-cm jejunum segment perfused with saline, the intravenous injection of 55 pmol/kg/min SP produced an increase in plasma SP concentration from 6 to 121 fmol/ml, an increase in intestinal secretion of water (from 102 to 275 μ l/min) and Na⁺ (from 20 to 23.2 μ Eq/min), and a decrease in Cl⁻ secretion (from 21.7 to 16.5 μ Eq/min) (McFadden et al., 1986).

Close intraarterial infusion of SP (causing a plasma concentration of 1–5 μ M) to the in vivo feline small intestine regularly evoked a net fluid secretion in vivo, accompanied by release into the blood of vasoactive intestinal polypeptide (Brunsson et al., 1995).

The problem of receptors and mediator systems involved in the in vivo and in vitro secretory effects of the tachykinins is extremely complex. NK1 and NK2 receptors seem to play a predominant role. In the canine intestine, implication of NK2 receptors in addition to NK1 receptors is demonstrated by the greater potency of eledoisin in comparison with physalaemin in eliciting watery discharges.

Pancreatic secretion. Physalaemin displayed a moderate, short-lasting stimulatory action on exocrine pancreatic secretion in the dog after threshold intravenous doses of 0.05 to 0.5 nmol/kg. Eledoisin was 30 times less active. The content of amylase in pancreatic juice was similar to that present in the secretion elicited by cholecystokinin (Bertaccini et al., 1967).

Like eledoisin, kassinin was virtually inactive on the dog pancreas. SP enhanced the basal output of pancreatic juice, amylase, and bicarbonate in dogs (20 pmol/kg/min, by close intraarterial infusion) and rats (Thulin and Holm, 1977; Konturek et al., 1981). However, the effect of the tachykinins was negligible in comparison with that of cholecystokinin.

In dispersed acinar cells prepared from guinea pig pancreas, physalaemin and eledoisin increased outflow of Ca²⁺, accumulated cyclic GMP, and released amylase without affecting cyclic AMP. The efficacy relative to caerulein was 29% for eledoisin and 17% for physalaemin (May et al., 1978; Jensen and Gardner, 1979). In the isolated rat pancreas, 0.1 to 1 nM SP inhibited cholecystochynin-induced amylase release and secretin-induced flow. Capsaicin displayed the same effects as SP, prob-

ably through release of endogenous SP (Kirkwood et al., 1999).

The functional significance of neuronal tachykinins in the gut has been the object of an extensive and thorough investigation (Holzer-Petsche, 1995; Holzer and Holzer-Petsche, 1997a, 1997b). All aspects of the tachykinin function in the gut have been taken in consideration, but it is extremely difficult to draw some firmly established conclusions from the enormous amount of data published. Pharmacological "in vitro" tests have clearly demonstrated that exogenous tachykining given alone are among the most active substances on the gastrointestinal motility of all examined vertebrate species, generally in the sense of clear-cut stimulation. Things are different in the intact organism. The tachykinins represent only one of the many active agents of peptide and nonpeptide nature (gastrin, cholecystokinin, motilin, enteroglucagon, bombesin, guanvlin, neuropeptide tvrosin, opioid peptides, noradrenaline, histamine, serotonin, acetylcholine, prostaglandins, ATP, etc.), and it is virtually impossible at present time to distinguish the part played in the functional control of gut motility by the tachykinins from that played by the array of the above active substances. It is highly probable that the tachykinin co-involvement may be considerably different in the different animal species and in the different gut sections. The use of tachykinin antagonists has given only partial and ambiguous results. In conclusion, whereas, there is no doubt that endogenous SP and NKA may play an important role by interaction with other enteric transmitters in the control of gastrointestinal motor activity, the weight of this role in health and disease remains to be defined.

Exogenous tachykinins seem to cause also secretion of fluid and electrolytes from the intestinal mucosa, and it has been suggested that endogenous tachykinins may play a messenger role in intestinal secretory pathway. The increase in the capillary permeability could contribute to increase in secretions. Moreover, there is evidence that the tachykinins participate in the hypersecretory. vascular, and immunological disturbance, associated with infection and inflammatory bowel disease (Holzer and Holzer-Petsche, 1997a, 1997b). This statement must be again accepted with caution, keeping in mind that the tachykinins are also in the case of secretions and, even more so, of immunological reactions only one of the numerous factors involved in these processes. To remain in the field of gut secretions, in recent years a peptide, guanylin, has been extracted and isolated from various organs, among which is the rat intestine (Currie et al., 1992) in which it seems to be costored with serotonin in some populations of the enterochromaffin cells (Cetin et al., 1994). Guanylin exhibits high sequence and structural homology with Escherichia coli heat-stable enterotoxin and, like this toxin, guanylin causes secretory diarrhea in rats, by activation of the guanylate cyclase C receptors. It is evident that guanylin may play an important role in the physiological regulation of electrolyte/water secretion in ion-transporting intestinal epithelium.

Some questions at this point are imperative. Do some enterochromaffin cell populations costore and cosecrete SP and guanylin? Could guanylin be costored with SP also in the nonargentaffin, but argyrophyl/acidophyl cells of the gut? Is it possible that guanylin is present also in carcinoid tumors, contributing to the diarrhea, which is one of the most frequent symptoms in the carcinoid syndrome? Should the answer to these questions be positive, the tachykinin contribution in the regulation of gut secretion could be attributable not to neuronal SP but to SP occurring in the highly neglected epithelial cells of the gut, probably provided with paracrine secretion.

C. Airways System

A detailed description of the effects of the tachykinins on isolated preparations of tracheal and bronchial musculature in the rat, guinea pig, ferret, hamster, and man is presented by Frossard and Advenier (1991), together with a discussion of the receptor types and subtypes involved in the response to the tachykinins. Under physiological conditions, tachykinins contribute, to some extent, in the regulation of the tone of the airways musculature, at least in some animal species.

In spontaneously breathing guinea pigs, graded doses of eledoisin (2–5 nmol/kg, intravenous) produced a dose-dependent reduction of the tidal volume associated with temporary tachypnoea. With 5 nmol/kg, the tidal volume approached zero, although the movements of the respiratory muscle had increased. Eledoisin was one-half as active as serotonin.

In mechanically respired guinea pigs, eledoisin reduced the inflation volume in a dose-dependent manner; a 50% reduction was obtained at a dose of 1 nmol/kg. Serotonin was 3 to 4 times less active (Gjuris and Westermann, 1965). These results were confirmed by Nilsson et al. (1977), who observed that intravenous injection of SP produced a dose-dependent elevation of insufflation pressure. SP (0.4 nmol/kg) increased the pressure by 100%, an effect that required doses of histamine 40 times higher. Similarly, Hua et al. (1984), found that kassinin, eledoisin, and NKA potently increased insufflation pressure. Physalaemin was less potent, and NKB still less so. Moreover, in guinea pig airways, physalaemin, eledoisin, and SP provoked an increase in microvascular permeability to protein by activating receptors localized in the endothelial cells, as assessed by Evans blue extravasation (Rogers et al., 1988). It is suggested that both NK1 and NK2 receptors are involved in the response to tachykinins by the guinea pig tracheobronchial tree (Ireland et al., 1991; Maggi et al., 1991).

In rats, the broncho-constrictor action elicited by the intravenous injection of tachykinins was inhibited largely by atropine (suggesting a release of acetylcho-

line), and by methysergide (suggesting a release of 5-HT from pulmonary mast cells). Eledoisin and kassinin were slightly but significantly more potent than the neurokinins but much more potent than SP (Joos et al., 1986). In anesthetized rabbits, peripheral administration of either SP or NKA (0.2–2 nmol/kg) produced a dose-related increase in rapidly adapting pulmonary stretch receptor activity without any significant changes in total lung resistance. NKA was less potent than SP, NKB was practically inactive. This effect is mediated by activation of both NK1 and NK2 receptors (Matsumoto et al., 1997).

In the anesthetized dog, challenge with aerosolized NKA (0.1–1%) produced a dose-dependent increase in lung resistance, a decrease in dynamic lung compliance, reduced tidal volume, and increased respiratory rate. Experiments with selective tachykinin antagonists suggest that in addition to the NK2 receptor the NK1 receptor may also be involved in the response to NKA by the dog respiratory tract. Cholinergic reflexes may play a small, but significant role in this response (Sherwood et al., 1977).

In humans, infusion of SP had little effect on airway function. A small increase in airway resistance was observed at low dose and converted to bronchodilation at high doses (3.2 pmol/kg/min) (Evans et al., 1988). Inhaled SP also failed to increase airway resistance in both normal and asthmatic subjects. NKA conversely induced a fall in specific airways conductance in patients with mild asthma, suggesting an activation of NK2 receptors (Joos et al., 1986). In man, it is rather doubtful from the available data that the tachykinins play some role in disease, such as asthma, and that tachykinin receptor antagonists may have a future in therapeutics of respiratory disease. One of the major symptoms of the carcinoid syndrome is asthmatic attack. It would be important to know whether these attacks benefit from administration of tachykinin antagonists.

D. Urogenital Tract

Exogenous tachykinins at extraphysiological concentrations produced variable degrees of stimulation of smooth muscle preparations of the urogenital tract, especially the urinary bladder and displayed differences in their agonistic potency not only depending on the different animal species but also on the various segments of the urinary tract. The different kinds of responses seem to be mediated through different types of receptors and seem to be brought about both by a direct effect on the bladder's smooth muscle and by an effect on intramural sensory nervous pathways ("micturition reflex") (Maggi and Meli, 1986; Maggi et al., 1986a, 1986b; Maggi 1991). Activation of rat bladder motility and the micturition reflex may be provoked by intravenous injection and by topical application of tachykinins on the serosal surface of the bladder.

Intravenous injection of the peptides elicited a phasic contraction of the bladder (increase of internal pressure) and an activation of a series of rhythmic contractions. Kassinin was the most potent peptide, followed by NKA (30%), NKB (20%), and SP (1%). Thus, endogenous tachykinins together with other active substances may contribute to the tone and motility, including micturition reflex, of the urinary bladder, the ureters, and the urethra, but the part actually displayed by the tachykinins remains to be established.

E. Immune System

The influence of the tachykinins on the immune system has been carefully reviewed in recent articles (Hartung and Toyka, 1989; McGillis et al., 1990; Eglezos et al., 1991; Maggi, 1997). Although there is increasing evidence that the tachykinins (especially SP and, subordinately NKA) play a role in neuro-immunomodulation. i.e., in the control and regulation of the immune response by the central and peripheral nervous system, the actual relevance and importance of the tachykinins in immunomodulation is uncertain. Tachykinins are merely one of the numerous factors that may directly or indirectly influence the immune system: all or nearly all peptidergic and aminergic neurotransmitters, the derivatives of arachidonic acid, and all of the many active substances synthesized and released by the immune cells.

Data supporting the influence of the tachykinins on the immune system are as follows: a) Occurrence of SP immunoreactive fibers in organs of the immune system, such as lymph nodes, thymus, and bone narrow. b) Presence of SP receptors in thymus and spleen and, above all, expression of these receptors in human circulating lymphocytes and monocytes, rabbit polymorphonucleates and leukocytes, guinea pig macrophages. c) Clearcut action of SP, in vitro and in vivo, on B- and T-cell proliferation, immunoglobulin secretion, cellular chemotaxis, and lymphocyte migration.

The integrity of the immune system is essential for life. The involvement of tachykinins in the control of the immune system in health is very difficult to be established and, at any rate, it seems to be of limited importance: knockout mice with disrupted preprotachykinin A gene were in good health. However, it is necessary to distinguish between physiology and pathology of the immune system. In pathological conditions, such as inflammatory processes, things are even more complicated, because of the enormous cascade of biochemical events that take place during inflammation and immune reaction. It is certainly possible that certain immune cell types are able to synthesize and release tachykinins (of extra-neuronal source) and that NK1 receptors play a role in mediating extravascular migration of granulocytes into inflamed tissues in response to various stimuli (Maggi, 1997). In human skin, exogenous tachykinins may cause wheal and flare, due to release of histamine, and may evoke plasma leakage through the capillaries in several but not all tissues. SP may also degranulate mast cells, but this is an effect attributable not to the intact SP molecule, but to its N-terminal segment acting on receptors independent from the classical tachykinin receptors. To demonstrate the complexity of participation of SP in inflammation, it has been shown in a recent paper (Wallace et al., 1998) that in a model of acute colitis in the rat and guinea pig, NK1 antagonists, although reducing the infiltration of granulocytes during the first 12 h after induction of colitis, failed after repeated administration during a 3-day period to affect granulocyte recruitment or severity of tissue injury.

F. Central Nervous System

The overall presence of tachykinins with their receptors in the CNS of mammals and in that of all examined submammalian species (see neuronal localization) constitutes the most pregnant and incontrovertible evidence that these peptides play in the CNS a very important role as neurotransmitter/neuromodulatory agents, as demonstrated by neurophysiological evidences directly showing this importance (Otsuka and Yoshioka, 1993).

In the CNS, tachykinins occur in large amounts particularly in areas involved in the central control of several peripheral autonomic functions (blood pressure, respiration, micturition, gastrointestinal motility, etc.), of essential functions (e.g., drinking behavior), of the affective and emotive life (stereotyped behavior, motility, anxiety, aggression, and pain), and of higher cerebral functions (learning and memory).

Blood pressure. Eledoisin (0.1–1 nmol/kg) injected into the cerebral ventricles of anesthetized rats produced a biphasic cardiovascular response that consisted of an initial fall of systemic blood pressure (8-15 mm Hg) followed by a rise (20–22 mm Hg). Abolition of the fall in blood pressure by phentolamine suggests central inhibition of sympathetic tone to vessels that afford peripheral resistance, whereas blockade of the delayed hypertensive phase by propranolol indicates a central activation of cardiac adrenoreceptors (Pearson et al., 1969). Different effects were obtained when 1 µg of eledoisin was intracerebroventricularly administered in conscious rats. The peptide, in fact, produced a longlasting rise in blood pressure (18 mm Hg) that was accompanied by behavioral excitement. Pretreatment with phentolamine, but not propranolol or morphine, prevented the pressor response, thus, indicating an α -adrenergic-mediated vasoconstriction (Lambert and Lang, 1970). Effects produced by intracerebroventricular injection of SP (10 nmol) were similar: increase in blood pressure, heart rate, and sympathetic efferent activity with visceral vasoconstriction and hindlimb vasodilation. The cardiovascular responses were accompanied by a behavioral defense reaction, including increased locomotion, scratching, skin biting, and grooming (Unger et al., 1988). Also NKA (10 nmol) elicited increase in blood pressure and heart rate (via sympathetic activity) (Takano et al., 1990).

Respiration. Intracerebroventricular injections of SP in rats (3–30 nmol) induced a dose-dependent stimulation of minute ventilation due to increase in total volume, although respiratory frequency was slightly reduced (Hedner et al., 1984). Similar respiratory effects were produced by application of SP on the dorsal surface of the medulla oblungata in newborn rabbits (Yamamoto and Lagercrantz, 1985).

Gastric acid secretion. The NK3 receptor preferring tachykinins (kassinin, NKB, and PGKII) given to rats by intracerebroventricular injection (0.01–10 nmol/rat) elicited a dose-related inhibition of gastric acid secretion. Kassinin and eledoisin were the most potent peptides, the other peptides showing the following order of relative potency: kassinin > NKB = PGKII \gg NKA > SP and physalaemin (inactive). Subcutaneous doses up to 20 nmol of eledoisin or kassinin were ineffective (Improta and Broccardo, 1990; Improta et al., 1996).

Gastric emptying. Administration of 0.1 nmol of either eledoisin or kassinin produced a 35 to 40% inhibition, and 10 nmol caused a 100% inhibition of gastric emptying of a liquid meal. The relative potency of other examined tachykinins was as follows: NKA, 50%; physalaemin, NKB, and PGKII 0.3%, thus suggesting a predominant involvement of NK2 receptors (Improta and Broccardo, 1990; Improta et al., 1996). In general, tachykinins are less potent in their inhibition of gastric emptying and gastric secretion than either bombesin or opioid peptides.

Colonic propulsion. Intracerebroventricular injections of PG-KII (0.1–100 ng/rat; threshold 1 ng/rat), a selective NK3 receptor agonist, produced a dose-related inhibition of colonic propulsion in the rat. Senktide had a weaker, but evident action, whereas NKB, the classical mammalian selective NK3 agonist was inactive, up to 10 μ g/rat (Broccardo et al., 1999). The interpretation of this surprising result is obscure. It is tempting to suggest the existence of different NK3 receptor subtypes.

Food intake. Intracerebroventricular injections of eledoisin and physalaemin (100–1000 pmol) did not reduce intake of milk or solid food by rats. Some inhibition of milk intake, observed at 100 ng doses, was accompanied by increased grooming and locomotion (Massi et al., 1986).

Thermoregulation. At doses up to 10 nmol, intracerebroventricular administration of tachykinins (kassinin, eledoisin, physalaemin, and SP) had no effect on the temperature of rats kept at room temperature (Broccardo and Improta, 1988).

Sexual behavior. In ovariectomized, estrogentreated female rats, bilateral injection of SP (50–1000 pmol) into the midbrain gray matter produced a rapid, long-lasting (3-h) increase in lordosis score, similar to that produced by luteinizing hormone-releasing hor-

mone (Dornan et al., 1987). Similarly, injections of SP (10–200 pmol) into the medial-preoptic-anterior-hypothalamic area in rats significantly shortened the interval to initiate copulation and reduced ejaculation latency (Dornan and Malsbury, 1989).

Drinking behavior. The effects of tachykinins on all aspects of drinking behavior were 1000 times less intense by peripheral than by central administration.

Intracerebroventricular pulse administration of eledoisin (threshold, 10 pmol/rat) potently inhibited water intake evoked by intracerebroventricular angiotensin II (100 pmol/rat), water deprivation, and cell dehydration. SP and physalaemin were far less potent; kassinin and NKA caused only a long-lasting inhibition of drinking due to cell dehydration. Brain areas sensitive to the antidipsogenic effect of eledoisin versus angiotensin II-induced drinking are the nucleus preopticus medialis, the nucleus anterior hypothalami, and the subfornical organ (Massi et al., 1988, 1990).

Eledoisin, kassinin, and, to a lesser extent, physalaemin caused release of vasopressin, with ensuing antidiuresis. SP was ineffective. Vasopressin release (particularly evident upon injection of the peptide into the hypothalamic paraventricular nucleus) seems to be mediated by central angiotensin, once NK3 receptors are activated (Polidori et al., 1989; Massi et al., 1991).

Kassinin, eledoisin, and, to a lesser extent, NKA (100 nmol/rat, intracerebroventricularly) displayed a potent and long-lasting inhibitory effect on salt intake. SP, physalaemin, and neurokinin B were far less effective, and thus, did not suggest NK1 receptor involvement, because SP was only poorly effective. The medial region of the amygdala seems to be main site of action of the tachykinins for inhibition of salt intake (Massi and Epstein, 1989; Massi et al., 1990).

The effects of the tachykinins on drinking behavior in other mammalian species (rabbit and sheep) were considerably less intense and less constant. However, in cats, intracerebroventricular injections of eledoisin (100 pmol) caused a remarkable (60%) and long-lasting (over 60 min) inhibition of angiotensin-induced drinking. Eledoisin was at least four times more potent than SP. Kassinin appeared virtually inactive (Barocelli et al., 1988).

In sharp contrast to the rat, tachykinins displayed a potent dipsogenic effect in the pigeon and, less evident, in the duck (De Caro et al., 1978, 1980). Physalaemin stimulated water intake even at intracerebroventricular doses as low as 10 pmol/pigeon. At the highest tested doses (1 nmol), the animal drank more water within a few minutes than would normally be consumed during a period of 16 to 24 h. The dipsogenic potency of physalaemin was 10 times less than that of angiotensin II, similar to that of eledoisin and kassinin, but 10 times greater than that displayed by NKA, and 100 times greater than that possessed by NKB. This order of potency does not seem consistent with the tachykinin re-

ceptor subtypes so far proposed. The dipsogenic effect of the tachykinins cannot be attributed to activation of angiotensin II receptors, because drinking was not reduced by administration of angiotensin II antagonists (De Caro et al., 1978, 1988; Massi et al., 1987).

The selective agonists of NK3 receptors (NKB, senktide, and PG-KII) potently inhibited ethanol intake in genetically alcohol-preferring rats; at intracerebroventricular doses from 10 pmol/rat PG-KII was 3 times more potent than senktide. At doses of 100 pmol/rat, only alcohol intake was inhibited in food-deprived rats, not food intake or prandial drinking, indicating that the effect on alcohol intake was behaviorally selective. PG-KII, a NK3 receptor tachykinin agonist, inhibited angiotensin II-induced drinking only at doses of 300 to 1000 pmol, producing also evident competitive behavior, locomotion, and inhibition of digestive behavior (Ciccocioppo et al., 1997).

Micturition reflex. Intracerebroventricular injection of SP (30 nmol) or capsaicin (25 μ g) elicited the micturition reflex in the rat, probably by acting directly on the brain micturitian centers (Dib et al., 1998).

Stereotyped and motor behavior. At intracerebroventricular doses of 0.6 nmol, SP(1–7) inhibited not only nociception but also aggressive and grooming behavior, while stimulating, like SP, investigative motor behavior. The C-terminal peptide fragment [pGlu⁶] SP(7–11) exerted opposite effects (Hall and Stewart, 1984). In producing a rigorous reciprocal hindlimb scratching accompanied by extensive grooming behavior, there was an impressive (approximately 1000 times) difference in responsiveness to intracerebroventricular injection of SP as a function of the genetic strain and age of mice, the old animals (4–5 months) being less sensitive than the young (1–2 months) animals (Hall et al., 1985).

Moreover, whereas the intracerebroventricular injection of all tested tachykinins (SP, physalaemin, NKA, eledoisin, and kassinin) produced in mice an enhancement of grooming and scratching behavior and a reduction of sniffing behavior, only SP increased hindlimb rearing behavior. This effect, unique to SP, was shared by the N-terminal metabolic fragment SP(1–7) and, very surprisingly, also by SP(1–6) (Hall et al., 1987).

The intracerebroventricular, but not intravenous, injection in gerbils of the two SP-like, NK1 receptor agonists, [Sar⁹,MetO₂¹¹]SP or DAla-[Pro⁹,Leu¹⁰]SP, and GR 73632, elicited in gerbils a characteristic repetitive hind paw tapping, which was not associated with an increase in locomotor activity and seemed to be involuntary. Peak response occurred within 5 to 10 min. GR 73632 was 70 times more potent than [Sar⁹,MetO₂¹¹]SP (ED₅₀ 0.7 nmol/gerbil), but the response induced by GR 73632 was less intense. Responses were significantly and dose dependently antagonized only by CNS penetrating NK1 receptor antagonists (Bristow and Young, 1994; Rupniak and Williams, 1994).

At intracerebroventricular doses of 100 to 400 pmol, the NK1 agonist GR 73632 significantly increased also in the guinea pig the locomotor activity. The effect was abolished by NK1 receptor antagonists and by haloperidol (Mason et al., 1992).

Results obtained by subcutaneous or intraperitoneal injection of SP (5 nmol) in mice were at variance. In fact, the peptide decreased spontaneous locomotor activity and counteracted amphetamine-induced hyperactivity. Spontaneous exploratory behavior was also lowered. It is possible that brain monoamines are implicated in these effects, with acceleration of dopamine turnover and retardation of serotonin turnover. Smaller doses of SP showed an antinociceptive morphine-like action in the hot plate test. These results are consistent with SP having a tranquilizing action in mice (Starr et al., 1978).

Aggressive behavior. The intracerebroventricular injection of 0.6 nmol/kg SP or of the N-terminal fragment SP(1–7) reduced fighting in mice made aggressive by prolonged isolation. This effect was enhanced by naloxone. In contrast, the shorter C-terminal analog of SP, [pGlu⁶]SP(7–11) increased the isolation-induced fighting, an effect that was antagonized by naloxone, demonstrating that the various peptide fragments of the SP molecule can exert opposite effects on a specific behavior and that the different effects of naloxone may be modulated by specific mechanisms (Hall and Stewart, 1984; Hall et al., 1987).

Learning and memory. SP administered subcutaneously influenced dose-dependently passive and active avoidance conditioning in mice. The retention of a single trial passive avoidance task was enhanced by 0.75 pmol/g. Higher or lower doses were less active or ineffective. SP did not alter the rate at which the mice learned an active avoidance task but increased the extinction of learning (Schlesinger et al., 1983). Similarly, in an appetite motivated learning task, mice injected subcutaneously with 0.75 pmol/g SP retained the task better than control animals, suggesting that SP-treated remembered original better $_{
m the}$ (Schlesinger et al., 1986). These results were confirmed by Hasenohrl et al. (1990), who found that enhancement of inhibitory avoidance learning produced in rats by SP (40 nmol/kg) was reproduced by the N-terminal fragment SP(1-7), but not by the C-terminal fragment [pGlu⁶]SP(6-11). Higher or lower doses of SP had no effect, demonstrating that the facilitating effect of the peptide was reflected by an U-shape dose-response function.

Rats given diazepam 20 min before the training on an inhibitory avoidance task showed an impaired retention. The amnesic effect of diazepam was blocked by 50 μ g/kg SP and 167 μ g/kg SP(1–7) but not by 134 μ g/kg SP(6–11). Thus, the amino acid sequence responsible for this effect may be encoded by the N-terminal fragment of SP (Costa and Tomaz, 1998).

Psychological stress: anxiety. The intracerebroventricular infusion of the NK1 agonist GR 73632 (0.1 nmol) in guinea pigs causes not only motor activation but also pronounced and long-lasting audible vocalization, markedly attenuated not only by the antidepressant drugs, but also by the NK1 receptor antagonist L 733.061. Similarly, the CNS-penetrant NK1 receptor antagonists, like the antidepressant and anxiolytic drugs, were able to inhibit vocalization evoked in guinea pig pups by transient maternal separation.

It is concluded that the selective pharmacological blockade of SP receptors is capable of inhibiting behavioral responses to psychological stress in a manner resembling the effect of clinically used psychotherapeutic agents (Kramer et al., 1998). However, no influence on anxiety (upon field assay) was seen in mice with disrupted gene of NK1 receptor (De Felipe et al., 1998).

Abstinence reaction during opioid withdrawal. It has been found that SP may modulate the abstinence reaction to opioid withdrawal. In fact the N-terminal fragment of SP, SP(1–7), may inhibit the intensity of the withdrawal reaction in morphine-dependent rats. Moreover, significant increases in concentration of SP(1–7) were observed in different brain areas during morphine withdrawal, indicating the involvement of the SP system during opioid withdrawal (Zhou et al., 1998).

Action on discrete selected brain areas. Cat subfornical organ. Neither physalaemin nor eledoisin produced activation of neurons in the cat subfornical organ upon direct application onto its surface (Felix, 1967).

Rat cerebral cortex. Iontophoretic application of SP, physalaemin, and eledoisin in the cerebral cortex of rats excited 91% of the spontaneously active neurons tested, including b-cells. Of the tachykinins, 0.8 μ M SP was the most potent, immediately followed by physalaemin and, at distance, by eledoisin. No excitation was induced in nonspontaneously active neurons (Phillis and Limacher, 1974).

Rat nigro-striatal system. The bilateral infusion of SP (2.25 nmol) into the substantia nigra produced a strong increase in stereotyped rearing and sniffing, with no concurrent enhancement of locomotion. After successive infusions, rearing response disappeared. In caudate-lesioned rats, behavioral stimulation by SP was blocked, suggesting that the response to SP is mediated through the nigro-striatal dopamine system (Kelley and Iversen, 1979).

In accordance with these results Tan and Tsou (1988) found that intranigral injection of kassinin, eledoisin, and SP (0.5 nmol) produced a marked dose-dependent increase of 3,4-dihydroxyphenylacetic acid and dopamine concentration in the ipsilateral striatum and a number of controlateral circlings. The rank order of activity was kassinin > eledoisin > SP.

SP(1-9) and SP(6-11) at 0.1 to 1 nM concentrations induced an increase in dopamine outflow from rat striatal slices. The effect of SP(6-11) was blocked by an

NK1 antagonist, whereas the effect of SP(1–9) was unaffected. The coincubation of the above fragments with intact SP revealed a negative interaction between fragments and substance P (Khan et al., 1998).

In fresh striatal slices, (3 H)SP bound specifically to one site from which it was displaced by SP, but not by SP(1–7) or SP(5–11). In contrast, 10 μ M SP(1–7) or SP(5–11) induced, like SP, a significant internalization of the NK1 receptor. It is suggested that SP fragments have high affinity with an NK1 receptor conformance, which is different from that labeled by (3 H)SP (Michael-Titus et al., 1999).

Hippocampus and entorhinal cortical cortex. Intrahippocampal administration of 10 pmol of SP triggered self-sustained status epilepticus in response to electrical stimulation of the perforant path for periods too brief (7 min) to have any effect in control rats (requiring a 30-min stimulation). The seizures were accompanied by high-amplitude electrographic paroxysmal activity that lasted many hours. Hippocampal damage resembling that known to occur in human epileptic seizures was blocked by SP receptor antagonists (RP 67.580).

In addition, in the status epilepticus, a rapid, dramatic increase of the expression of preprotachykinin A mRNA, of SP in several brain areas, and of the glutamate release from hippocampal slices has been shown. It is concluded that enhanced expression of SP during self-sustained status epilepticus may modulate hippocampal excitability and play a critical role in the maintenance of status epilepticus (Liu et al., 1999a). In hippocampal dentate gyrus granule cells, SP produces a robust enhancement on *N*-methyl aspartate channel function, prolonging the opening of the channels. Thus, SP provokes an enhancement of the excitatory amino acid-mediated excitability (Lieberman and Mody, 1998).

In experiments in vitro on slices of rat entorhinal cortical cortex, it was found that spontaneous epileptiform discharges evoked by the GABA receptor antagonist bicuculline were reduced in frequency and sometimes in duration by SP. Excitatory synaptic potentials mediated by aspartate were not affected by the peptide (Maubach et al., 1998).

Rat ventral tegumental area. SP and related tachykinins administered either intracerebroventricularly (0.1–20 μ g) or directly into the ventral tegumental area of the rat mesencephalon (0.5 nmol) caused increased locomotor activity, grooming behavior and wetdog shakes. Kassinin, eledoisin, and NKA elicited the greatest locomotor activity and wetdog shakes, whereas SP and physalaemin were not effective in producing full-body grooming (Elliott and Iversen, 1986). Similarly, mice infused with SP (3 nmol) into the ventral tegumental area exhibited a long-lasting increase in spontaneous activity with rearing and sniffing (Kelley et al., 1979).

Rat nucleus tractus solitari. Microinjection of SP (0.7 pmol) or NKA (0.9 pmol) in the nucleus caused prompt, transient hypotension and bradycardia, suggesting that the tachykinins may act as neurotransmitters of the baroreceptors of the nucleus (Nagashima et al., 1989). The same results were obtained with microinjection of either SP or SP(1–7) (60 pmol), and the effects of SP were blocked when cleavage of SP was inhibited by phosphoramidon, which alone failed to block the depressor and bradycardic effect of SP(1–7), suggesting that only the N-terminal fragment was active and not the intact SP molecule (Hall et al., 1989).

Rat nucleus accumbens. SP and [pGlu⁶]SP(7–11) attenuated passive avoidance behavior when at picomolar amounts were injected into the n. accumbens. The N-terminal fragment, SP(1–7) had an opposite effect, facilitating passive avoidance behavior (Gaffori et al., 1984). However, intraaccumbens injection of SP at nanomolar doses had no significant behavioral effects in regard to motility and to conditioned place preference (Schildein et al., 1998).

Rat nucleus basalis magnocellularis. SP (1 pmol) injected into the n. basalis magnocellularis region exerted anxiolytic-like effects in the rat as shown by more time spent on the open arms of the plus-maze test and in social interaction. When administered intraperitoneally, SP had a biphasic dose-dependent effect: anxiolytic action at 40 nmol/kg, and anxiogenic action at 400 nmol/kg (Hasenohrl et al., 1998).

Rat spinal cord. On isolated spinal cord of newborn rats, the tachykinins at 10 to 100 nM concentrations caused depolarization of motoneurones. The rank order of potency of the examined tachykinins was physalaemin > NKB = kassinin = SP > NKA (Matsuto et al., 1984).

Bullfrog spinal cord. SP, physalaemin, and eledoisin exerted a strong excitatory action on motoneurones of isolated bullfrog spinal cord. On a molar basis, SP was about 200 times, physalaemin 1500 times, and eledoisin 2000 times more active than L-glutamate in depolarizing the motoneurones. Because the depolarizing action of substance P and related peptides was blocked by Ca²⁺ deficiency by tetrodotoxin, it is likely that the peptides have a direct action on the motoneurones. Moreover, Konishi and Otsuka (1974) suggested SP to be an excitatory transmitter of primary sensory neurons.

Molluscan ganglia. Direct application of physalaemin on isolated esophageal ganglia of the mollusc $Achatina\ fulica$ produced excitation of an identified tonically autoactive, giant neuron (TAN). By bath application of physalaemin (200 μ g/ml), the frequency of the TAN's spike discharge increased more than twice. Micro-drop application (3.5 ng) resulted in the biopotential of TAN showing a slight hyperpolarization followed by a marked depolarization. The excitatory effect of physalaemin clearly was predominant. No other peptides examined, including SP and eledoisin, had any effect (Takeuchi et

al., 1976). After trypsin or chymotrypsin treatment, physalaemin lost its effect, and a clear inhibitory effect emerged on the same TAN. Among the peptide fragments obtained by chymotryptic digestion, the tripeptide Lys-Phe-Tyr- appeared to be responsible for the strong inhibitory effect on TAN. The critical concentration of the tripeptide by bath application was 6 to 20 μ g/ml, whereas by micro-drop, amounts as low as 0.3–0.5 ng caused a marked inhibition of the TAN biopotential (Takeuchi and Sakai, 1977; Takeuchi et al., 1977a, 1977b).

From the pharmacological data here presented and from other experiments on mutant mice, however, it seems clear that the tachykinins are not essential for life or for most of the above functions and expressions of the CNS activity. Mutant mice had no gross physical abnormalities, were similar in size and weight to wild mice, appeared healthy over a period of at least 6 months, and were fertile with normal litter size and maternal behavior. They also appeared normal on a rotating rod and in open field activity (Cao et al., 1998; De Felipe et al., 1998; Zimmer et al., 1998). Tachykinins are probably a very important link, but only one link, of the extremely complex chain of events underlying the chemical conveyance of information within the CNS. The part played by the tachykinins in the array of transmitters/modulator molecules occurring in the CNS is not yet completely understood. Thus, as pharmacological and physiological data on the role of tachykinins in the CNS are derived essentially from experiments carried out in mice and rats, it is even harder to conceive that they are transferable, sic et simpliciter, to higher mammals and to man.

Moreover, the SP(1–7) fragment seems to be involved not only in analgesia but also in aggressive and grooming behavior (Hall and Stewart, 1984; Hall et al., 1987), aggressive behavior by prolonged isolation (Hall and Stewart, 1984), hindlimb rearing behavior (Hall et al., 1987), learning and memory (Hasenohrl et al., 1998), central elicited hypotension and bradycardia (Hall et al., 1989), diazepam amnesic effect (Costa and Tomaz, 1998), and the abstinence reaction to morphine withdrawal in morphine dependent rats (Zhou et al., 1998).

SP(1–7) may be an endogenous modulator of SP actions in the brain (Herrera-Marschitz et al., 1990). The intranigrally injected fragment acted as a very potent antagonist against responses induced by intranigral injection of SP (dopamine release in the striatum with consequent behavioral effects) and also of physalaemin (Sakurada et al., 1990b). Thus, it seems demonstrated, beyond any doubt, that SP(1–7) may act in the CNS as neurotransmitter/neuromodulator.

All of these findings raise the following questions, with the possibility that the entire problem of the function of tachykinins in the CNS should be revised:

1) To what extent are the central actions of SP attributable to the intact SP molecule and to what extent does

SP require enzymatic cleavage with formation of its metabolite SP(1-7) to become active? SP acts certainly as intact molecule a) in the case that its central effects are reproducible also by other selective NK1 agonists. such for example physalaemin; b) in the case that its action is inhibited by selective NK1 receptor antagonists; and c) in the case that its action is potentiated by enzyme inhibitors, blocking fragmentation of SP. The observation that phosphoramidon (2-2000 pmol) (an endopeptidase inhibitor) and, to a lesser extent, bestatin (an aminopeptidase inhibitor) remarkably increased and prolonged the behavioral responses (scratching, biting, and licking) induced in mice by SP suggests the importance of endopeptidases in terminating the effects of the SP intact molecule (Sakurada et al., 1990a, 1990b). On the contrary, it is probable that only SP(1-7) fragment is active a) when its action is not reproducible by neither SP or other tachykinin agonists: b) when actions of SP(1-7) are not blocked by NK1 antagonists; and c) when the effect of SP is blocked by phosphoramidon, which inhibits formation of SP(1-7). For example, hypotension and bradycardia elicited by injection of SP in the rat nucleus tractus solitarius are blocked by the endopeptidase inhibitors (Hall et al., 1989).

2) Lack of binding of SP(1-7) to any of three classical tachykinin receptors, presupposes the existence of other selective binding sites. Data that, however, need confirmation, have demonstrated the existence of two populations of binding sites in the mouse spinal cord capable of binding reversibly (3H)SP(1-7) (Igwe et al., 1990). Specific agonists for NK1, NK2, and NK3 receptors did not compete at the binding of SP(1-7). These results support the existence of an N-terminal directed SP-receptor. The fact that DAMGO, a μ -opioid agonist, was active in displacing the ligand SP(1-7) is surprising.

The existence of different binding sites for the C-terminal fragment and the N-terminal fragment of SP was indirectly demonstrated also by Khan et al. (1998) who found that the increased dopamine outflow from rat striatal slices produced by SP(6–11) was blocked by a NK1 antagonist, whereas the outflow elicited by SP(1–9) was unaffected.

3) At present, it is not clear which of the amino acid residue(s) in the SP sequence are responsible for the central effects of the heptapeptide. Generally experiments have been carried out with SP(1–7) and SP(1–8), suggesting that the Phe⁷ residue plays an important role, but in some experiments, SP(1–6) also proved to be active. SP-like peptides, sharing as many as five to six amino acid residues with the N-terminal heptapetide of SP, are found in reptile brain and intestine (bufokinin) and in fish brain (trout SP and cod SP). It would be of general interest to check whether these N-terminal fragments (1–7) are active in the CNS of the pertinent species and of mammals.

4) Transgenic mice with the disrupted the gene encoding the NK1 receptor (De Felipe et al., 1998) could be an

excellent material for investigating if SP (1–7) still displays its central effects and, in this case, to demonstrate unequivocally, the existence of receptors activated only by the N-terminal fragment of SP.

G. Pain

Much evidence has accumulated to suggest that SP is synthesized in the periphery by small-diameter sensory "pain fibers" and then, upon intense peripheral stimulation released into the dorsal horn, as a first step, through activation of NK1 receptors of transmission of pain information into the CNS. As a consequence, there is central hyperexcitability and increased sensitivity to pain. However, SP is also largely present, together with its NK1 receptor, in several brain areas. It is beyond question that brain SP contributes to pain perception and elaboration. But how, and to what extent? To answer this fundamental question it seems opportune to discuss separately the problem of SP and pain in the periphery (until the dorsal horns) and in the brain.

Periphery. The first association of SP with pain was made by Lembeck and Holzer (1979), who suggested that SP, together with other neuropeptides, may be released from the peripheral sensory nerve fibers in the skin, muscle, and joints. This release was thought to be involved in "neurogenic inflammation", a local painful inflammatory response to certain types of injury or infection, such as that caused by the classical irritant capsaicin.

By intraperitoneal administration, SP (0.8–3.2 nmol/mouse) displayed either no analgesic effect (Growcott and Shaw, 1979) or predominantly a clear antinociceptive action. SP antinociception was found at 10 to 20 pmol in the mouse (Stewart et al., 1976), at 0.8 to 3.2 nmol in the mouse (Starr et al., 1978), and at 0.2 to 0.8 nmol/kg in the rat (Mohrland and Gebhart, 1979).

SP injected into the lumbar subarachnoidal space of rats depressed the tail-flick response in a dose-dependent manner (ED_{50} 1.2 nmol/rat). Maximum effect was reached after 20 min and lasted 30 min. The antinociceptice effect of SP was abolished by naloxone (Doi and Jurna, 1981).

At a dose of 7.5 nmol, SP depressed the motor response evoked by supramaximal stimulation of the sural nerve and also reduced the activity of part of the ascending neurons of the spinal cord evoked by stimulation of C-afferent fibers. The depressive effect in ascending nociceptive activity was slow in onset, lasted longer than 60 min, and was abolished by naloxone (Doi and Jurna, 1982). In the superfused spinal cord of rats and cats, iontophoretic application of SP produced a long-lasting excitation of the dorsal horn neurons similar to that elicited by noxious cutaneous stimuli; SP was released from dorsal horns after stimulation of sensory neurons by capsaicin, and this release was completely inhibited by morphine (Yaksh et al., 1980).

Intrathecal injection of SP or NKA (10–100 pmol), which in the mouse caused a dose-dependent reciprocal hindlimb scratching, licking, and biting response directed to the caudal part of the body, also decreased latency in the tail-flick assay but did not alter reaction in the hot plate test. These effects are interpreted as indicative of a nociceptive behavior (Hylden and Wilcox, 1981; Seybold et al., 1982; Gamse and Saria, 1986).

SP, physalaemin, and eledoisin intrathecally injected in rat have been reported to cause hyperalgesia in the tail-flick test. Hyperalgesia produced by the tachykinins was dose-dependent, was maximal 10 to 20 min after injection, and lasted 30 min. The rank order of potency was: physalaemin > SP > eledoisin. Desensitation to the effects of the peptides was observed after three successive injections of the peptide (Moochhala and Sawynok, 1984).

After microdyalisis of SP or NKA into the dorsal horn of anesthetized monkeys, it was observed that neither peptide had significant effects on the background activity or the response to mechanical or thermal stimulation of the skin. However, each peptide produced significant increases in the response to simultaneous or subsequent iontophoretic application of excitatory amino acids (glutamic acid). Thus, it seems that tachykinins facilitate responses of dorsal horn neurons to excitatory amino acids or to cutaneous stimuli (Dougherty et al., 1995).

Similarly, the progressive hypersensitivity of spinal flexor motoneurons induced by repeated peripheral stimulation of inflamed tissues in decerebrated rats was attenuated by the subcutaneous injection of the NK1 antagonist RP 67580, indicating that SP is involved in mediating progressive hypersensitivity during inflammation (Ma and Woolf, 1997).

Recent, decisive demonstration of the important role of SP in nociception has been afforded by experiments with nociceptin and by experiments on mutant mice in which either the preprotachykinin A gene or the gene encoding the NK1 receptor was disrupted. Inoue et al. (1998) demonstrated that the nociceptin/orphanin FQ-induced nociceptive response is brought about in mice by SP release from peripheral nerve endings of nociceptive primary afferent neurons. After intraplantar injection into the hindlimb of mice of nociceptin ($EC_{50} = 0.31$ fmol), there was a 25 to 70% increase in the flexor-reflex response, which was abolished by pretreatment of mice with an NK1 tachykinin receptor antagonist or with the SP-depleting agent capsaicin, but not by pretreatment with NK2 antagonists. Similarly, nociceptin was completely ineffective in mice with targeted disruption of the NK1 receptor gene.

It has been demonstrated that the knockout mice, which presented a disruption of the gene encoding the NK1 receptor (with consequent blockade of the activity of SP but not of NKA or SP(1–7) (De Felipe et al., 1998), and the mutant mice, which presented a disruption of the preprotachykinin A gene (with consequent lack of expression of SP, SP(1–7), and NKA) (Cao et al., 1998; Zimmer et al., 1998), did not show any changes to acute

pain threshold in mechanical, electrical, chemical, or thermal nociceptive tests, but their responses were blunted in tests that involved more intense noxious stimuli. The importance of SP/NKA seems to apply only to a certain "window" of pain intensity, and when the intensity of the pain stimulation was further increased, the response of the knockout mice did not differ from those of wild mice. The fact that behaviorally acute nociceptive threshold (tail-flick and hot plate assays) were not affected by gene disruption would imply lack of any activation of NK1 receptor and of any involvement of SP in the above assays.

In contrast, when sensory nerves are subjected to an intense period of noxious stimulation, normal animals show a "wind up phenomenon", i.e., an amplification and intensity coding of nociceptive reflexes, which indicate a sensitization of the CNS mechanisms by intense stimulation. The "wind up" was completely absent in the NK1 receptor lacking mice. Thus, SP seems to play an unexpected role for full development of stress-induced analgesia and also for the aggressive response to territorial challenge. Mutant mice did not present any change in anxiety tests. However, the fact that aggression but not anxiety (open field assay) was blunted in mutant mice would again indicate that NK1 receptors and SP are not involved in anxiety even if, in another anxiety assay (vocalization in guinea pig pups by transient maternal separation), the NK1 receptor agonists increased vocalization and the NK1 antagonists remarkably attenuated this effect (Kramer et al., 1998).

In agreement with the above data, Zimmer et al. (1998) observed that knockout mice displayed no significant pain responses after formalin injection, but have an increased pain threshold in the hot plate test. In addition, the mutant mice reacted normally in the tail-flick test assay and acetic acid-induced writhing test.

The conclusion is that mutant mice develop hypoalgesia in some assays, but not in others, probably depending on the apparent levels (spinal or supraspinal) in which the involved pain mechanisms are situated. We further suggest that it is possible that enkephalins and SP modulate nociceptive inputs antagonistically and determine whether a nociceptive stimulus is experienced as pain.

Brain. Results obtained by intracerebroventricular injection of SP and other tachykinins, on pain sensation are very complex, conflicting, and open to unexpected speculations.

The intracerebroventricular injection of 2 to 2000 pmol of SP did not affect the hot plate test in mice (Hayes and Tyres, 1979). Similarly, intrathecal SP (10–10000 pmol) in rats did not significantly affect pain threshold in various analgesic tests (paw pressure, tail immersion, and hot plate test). Moreover, Malthe-Sphirenssen et al. (1978) found that neither intracerebroventricular injection nor injection into the periaqueductal gray of high doses of SP (30 nmol) induced analgesia in

rats. Frederickson et al. (1978), in turn, showed that SP produced analgesia in mice when administered in very small doses (1.25–5 pmol/mouse) by intraventricular route. The analgesic effect was blocked by naloxone. At doses greater than 50 pmol, this effect was lost and hyperalgesia appeared when these doses were combined with naloxone, analgesia when combined with baclofen. Thus, SP may have a dual action in brain, releasing endorphin at very low doses and directly exciting neuronal activity in nociceptive pathways at higher doses.

All of these results are, however, in conflict with a considerable amount of observations demonstrating that intracerebroventricular SP or SP injected into discrete brain areas is predominantly an analgesic, pain-blunting substance.

Malick and Goldstein (1978) found that, after injection into the periaqueductal gray, SP (EC₅₀ = 0.7 nmol/rat) displayed a long-lasting (30–60 min) analgesic effect in the tail-flick test. SP was approximately 25 times more potent than morphine, and its effect was significantly antagonized by naloxone. Similarly, low doses of SP (10 pmol) applied to the subarachnoid space of the rat potentiated morphine analgesia (0.1 to 0.5 μ g) in the rat tail-flick test, either by facilitating release of endogenous opioids or by modulating opioid receptors.

Stewart et al. (1976, 1982) also demonstrated that centrally administered SP displayed a clear-cut analgesic action. A first important observation was that SP antinociception appeared after a lag of approximately 30 min, even after intracerebroventricular injection, suggesting that the peptide may first require, to become active, an enzymic cleavage at the Phe⁷-Phe⁸ bond, leading to release of the N-terminal fragment SP(1-7) (Hall et al., 1989). This fragment displayed a clear-cut antinociceptive action in the hot plate test, either by intracerebroventricular injection (5 pmol/mouse) or intraperitoneal injection (15-20 pmol/ mouse). SP(1-6) and SP(1-4) did not show any significant analgesic effect. Prior treatment with naloxone abolished the effect of SP(1-7). In comparison with intact SP, the SP(1–7) fragment displayed its antinociceptive effect only within a narrow dose range and as expected, had a shorter lag in onset and a shorter duration of action.

Recently, the effects of two amphibian tachykinins, the NK1 receptor agonist PG-SPI and the NK3 receptor agonist PG-KII, and the mammalian tachykinins SP, NKA, and NKB on the reaction time to a painful radiant heat stimulus (tail-flick test in rats) after intracerebroventricular injection were investigated and compared (Improta and Broccardo, 2000). PG-SPI and PG-KII (1, 5 and 10 μ g) and SP (10 μ g) significantly increased the reaction time, whereas NKA and NKB did not. Like analgesia evoked by exogenous SP, PG-SPI-evoked analgesia was blocked by pretreatment with naloxone. Naloxone left PG-KII antinociception unchanged, but the NK3 receptor selective antagonist markedly reduced it. All of these findings suggest NK1 and NK3 tachyki-

nin receptor system involvement in supraspinal analgesia in rats.

We have discussed in some details this topic on pain because of its great interest in pharmacology, pathology, and therapeutics even if the question "SP equals pain substance?" by Iversen (1998) is still open and has no definite answer, depending on pain intensity (windows!), nature of pain, and methods used to assess response to painful stimuli.

There is evidence that SP plays a role in transmission of pain sensation and its elaboration in the CNS. Evidence is more convincing in the periphery, from sensory nerve endings to the dorsal horns of the spinal cord (SP = pain substance), less so in the CNS, because of a large number of conflicting results and on the still not clear involvement of SP(1-7).

At any rate, in pain control, there is certainly a close interplay between opioid peptides and SP with the concomitant participation of excitatory and inhibitory amino acids, monoamines and other neuropeptides as well. In human beings, the problem of pain is further complicated by its heavy emotional component. The involvement of SP in defense against stress conditions (anxiety and aggression) is highly probable, but again the importance of this involvement remains to be established. It seems that the monoaminergic system plays here a predominant role and that SP or SP(1–7), like other neuropeptides, displays a modulating effect. Again, results obtained in rats and mice are transferable to human beings with caution.

H. Neurogenic Inflammation

Electrical, mechanical, and chemical stimulation of the C-fibers in sensory neurons causes an axon reflex taking place in the branchings of sensory nerves. The consequence is the neurogenic inflammation: pain, vasodilation (flare), and plasma extravasation.

Antidromic vasodilation is mediated by a neurotransmitter at the sensory nerve endings in the skin. Similarly, plasma extravasation elicited by antidromic stimulation also seemed to be provoked by a mediator released from pain sensitive nerve terminals (Jancso et al., 1967).

Among the many transmitters suggested in this connection were acetylcholine, noradrenaline, ATP, bradykinin, histamine, 5-HT, and prostaglandins. At the present time, SP fulfills the criteria for being accepted as the main mediator for all components of antidromic stimulation (Lembeck and Holzer, 1979; Pernow, 1985).

- i. SP is present in the C-fibers of the sensory neurons and is released from these fibers during antidromic stimulation.
- Close arterial administration of SP causes vasodilation and plasma extravasation, thus, mimicking the effect of antidromic stimulation.

iii. Capsaicin, which depletes SP in sensory neurons, almost completely blocks vasodilation and neurogenic plasma extravasation.

The above criteria were completed and remarkably strengthened by more recent data:

- iv. The nociceptin/orphanin-induced nociceptive response is brought about in mice by SP release from peripheral endings of nociceptive primary afferent neurons (Inoue et al., 1998), supporting the view that also pain in neurogenic inflammation is due to release of SP.
- v. In mutant mice with disrupted preprotachykinin A gene, neurogenic inflammation produced by topical application of capsaicin was almost absent, whereas in non-neurogenic paw edema produced by complete Freund's adjuvant neurogenic inflammation was the same in wild-type and mutant mice (Cao et al., 1998). However, there is some doubt about the fact that SP is the unique direct or indirect (through release of histamine from the mast cells) agent responsible for the vasodilation and plasma extravasation seen in neurogenic inflammation. Two points deserve attention. The first is that SP is costored and coreleased from sensory nerve endings with calcitonin gene-related peptide, which displays a potent edema producing activity; the second is that the histamine-releasing activity of SP, which remarkably contributes to plasma extravasation and edema, has been attributed not to the intact SP molecule but to its N-terminal fragment (1-7). Moreover there are data, which need confirmation, showing that antidromic stimulation may not always release SP but other active agents.

In summing up, there is little doubt that neurogenic inflammation represents the most striking and credible example of a decisive, if not unique, involvement of SP in a physiopathological process.

I. Miscellaneous Pharmacological Actions

1. Lachrymal Secretion. Physalaemin given intravenously and at threshold doses of 0.03 to 0.3 nmol/kg was a potent stimulant of lachrymal secretion in the dog. At a dose of 1 nmol/kg, which caused an intense drop of blood pressure, the increase in secretion was 400%.

Like those of the dog, the lachrymal glands of the gerbil seem to be very sensitive to SP. In fact, in this animal, the non-natural NK1 agonist, [Sar⁹,MetO₂¹¹]SP, induced by intravenous injection of doses over 0.007 nmol/gerbil an immediate dose-dependent chromodachryorrhoea, which was blocked by the NK1 antagonist, CP 99,994. Doses of the agonist 180 times higher were required to elicit the same effect by intracerebroventricular injection, demonstrating that the point of attack of the peptide was on peripheral NK1 receptors. The same results were obtained

with intravenous injection (0.3 nmol/gerbil) of another synthetic SP-like peptide, GR 73632 (D-Ala[Pro⁹,MeLeu¹⁰]SP). There was no involvement of the cholinergic system. NKA and NKB receptor agonists were ineffective (Bristow and Young, 1994).

The rat was much less sensitive (intravenous threshold 2.5–5 nmol/kg) and the maximum increase in lachrymal secretion never exceeded 100% (De Caro and Cordella, 1965; Bertaccini et al., 1966).

Lachrymal glands of the rabbit seem to be rather insensitive to tachykinins. In fact, eledoisin, given by close intra-arterial injection, at doses up to 1 nmol, failed to modify fluid and protein secretion (Dartt et al., 1988). In humans, given by eye drops, 10 to 20 nmol of physalaemin provoked a 95% increase in lachrymal secretion, with intense conjunctival hyperaemia and sometimes-moderate chemosis (De Caro and Cordella, 1965). The same amount of eledoisin was ineffective in normal human subjects but showed striking stimulatory effects in patients suffering from hypofunctional lachrymal glands. Lachrymal secretion increased up to 200% and the effect lasted for several hours during which it decreased slowly (Impicciatore et al., 1973).

2. Histamine Release. After perfusion of the rat isolated hindlimb with 10 nmol/min of SP, kassinin, eledoisin, and NKA, it was shown that only SP produced a significant increase in the histamine concentration in the perfusion liquid, from 263 to 750 ng histamine/min. The three other tachykinins were inactive (Erjavec et al., 1981; Holzer-Petsche et al., 1985). Similarly, only SP released histamine from peritoneal mast cells, whereas eledoisin and kassinin were ineffective (Erjavec et al., 1981; Pietrowski et al., 1984). In their ability to release histamine from the rat peritoneal mast cells neurotensin, kallidin and SP were the most potent agonists. Surprisingly an undecapeptide SP antagonist behaved as superagonist.

Only compounds with positive charges at their N-terminals caused a noncytotoxic release of histamine from rat mast cells. It is evident that SP acts by a nonspecific mechanism not related to activation of mast cell NK1 receptor and, hence, not related to the tachy-kinin nature of SP (Devillier et al., 1985, 1989).

A histamine release by 0.1 to 10 μ M SP, with consequent flare, wheal, and itching, has been also demonstrated in human skin, where the peptide was 100 times more potent as histamine liberator than in the rat peritoneal cells. The skin reactions were blocked by antihistaminic drugs (Hagermark et al., 1978). In more detail, in human skin (volar surface of the forearm), 6.25 to 25 pmol of SP produced a dose-dependent flare and wheal response. Only peptides having one or more basic residues at their N-terminal region were effective in producing flare: eledoisin and SP(1–7) were 17 and 7 times less active than SP. Wheal production, on the contrary, was not dependent on basic residues: physalaemin was the most potent agent, SP was one-half as potent, and ele-

doisin was 20 times less potent. Pretreatment with a histamine antagonist reduced both flare and wheal responses; pretreatment with capsaicin reduced the flare but not the wheal response, indicating that in response to the tachykinins, both mast cells and SP-containing primary neurons are involved (Foreman et al., 1983). It has been suggested that the histamine-releasing property of SP derives essentially from cooperation between the basic N-terminal tetrapeptide and the C-terminal heptapeptide with the two successive Phe-residues at position 7 and 8 (Mazurek et al., 1981).

To support the validity of this hypothesis, it would be interesting to check the histamine-releasing activity of carassin(12–21) with three basic residues in the N-terminal tetrapeptide, but with the sequence Phe-Val, instead of Phe-Phe and of bufokinin, with 2 basic and 1 acid residue in the N-terminal pentapeptide and the sequence Phe-Tyr, instead of Phe-Phe.

VI. Tachykinins in Human Diseases and Therapeutics

Research on this topic is rather scant and at early stage of advancement. Pharmaceutical companies are highly interested in diseases possibly attributable, at least in part, to excess or deficiency in tachykinin production and/or release. However, because no tachykinin agonists are hitherto known to possess an appreciable capacity to cross the blood-brain barrier, the focus of interest lies, at present time, on the tachykinin antagonists, which are generally of nonpeptide nature and are often brain-penetrating molecules.

A. Tachykinin Receptor Agonists

In patients suffering from arteriosclerosis obliterans of the legs, the effects of eledoisin (15–35 nmol) injected into the femoral artery have been studied (Szam et al., 1966). The angiorheogram and the pulse volume increased considerably in the majority of patients. Fleeting side effects and slight decrease in blood pressure were also observed. Unfortunately this promising clinical trial was not extended: inconveniences of intraarterial infusion discouraged continuation of experiments. Administration, by eye drops, of eledoisin or physalaemin (10-50 nmol, 1-4 times daily) increased lachrymal secretion and ameliorated the Sjögren syndrome and other forms of keratoconjunctivitis sicca due to deficit of lachrymal secretion (De Caro et al., 1969; Jaeger et al., 1985; Jaeger, 1988). Because of the relative rarity of the disease and the existence of other therapeutical approaches, these "orphan" drugs did not further arouse the interest of the pharmaceutical companies. However, in an organ culture of rabbit cornea, it was observed that SP alone (not NKA, NKB, eledoisin, kassinin, or physalaemin) at any concentration (50 ng/ml-50 µg/ml) did not affect epithelial migration but enhanced the stimulant effect (Nakamura et al., 1997). In the search for the SP fragment responsible for the above effects, it has been found that both SP and its C-terminal tetrapeptide, SP(8–11), acted synergistically with insulin-like growth factor 1 on wound healing of rabbit cornea (Nakamura et al., 1999). There was both stimulations of epithelial migration in vitro and of attachment of corneal epithelial cells to a fibronectin matrix. Moreover, the combination of insulin-like growth factor 1, SP(8–11), and integrins by topical application facilitated wound closure in vivo.

As far as it concerns the neurodegenerative and other CNS disorders, it has been suggested that tachykinins may have both neuroprotective and neurodegenerative properties (Raffa, 1998). Among the degenerative diseases of the CNS, in which a deficit of tachykinins is clearly evident, is Huntington's disease or Chorea. In this autosomal hereditary disorder, a marked decrease in immunoreactive SP fiber density in the regions showing the greatest histopathological destruction, particularly in the substantia nigra, has been reported. Of course, whereas SP has nothing to do with the etiology of Huntington's disease, it could be responsible, at least in part, for the symptoms of this disease: choreiform movements, personality disturbances, and cognitive decrease to dementia.

The intervention of tachykinins in other CNS disorders is more controversial: amylotrophic lateral sclerosis, Parkinson's disease (decrease of SP in substantia nigra), schizophrenia (no significant change in SP content in brain areas), and depression (predominant data showing elevated levels of SP).

The problem of how tachykinins participate in the cerebral aging is also a matter of investigation and debate. Here, only a few studies dealing with the relationship between β -amyloid protein and the tachykinins are reviewed. β-Amyloid is a 39- to 43-amino acid polypeptide that is the primary constituent of senile plagues and cerebrovascular deposits in Alzheimer's disease and Down syndrome. Although the protein has been characterized biochemically, neither its primary biological significance nor its role in the pathogenesis of Alzheimer's disease is completely known. It has been shown that, in cultures of rat embryonic hippocampal cells, the β -amyloid protein is neurotrophic in undifferentiated cells and at low concentrations, but it is neurotoxic in mature neurons and at higher concentrations (Yankner et al., 1990). Neurotrophic and neurotoxic effects of the protein were mediated by its fragment 25 to 35, which shows important homologies to the sequences of SP and other tachykinins. However, the problem on the possible involvement of tachykinins (namely SP) in the pathogenesis of Alzheimer's disease cannot be considered solved. In fact, Zhao et al. (1993) found that amyloid β -protein (1–40) was toxic to NB41A3 neuroblastoma cells in serum-free culture, as judged by decreasing cell number and release of lactic dehydrogenase, and that this toxicity was inhibited by the concurrent treatment of the cells with 1 μM physalaemin. In turn, Kimura and Schubert

(1993) observed that amyloid β -protein (1–40) weakly activated, for itself, the tachykinin receptors, but that in the presence of glutamate, the amyloid β -protein produced an activation of both the tachykinin receptors (especially the NK1 receptors) and the phosphatidylinositol turnover. There is the possibility that an overproduction of amyloid β -protein disturbs normal neuronal transmission by activating the SP receptor in synergy with glutamate or by acting as a SP antagonist by itself. The resulting compromised synapses could lead to the dementia of the Alzheimer's disease.

B. Tachykinin Receptor Antagonists

Based on the knowledge of distribution of tachykinin receptors and pharmacological effects of the tachykinins, it may be hypothesized that receptor antagonists may have several therapeutic applications. With regard to NK1 receptor antagonists, their therapeutical use has been hypothesized in the treatment of pain and emesis and, in the periphery, in the treatment of several inflammatory diseases including arthritis, inflammatory and motor diseases, and cystitis (Quartara and Maggi, 1998).

At present time, the only documented clinical trial with tachykinin antagonists, more precisely with a SP antagonist, is that carried out in the treatment of moderate to severe major depression by a large team of researchers, starting from the observation that, like clinically used antidepressant and anxiolytic drugs, also SP antagonists suppressed isolation-induced vocalization in the guinea pig (Kramer et al., 1998).

In a placebo-controlled trial, it has been found that in patients suffering from depression, the SP antagonist MK-869 displayed an antidepressive effect greater than that displayed by the first choice drug in depression. paroxetin, and side effects, always in comparison with paroxetin, were less intense. The mechanism of action of MK-869 is, at present, not completely understood. This TK receptor antagonist does not interact with monoamine systems (inhibition of re-uptake of serotonin and/or noradrenaline) like the known antidepressant drugs do; thus, it cannot be excluded that MK-869 does not act only through NK1 receptor blockade. Moreover, like the known antidepressive drugs, MK-869 acts only after 2 to 3 weeks, suggesting the possibility that all antidepressant drugs act via an as yet unclear "common pathway" mechanism (Wahlestedt, 1998).

The enormous theoretical and therapeutical interest that SP antagonists may represent well tolerated antidepressant drugs is obvious. However, the successful therapeutical use of tachykinin antagonists in humans requires some precise accomplishments: a) knowledge of the tachykinin receptor types and subtypes occurring in the different human organs and tissues and evidence that the antagonists on trial compete exactly with the wanted binding site. This is because of the heterogeneity of all tachykinin receptor types; b) lack of important toxicity (side effects) even by long-term administration. Antagonists are generally synthetic, nonpeptide, and non-natural molecules; c) lack of appreciable agonistic activity; and d) brain-permeability for antagonists destined to act in the CNS.

VII. General Conclusions

We have previously shown that tachykinins constitute one of the largest families of peptides in all of the world whose members are present in all animal species from lower invertebrates to mammals. There is no nervous system, from the most primitive in coelenterates to the most developed and complex human CNS, that is lacking a tachykininergic system.

What is the functional significance of this spectacular display of tachykinin-secreting fibers and their receptors? It is beyond doubt that neuronal tachykinins play an important role in neurotransmission/neuromodulation both in the CNS and in periphery. This is demonstrated by the overall occurrence of tachykinins in the brain and other nervous structures from the lowest invertebrates to mammals. Although important, the tachykinin peptide family represents only one of the numerous peptide and nonpeptide families involved in neurotransmission and neuromodulation. Members of these families are expressed in a variety of tissues, and very frequently a tachykinin is costored and cosecreted by the nerve endings with other peptides or biogenic amines. Moreover, the tachykinins, like all other neuropeptides, may enter in competition, positive or negative, with a number of active extraneuronal compounds originating in blood (bradykinin and angiotensin) or in compact or diffuse endocrine organs.

Tachykinins, with their variable primary structure seem to be adapted to display, in the better way, their function in the different invertebrate and vertebrate phyla. In all examined species, and especially in mammals (the phylum more thoroughly studied), tachykinins elicit a spectrum of biological activity (both in the CNS and in the periphery), which may vary conspicuously in the different species and even in the various strains of single species, again strongly supporting the concept of a general, important functional significance of these peptides.

Transgenic mice with disrupted preprotachykinin A gene or with disrupted NK1 receptor gene are in good health conditions and fertile. This demonstrates that the tachykinins are not essential for life and health, at least in mice but probably in the other mammalian species as well, and points to the well known great adaptability of living organisms and the plasticity of homeostatic mechanisms. At present, we do not know any pathological syndrome attributable entirely or predominantly to excess or defect of tachykinin production and release. No function of the various organs and systems in health and disease seems to depend entirely on the tachykininergic

system, and tachykinins seem to be only one arm of the complex mechanism that regulates body functions.

References

- Alumets J, Hakanson R, Ingemansson S, and Sundler F (1977) Substance P and 5-HT in granules isolated from an intestinal argentaffin carcinoid. *Histochemistry* **52:**217–222.
- Anastasi A and Erspamer V (1962) Occurrence and some properties of eledoisin in extracts of posterior salivary glands of *Eledone*. Br J Pharmacol Chem 19:326–354
- Anastasi A, Erspamer V, and Cei JM (1964) Isolation and amino acid sequence of physalaemin, the main active polypeptide of the skin of *Physalaemus fuscomaculatus*. Arch Biochem Biophys 108:341–348.
- Anastasi A, Erspamer V, and Endean R (1975) Structure of uperolein, a physalae-min-like endecapeptide occurring in the skin of *Uperuleia rugosa* and *Uperoleia marmorata*. Experientia 31:394–395.
- Anastasi A and Falconieri Erspamer G (1970) Occurrence of phyllomedusin, a physalaemin-like decapeptide, in the skin of *Phyllomedusa bicolor. Experientia* **26**:866–867.
- Anastasi A, Montecucchi P, Erspamer V, and Visser J (1977) Amino acid composition and sequence of kassinin, a tachykinin dodecapeptide from the skin of the African frog Kassina senegalensis. Experientia 33:857–858.
- Aros B, Wenger T, Vigh B, and Vigh-Teichmann I (1980) Immunocytochemical localization of substance P and ACTH-like activity in the central nervous system of the earthworm Lumbricus terrestris L. Acta Histochem 66:262-268.
- Bannon MJ, Poosch MS, Haverstick DM, Mandal A, Xue IC, Shibata K, and Dragovic LJ (1992) Preprotachykinin gene expression in the human basal ganglia: characterization of mRNAs and pre-mRNAs produced by alternate RNA splicing. *Mol Brain Res* 12:225–231.
- Barber WD, Stevenson GD, and Burks TF (1987) Tachykinins: local gastric effects and brainstem responses. *Am J Physiol* **252**:365–373.

 Barja F and Mathison R (1982) Adrenergic and peptidergic (substance P and vaso-
- Barja F and Mathison R (1982) Adrenergic and peptidergic (substance P and vaso-active intestinal polypeptide) innervation of the rat portal vein. *Blood Vessels* 19:263–272.
- Barocelli E, Chiavarini M, Impicciatore M, Massi M, and De Caro G (1988) Inhibitory effect of intracranial injections of tachykinins on angiotensin-induced drinking in the cat. *Pharmacol Biochem Behav* **31**:493–497.
- Benedeczky I, Kiss JZ, and Somogyi P (1982) Light and electric microscopic localization of substance P-like immunoreactivity in the cerebral ganglion of locust with a monoclonal antibody. *Histochemistry* **75**:123–131.

 Beretta Anguissola A, Feruglio FS, Campus S, Chiandussi L, Pandolfo G, and Berti
- Beretta Anguissola A, Feruglio FS, Campus S, Chiandussi L, Pandolfo G, and Berti G (1966) The effects of eledoisin and bradykinin on the general and visceral circulation, in *Hypotensive Peptides* (Erdos G, Back N, Sicuteri F, and Wilde AF eds) pp 430–440, Springer-Verlag, New York.
- Bergamaschi M and Glasser AH (1963) Effect of the endecapeptide eledoisin on the coronary blood flow: comparison with bradykinin, nitroglycerin and epinephrine in the dog. Circ Res 13:329–335.
- Bergamaschi M and Glasser AH (1964) Peripheral vasodilator action of eledoisin, bradykinin and nitroglycerin in anaesthetized dogs. Circ Res 15:371–379.
- Bergamaschi M, Glasser AH, and Silvestri R (1966) Vasodilating action of eledoisin on the intact and denervated skeletal muscle preparation of the dog. *Arzneimittelforschung* 16:1668–1671.
- Bergstrom M, Theodorsson E, Norheim I, and Oberg K (1995) Immunoreactive tachykinins in 24 h collections of urine from patients with carcinoid tumours: characterization and correlation with plasma concentrations. Scand J Clin Lab Invest 55:679–689.
- Bernardi L, Bosisio G, Chillemi F, De Caro G, De Castiglione R, Erspamer V, Glaesser A, and Goffredo O (1964) Synthetic peptides related to eledoisin. *Experientia* 20:306–309.
- Bertaccini G (1980) Peptides of amphibian skin active on the gut. Tachykinins and caeruleins. Isolation, structure and basic functions, in *Comprehensive Endocrinology, Gastrointestinal Hormones* (Jerzyzzglasszz GB ed) pp 315–341, Raven Press, New York.
- Bertaccini G (1982) Substance P, in *Handbook of Experimental Pharmacology* (Bertaccini G ed) Vol 59/II, pp 85–105, Springer, Berlin.
- Bertaccini G, Cei JM, and Erspamer V (1965) Occurrence of physalaemin in extracts of the skin of *Physalaemus fuscomaculatus* and its pharmacological actions on extravascular smooth muscle. *Br. J. Pharmacol.* **25**:363–379.
- Bertaccini G and Coruzzi G (1977) Action of some natural peptides on the stomach of the anaesthetized rat. Naunyn Schmiedebergs Arch Pharmacol 298:163–166.
- Bertaccini G and De Caro G (1965) The effect of physalaemin and related peptides on salivary secretion. J Physiol 181:68–81.
- Bertaccini G, De Caro G, and Cheli R (1966) Enlargement of salivary glands in rats after chronic administration of physalaemin or isoprenaline. *J Pharm Pharmacol* 18:312–316
- Bertaccini G, De Caro G, and Impicciatore M (1967) Effects of physalaemin on some exocrine secretions of dogs and rats. *J Physiol* 193:497–511.
- Bianchi Porro G, Della Porta P, and Maiolo AT (1965) Portata distrettuale e metabolismo dell'encefalo sotto infusione continua di eledoisina. *Atti Accad Med Lombarda* 20:313-316.
- Bishop AE, Hamid QA, Adams C, Bretherton-Watt D, Jones PM, Denny P, Stamp GW, Hurt RL, Grimelius L, Harmar AJ, et al. (1989) Expression of tachykinins by ileal and lung carcinoid tumors assessed by combined in situ hybridization, immunocytochemistry and radioimmunoassay. *Cancer* 63:1129-1137.
- Bjorklund H, Dalsgaard CJ, Johnsson CE, and Hermansson A (1986) Sensory and autonomic innervation of non-hairy and hairy human skin. An immunohistochemical study. *Cell Tissue Res* **243**:51–57.
- Blitz DM, Christie AE, Marder E, and Nusbaum MP (1995) Distribution and effects

- of tachykinin-like peptides in the stomatogastric nervous system of the crab $Cancer\ borealis.\ J\ Comp\ Neurol\ 354:282-294.$
- Bonner TI, Affolter HU, Young AC, and Young WS (1987) A cDNA encoding the precursor of the rat neuropeptide, neurokinin B. Brain Res 388:243–249.
- Bradford AM, Raftery MJ, Bowie JH, Tyler MJ, Wallace JC, Adams GW, and Severini C (1996) Novel uperin peptides from the dorsal glands of the Australian foodplain toadlet *Uperoleia inundata*. Aust J Chem 49:475–484.
- Bristow LJ and Young L (1994) Chromodacryorrhoea and repetitive hind paw tapping: models of peripheral and central tachykinin NK1 receptor activation in gerbils. Eur J Pharmacol 253:245–252.
- Broccardo M and Improta G (1988) Effect of central administration of tachykinins on thermoregulation and gastric emptying in rats. Regul Pept 22:40.
- Broccardo M, Improta, and Tabacco A (1999) Central tachykinin NK3 receptors in the inhibitory action on the rat colonic propulsion of a new tachykinin PG-KII. Eur J Pharmacol 376:67-71.
- Brodin E, Lindefors N, Dalsgaard CJ, Theodorsson-Norheim E, and Rosell S (1986) Tachykinin multiplicity in rat central nervous system as studied using antisera raised against substance P and neurokinin A. Regul Pept 13:253–272.
- Brown CL and Hanley MR (1981) The effects of substance P and related peptides on alpha-amylase release from rat parotid gland slices. Br J Pharmacol 73:517–523.
- Brunsson I, Fahrenkrug J, Jodal M, Sioqvist A, and Lundgren O (1995) Substance P effects on blood flow, fluid transport and vasoactive intestinal polypeptide release in the feline small intestine. J Physiol 483:727–734.
- Burcher E, Atterhog JH, Pernow B, and Rosell S (1977) Cardiovascular effects of substance P: effects on the heart and regional blood flow in the dog, in $Substance\ P$ (von Euler US and Pernow B eds) pp 261–268, Raven Press, New York.
- Bush FF and Gupta BC (1988) Immunohistochemical localization of regulatory peptides and serotonin in six species of trematode parasites. Comp Biochem Physiol C 91:565-570.
- Cantalamessa F, De Caro G, and Perfumi M (1975) Effects of chronic administration of eledoisin or physalaemin on the rat salivary glands. *Pharmacol Res Commun* 7:259–271.
- Cao YQ, Mantyh PW, Carlson EJ, Gillespie AM, Epstein CJ, and Basbaum AI (1998)
 Primary afferent tachykinins are required to experience moderate to intense pain.
 Nature (Lond) 392:390-394.
- Caprilli R, Frieri G, Palla G, and Broccardo M (1975) Effects of eledoisin on gastro-intestinal electrical activity, in *Abstracts Symposium on Physiology and Pharmacology of Smooth Muscle*, p 12, Varna.
- Cascieri MA, Huang RRC, Fong TM, Cheung AH, Sadowsky S, Beer E, and Strader CD (1992) Determination of the amino acid residues in substance P conferring selectivity and specificity for the rat neurokinin receptors. *Mol Pharmacol* 41: 1096–1099
- Cetin Y, Kuhn M, Kulaksiz H, Adermann K, Bargsten G, Grube D, and Forssmann WG (1994) Enterochromaffin cells of the digestive system: cellular source of guanylin, a guanylate cyclase activating peptide. *Proc Natl Acad Sci USA* **91**: 2935–2939.
- Champagne DE and Ribeiro JM (1994) Sialokinin I and II: vasodilatory tachykinins from the yellow fever mosquito Aedes aegypti. Proc Natl Acad Sci USA 91:138– 149.
- Chang MM and Leeman SE (1970) Isolation of a sialagogic peptide from bovine hypothalamic tissue and its characterization as substance P. J Biol Chem 245: 4784–4790.
- Chang MM, Leeman SE, and Niall HD (1971) Amino-acid sequence of substance P. Nat New Biol 232:86–87.
- Chretien M, Sikstrom R, Lazure C, Mbikay M, Nasiannet S, Marcinkiewicz M, and Seida NG (1989) Expression of the diversity of neuronal and hormonal peptides in the cleavage of precursor molecules, in *Peptide Hormones as Prohormones* (Martinex J ed) pp 1–24, Ellis Horwood, London.
- Christie AE, Lundquist CT, Nassel DR, and Nusbaum MP (1997) Two novel tachy-kinin-related peptides from the nervous system of the crab Cancer borealis. J Exp Biol 200:2279-2294.
- Ciccocioppo R, Panocka I, Polidori C, De Caro G, Regoli D, and Massi M (1997) Stimulation of tachykinin NK3 receptors in the nucleus basalis magnocellularis reduces alcohol intake in rats. Peptides 18:1349–1355.
- reduces alcohol intake in rats. Peptides 18:1349–1355.
 Clottens FL, Meola SM, Coast GM, Hayes TK, Wright MS, Nachman RJ, and Holman GM (1993) Characterization of an antiserum against an achetakinin I-analog and its use for the localization of culekinin depolarizing peptide II in the mosquito Culex salinarius. Regul Pept 49:145–157.
- Conlon JM, Adrian TE, and Secor SM (1997) Tachykinins (substance P, neurokinin A and neuropeptide gamma) and neurotensin from the intestine of the Burmese python. *Phython molurus. Pentides* 18:1505–1510.
- python. *Phython molurus. Peptides* **18:**1505–1510. Conlon JM, Deacon CF, O'Toole L, and Thim L (1986a) Scyliorhinin I and II, two novel tachykinins from dogfish gut. *FEBS Lett* **200:**111–116.
- Conlon JM, Deacon CF, Richter G, Schmidt WE, Stockmann F, and Creutzfeldt W (1986b) Measurement and partial characterization of the multiple forms of neurokinin A-like immunoreactivity in carcinoid tumours. Regul Pept 13:183–196.
- Conlon JM, Katsoulis S, Schmidt WE, and Thim L (1988) [Arg³]substance P and neurokinin A from chicken small intestine. *Regul Pept* **20:**171–180.

 Conlon JM, O'Harte F, Peter RE, and Kah O (1991) Carassin: a tachykinin that is
- Conlon JM, O'Harte F, Peter RE, and Kah O (1991) Carassin: a tachykinin that is structurally related to neuropeptide-gamma from the brain of the goldfish. J Neurochem 56:1432–1436.
- Conlon JM, Schafer G, Schmidt WE, Lazarus LH, Becker HD, and Creutzfeldt W (1985) Chem and immunochemical characterization of substance P-like immunoreactivity and physalaemin-like immunoreactivity in a carcinoid tumour. Regul Pept 11:117–132.
- Conlon JM and Thim L (1988) Isolation of the thachykinin, des[Ser¹, Pro²]scyliorhinin II from the intestine of the ray *Torpedo marmorata*. Gen Comp Endocrinol 71:383–388.
- Conlon JM, Warner FJ, and Burcher E (1998) Bufokinin: a substance P-related peptide from the gut of the toad. *Bufo marinus* with high binding affinity but low selectivity for mammalian tachykinin receptors. *J Pept Res* **51**:210–215.

- Cooper PE, Fernstrom MH, Rorstad OP, Leeman SE, and Martin JB (1981) The regional distribution of somatostatin, substance P and neurotensin in human brain. *Brain Res* 218:219–232.
- Costa JC and Tomaz C (1998) Posttraining administration of substance P and its N-terminal fragment block the amnestic effects of diazepam. Neurobiol Learn Mem 69:65–70.
- Courtice GP, Burcher E, Carlo-Stella R, and Conlon JM (1993) Cardiovascular effects of amphibian and mammalian tachykinins in the toad *Bufo marinus*. Neuropeptides 24:171–176.
- Creutzfeldt W (1996) Carcinoid tumors: development of our knowledge. World J Surg ${f 20:}126-131.$
- Creutzfeldt W and Stockmann F (1987) Carcinoids and carcinoid syndrome. Am J Med 82:4-16.
- Currie MG, Folk KF, Kato J, Moore RJ, Hamra FK, Duffin KL, and Smith CE (1992) Guanylin: an endogenous activator of intestinal guanylate cyclase. *Proc Natl Acad Sci USA* 89:947–951.
- Dalsgaard CJ, Haegerstrand A, Theodorsson-Norheim E, Brodin E, and Hokfelt T (1985) Neurokinin A-like immunoreactivity in rat primary sensory neurons: coexistence with substance P. Histochemistry 83:37–39.
- Dartt DA, Schulman M, Gray KL, Rossi SR, Matkin C, and Gilbard JP (1988) Stimulation of rabbit lacrimal gland secretion with biologically active peptides. Am J Physiol 254:G300-G306.
- De Caro G (1963) Azione dell'eledoisina sulla permeabilità capillare nell'uomo, nella cavia e nel ratto. Arch Int Pharmacodyn 146:27–39.
- De Caro G and Cordella M (1965) Effetti della fisalaemina sulla secrezione lacrimale. Annali Ottalmol Clin Ocul 91:933–939.
- De Caro G, Cordella M, and Miani P (1969) The treatment of Sjoegren's syndrome with physalaemin. *Ophthalmologica* **158**:284–287.
- De Caro G, Farruggia L, Minardi E, and Novarini A (1966) Hypotensive effect of eledoisin, physalaemin and related peptides in man. NaunynSchmiedebergs Arch Pharmakol Exp Pathol 254:194–198.
- De Caro G, Mariotti M, Massi M, and Micossi LG (1980) Dipsogenic effect of angiotensin II, bombesin and tachykinins in the duck. *Pharmacol Biochem Behav* 13:229–233.
- De Caro G, Massi M, and Micossi LG (1978) Potent dipsogenic effect of physalaemin in the pigeon. *Pharmacol Res Comm* **10:**861–866.

 De Caro G, Perfumi M, and Massi M (1988) Tachykinins and body fluid regulation,
- De Caro G, Perfumi M, and Massi M (1988) Tachykinins and body fluid regulation, in Progress in Psychobiology and Physiological Pharmacology (Epstein AE and Morrison AR eds) Vol 13, pp 31–66, Academic Press, New York.
- De Felipe C, Herrero JF, O'Brien JA, Palmer JA, Doyle CA, Smith AJ, Laird JM, Belmonte C, Cervero F, and Hunt SP (1998) Altered nociception, analgesia and aggression in mice lacking the receptor for substance P. Nature (Lond) 392:394–397
- De Pasquale NP and Burch GE (1966) Digital vascular responses to intraarterial injections of bradykinin, kallidin and eledoisin in man. Circulation 34:211-217.
- Devillier P, Drapeau G, Renoux M, and Regoli D (1989) Role of the N-terminal arginine in the histamine-releasing activity of substance P, bradykinin and related peptides. Eur J Pharmacol 168:53–60.
- Devillier P, Renoux M, Giroud JP, and Regoli D (1985) Peptides and histamine release from rat peritoneal mast cells. Eur J Pharmacol 117:89-96.
- Dib B, Corsi MM, Fulgenzi A, Ferrero ME, and Falchi M (1998) Intracerebroventricular injection of capsaicin and substance P provokes the micturition reflex in the rat. Int J Tissue React 20:109–114.
- $\label{eq:Doi T and Jurna J (1981) Intrathecal substance P depresses the tail-flick response: antagonism by naloxone. Naunyn-Schmiedeberg's Arch Pharmacol {\bf 317:}135-139.$
- Doi T and Jurna J (1982) Intrathecal substance P depresses spinal motor and sensory responses to stimulation of nociceptive afferents: antagonism by naloxone. Naunyn-Schmiedeberg's Arch Pharmacol 319:154–160.
- D'Orléans-Juste P, Dion S, Drapeau G, and Regoli D (1986) Different receptors are involved in the endothelium-mediated relaxation and the smooth muscle contraction of the rabbit pulmonary artery in response to substance P and related neurokinins. Eur J Pharmacol 125:37–44.
- D'Orléans-Juste P, Dion S, Mizrahi J, and Regoli D (1985) Effects of peptides and non-peptides on isolated arterial smooth muscle: role of endothelium. *Eur J Pharmacol* 114:9–21.
- Dornan WA and Malsbury CW (1989) Peptidergic control of male rat sexual behavior: the effects of intracerebral injections of substance P and cholecystokinin. *Physiol Behav* **46**:547–556.
- Dornan WA, Malsbury CW, and Penney RB (1987) Facilitation of lordosis by injection of substance P into the midbrain central gray. Neuroendocrinology 45:498-506
- Dougherty PM, Palecek J, Paleckova V, and Willis WD (1995) Infusion of substance P or neurokinin A by microdialysis alters responses of primate spinothalamic tract neurons to cutaneous stimuli and to iontophoretically released excitatory amino acids. *Pain* **61**:411–425.
- Douglas FL, Palkovits M, and Brownstein MJ (1982) Regional distribution of substance P-like immunoreactivity in the lower brainstem of the rat. Brain Res 245:376-378
- Eglezos A, Andrews PV, Boyd RL, and Helme RD (1991) Modulation of the immune response by tachykinins. *Immunol Cell Biol* **69:**285–294.
- Eklund B, Jogestrand T, and Pernow B (1977) Effect of substance P on resistance and capacitance vessels in the human forearm, in $Substance\ P$ (von Euler US and Pernow B eds) pp 275–285, Raven Press, New York.
- Ekstrom J, Mansson B, and Tobin G (1983) Relative secretory contributions of the three major salivary glands of the rat in response to substance P and supersensitivity. Arch Oral Biol 28:859-863.
- El-Salhy M, Falkmer S, Kramer KJ, and Speirs RD (1983) Immunohistochemical investigations of neuropeptides in the brain, corpora cardiaca and corpora allata of an adult lepidopteran insect (*Manduca sexta*) L. Cell Tissue Res 232:295–317.
- Elliott PJ and Iversen SP (1986) Behavioural effects of tachykinins and related peptides. Brain Res 381:68-76.

- Emmelin N and Lenninger S (1967) The "direct" effect of physalaemin on salivary gland cells. Br J Pharmacol 30:676–680.
- Emmelin N, Ohlin P, and Thulin A (1969) The pharmacology of salivary myoepithelial cells in dogs. *Br J Pharmacol* **37:**666–679.
- Emson PC, Arregul A, Clement Jones V, Sandberg BE, and Rossor M (1980) Regional distribution of methionine-enkephalin and substance P-like immunoreactivity in normal human brain and in Huntington's disease. *Brain Res* 199:147–160.
- Ericsson A, Geenen V, Robert F, Legros JJ, Vrindts-Gevaert Y, Franchimont P, Brene S, and Persson H (1990) Expression of preprotachykinin-A and neuropeptide Y messenger RNA in the thymus. *Mol Endocrinol* 4:1211–1218.
- Erjavec F, Lembeck F, Florjanc-Irman T, Skofitsch G, Donnerer J, Saria A, and Holzer P (1981) Release of histamine by substance P. Naunyn-Schmiedeberg's Arch Pharmacol 317:67-70.
- Erspamer V (1949) Ricerche preliminari sulla moschatina. Experientia 5:79.
- Erspamer V (1981) The tachykinin peptides family. Trends Neurosci 4:267–269.
- Erspamer V (1994) Bioactive secretions of the integument, in *Amphibian Biology. I. The Integuments* (Heatwole H and Barthalmus GT eds) pp 178–350, Surrey Beatty & Sons, Chipping Norton, NSW, Australia.
- Erspamer V, Anastasi A, Bertaccini G, and Cei JM (1964) Structure and pharmacological actions of physalaemin, the main active polypeptide of the skin of *Physalaemus fuscomaculatus*. Experientia 20:489-490.
- Erspamer V and Falconieri Erspamer G (1962) Pharmacological actions of eledoisin on extravascular smooth muscle. *Br J Pharmacol* 19:337–354.
- Erspamer V and Glasser A (1963) The action of eledoisin on the systemic arterial blood pressure of some experimental animals. Br J Pharmacol 20:516–527.
- Evans TW, Dixon CM, Clarke B, Conradson TB, and Barnes PJ (1988) Comparison of neurokinin A and substance P on cardiovascular and airway function in man. Br J Clin Pharmacol 25:273–275.
- Felix D (1967) Peptide and acetylcholine action on neurons of the cat subfornical organ. Naunyn-Schmiedeberg's Arch Pharmacol 292:15-20.
- Figini M, Emanueli C, Grady EF, Kirkwood K, Payan DG, Ansel J, Gerard C, Geppetti P, and Bunnett N (1997) Substance P and bradykinin stimulate plasma extravasation in the mouse gastrointestinal tract and pancreas. Am J Physiol 272:G785—G793.
- Fingerman M, Hanumante MM, Kulkarni GK, Ikeda R, and Vacca LL (1985) Localization of substance P-like, leucine-enkephalin-like, methionine-enkephalin-like, FMRFamide-like immunoreactivity in the eyestalk of the fiddler crab *Uca pugilator. Cell Tissue Res* 241:473–477.
- Foreman JC, Jordan CC, Oehme P, and Renner H (1983) Structure/activity relationships for some substance P-related peptides that cause wheal and flare reactions in human skin. *J Physiol* **335**:449–465.
- Frederickson RC, Burgis V, Harrel CE, and Edwards JD (1978) Dual actions of substance P on nociception: possible role of endogenous opioids. *Science (Wash DC)* 199:1359–1362.
- Fregnan GB and Glasser AH (1968) Vasodilating activity of the natural polypeptide physalaemin on hind limb and coronary vascular beds of dog. Comparison with eledoisin and nitroglycerin. Arch Int Pharmacodyn Ther 171:435–448.
- Fritsch HA, Van Noorden S, and Pearse AG (1979) Localization of somatostatin-, substance P- and calcitonin-like immunoreactivity in the neural ganglion of Ciona intestinalis L (Ascidiaceae). $Cell\ Tissue\ Res\ 202:263-274$.
- Fritsch HA, Van Noorden S, and Pearse AG (1980) Substance P-, neurotensin- and bombesin-like immunoreactivities in the gill epithelium of Ciona intestinalis L. Cell Tissue Res ${\bf 208:}467-473.$
- Fritsch HA, Van Noorden S, and Pearse AG (1982) Gastro-intestinal and neurohormonal peptides in the alimentary tract and cerebral complex of *Ciona intestinalis* (Ascidiaceae). *Cell Tissue Res* **223**:369–402.
- Frode-Saleh TS, Calixto JB, and Medeiros YS (1999) Analysis of the inflammatory response induced by substance P in the mouse pleural cavity. *Peptides* **20:**259–265
- Frossard N and Advenier C (1991) Tachykinin receptors and the airways. *Life Sci* **49:**1941–1953.
- Fujisawa J, Muneaka Y, Tabahashi T, Takao T, Shimonishi Y, Kubota I, Ikeda T, Minakata H, Nomoto K, Kiss T, and Hiripi L (1993) An invertebrate-type tachy-kinin isolated from the freshwater bivalve mollusc, Anodonta cygnea, in Peptide Chemistry (Okoda ed) pp 161–164, Protein Research Foundation, Osaka, Japan.
- Furchgott RF (1983) Role of endothelium in responses of vascular smooth muscle. Circ Res **53**:557–573.
- Furchgott RF (1984) The role of endothelium in the response of vascular smooth muscle to drugs. Annu Rev Pharmacol Toxicol 24:176–197.
- Furness JB, Papka RE, Della NG, Costa M, and Eskay RL (1982) Substance P-like immunoreactivity in nerves associated with the vascular system of guinea-pigs. Neuroscience 7:447–459.
- Gaffori O, Stewart JM, and de Wied D (1984) Influence of substance P and fragments on passive avoidance behavior. *Experientia* **40**:89–91.
- Gale JS, Bird ED, Spoke EG, Ivejsen LL, and Jessel T (1978) Human brain substance P: distribution in controls and Huntington's chorea. *J Neurochem* **30:**633–634.
- Gamse R, Mroz E, Leeman S, and Lembeck F (1978) The intestine as source of the immunoreactive substance P in plasma of the cat. Naunyn-Schmiedeberg's Arch Pharmacol 305:17–21.
- Gamse R and Saria A (1986) Nociceptive behavior after intrathecal injections of substance P, neurokinin A and calcitonin gene-related peptide in mice. Neurosci Lett 70:143-147.
- Gamse R, Saria A, Bucsics A, and Lembeck F (1981) Substance P in tumours: pheochromocytoma and carcinoid. *Peptides* (Suppl 2):275–280.
- Gates TS, Zimmerman RP, Mantyh CR, Vigna SR, Maggio JE, Welton ML, Passaro EP, and Mantyh PW (1989) Substance P and substance K receptor binding sites in the human gastrointestinal tract: localization by autoradiography. *Peptides* 9:1207–1219.
- Geppetti P, Maggi CA, Zecchi-Orlandini S, Santicioli P, Meli A, Frilli S, Spillantini MG, and Amenta F (1987) Substance P-like immunoreactivity in capsaicinsensitive structures of the rat thymus. Regul Pept 18:321–329.

- Gibbins IL, Furness JB, Costa M, MacIntyre I, Hillyard CJ, and Girgis S (1985) Co-localization of calcitonin gene-related peptide-like immunoreactivity with substance P in cutaneous, vascular and visceral sensory neurons of guinea pigs. *Neurosci Lett* 57:125–130.
- Gjuris Y and Westermann E (1965) Bronchoconstrictor action of bradykinin, kallidin and eledoisin. Nature (Lond) 205:402–403.
- Goldberg D, Nusbaum MP, and Marder E (1988) Substance P-like immunoreactivity in the stomatogastric nervous systems of the crab Cancer borealis and the lobster Panulirus interruptus and Homarus americanus. Cell Tissue Res 252:515–522.
- Grady EF, Baluk P, Bohm S, Gamp PD, Wong H, Payan DG, Ansel J, Portbury AL, Furness JB, McDonald DM, and Bunnett NW (1996) Characterization of antisera specific to NK1, NK2 and NK3 neurokinin receptors and their utilization to localize receptors in the rat gastrointestinal tract. J Neurosci 16:6975–6986.
- Grimmelikhuijzen CJ, Balfe A, Emson PC, Powell D, and Sundler F (1981) Substance P-like immunoreactivity in the nervous system of hydra. *Histochemistry* **71**:325–333.
- Growcott JW and Shaw JS (1979) Failure of substance P to produce analgesia in the mouse. Br J Pharmacol **66**:129P.
- Guard S and Watson SP (1991) Tachykinin receptor types: classification and membrane signaling mechanism. Neurochem Int 18:149–165.
- Gustafsson MK (1987) Immunocytochemical demonstration of neuropeptides and serotonin in the nervous systems of adult *Schistosoma mansoni*. *Parasitol Res* **74**:168–174.
- Gustafsson MK, Lehtonen MA, and Sundler F (1986) Immunocytochemical evidence for the presence of "mammalian" neurohormonal peptides in neurones of the tapeworm Diphyllobathrium dendriticum. Cell Tissue Res 243:41–49.
- Haefeli A and Hurlimann A (1962) Substance P, a highly active naturally occurring polypeptide. Experientia 18:297–303.
- Hagermark O, Hokfelt T, and Pernow B (1978) Flare and itch induced by substance P in human skin. J Invest Dermatol 71:233–235.
- Hall ME, Grantham P, Limoli J, and Stewart JM (1987) Effects of substance P and neurokinin A (substance k) on motor behavior: unique effect of substance P attributable to its amino-terminal sequence. *Brain Res* **420:**82–94.
- Hall ME, Grantham PA, and Stewart JM (1985) Age and strain differences in some behavioral effects of intracranial substance P. *Peptides* **6:**363–368.
- Hall ME, Miledy F, and Stewart JM (1989) The role of enzymatic processing in the biological actions of substance P. *Peptides* 10:895–901.
- Hall ME and Stewart JM (1984) Modulation of isolation-induced fighting by N- and C-terminal analogs of substance P: evidence for multiple recognition sites. Peptides 5:85–89.
- Hallgren A, Flemstrom G, Hellstrom PM, Lordal M, Hellgren S, and Nylander O (1997) Neurokinin A increases duodenal mucosal permeability, bicarbonate secretion and fluid output in the rat. Am J Physiol 273:G1077–G1086.
- Hanley MR, Lee CM, Michell RHM, and Jones L (1980) Similar effects of substance P and related peptides on salivation and on phosphatidylinositol turnover in rat salivary glands. *Mol Pharmacol* 18:78–83.
- Hartung HP and Toyka HV (1989) Substance P, the immune system and inflammation. Int Rev Immunol 4:229–249.
- Hasenohrl RU, Gerhardt P, and Huston JP (1990) Substance P enhancement of inhibitory avoidance learning: mediation by the N-terminal sequence. *Peptides* 11:163–167.
- Hasenohrl RU, Jentjens O, De Souza Silva MA, Tomaz C, and Huston JP (1998) Anxiolytic-like action of neurokinin substance P administered systematically or into the nucleus basalis magnocellularis region. Eur J Pharmacol 354:123–133.
- Hauge A, Lunde PK, and Waaler BA (1966) The effect of bradykinin, kallidin and eledoisin upon the pulmonary vascular bed of an isolated blood-perfused rabbit lung preparation. Acta Physiol Scand 66:269–277.
- Hayes A and Tyres M (1979) Effects of intrathecal and intracerebroventricular injections of substance P on nociception in the rat and mouse. Br J Pharmacol 66:488P.
- Hedner T, Wessberg P, and Jonason J (1984) Interaction of substance P with the respiratory control system in the rat. J Pharmacol Exp Ther 228:196–201
- Herrera-Marschitz M, Terenius L, Sakurada T, Reid MS, and Ungerstedt U (1990)

 The substance P(1–7) fragment is a potent modulator of substance P actions in the brain. *Brain Res* 521:316–320.
- Hokfelt T, Holets VR, Staines W, Meister B, Melander T, Schalling M, Schultzberg M, Freedman J, Bjorklund H, and Olson L (1986) Coexistence of neuronal messengers: an overview. Prog Brain Res 68:33-70.
- Hokfelt T, Johansson O, Kellerth JO, Ljungdahl A, Nilsson G, Nygards A, and Pernow B (1977) Immunohistochemical distribution of substance P, in *Substance P* (von Euler US and Pernow B eds) pp 117–145, Raven Press, New York.
- Hokfelt T, Kellerth JO, Nilsson G, and Pernow B (1975) Substance P: localization in the central nervous system and in some primary sensory neurons. Science (Wash DC) 190:889-890.
- Holm I, Thulin L, and Hellgren M (1978) Anticholeretic effect of substance P in anaesthetized dogs. Acta Physiol Scand 102:274–280.
- Holstein B and Cederberg C (1986) Effects of tachykinins on gastric acid and pepsin secretion and on gastric outflow in the Atlantic cod *Gadus morhua*. Am J Physiol **250**:G309–315.
- Holzer P (1985) Stimulation and inhibition of gastrointestinal propulsion induced by substance P and substance K in the rat. Br J Pharmacol 86:305–312.
- Holzer P (1991) Capsaicin: cellular targets, mechanisms of action and selectivity for tin sensory neurons. Pharmacol Rev 43:143–201.
- Holzer P and Holzer-Petsche U (1997a) Tachykinins in the gut. Part I. Expression, release and motor function. *Pharmacol Ther* 73:173–217.
- Holzer P and Holzer-Petsche U (1997b) Tachykinins in the gut. Part II. Roles in neural excitation, secretion and inflammation. *Pharmacol Ther* 73:219–263.
- Holzer-Petsche U (1995) Tachykinin receptors in gastrointestinal motility. Regul Pept $\bf 57:19-42$.
- Holzer-Petsche U, Lembeck F, and Seitz H (1987) Contractile effects of substance P

- and neurokinin A on the rat stomach in vivo and in vitro. Br J Pharmacol $\bf 90:$ 273–279.
- Holzer-Petsche U, Schimek E, Amann R, and Lembeck F (1985) In vivo and in vitro actions of mammalian tachykinins. Naunyn-Schmiedeberg's Arch Pharmacol 330: 130–135.
- Hua X, Lundeberg JM, Theodorsson-Norheim E, and Brodin E (1984) Comparison of cardiovascular and bronchoconstrictor effects of substance P, substance K and other tachykinins. Naunyn-Schmiedeberg's Arch Pharmacol 328:196–201.
- Hunter JC, Hannah PA, and Maggio JE (1985) The regional distribution of kassininlike immunoreactivity in central and peripheral tissues of the cat. *Brain Res* 341:228-232.
- Hylden JL and Wilcox GL (1981) Intrathecal substance P elicits a caudally-directed biting and scratching behavior in mice. Brain Res 217:212–215.
- Igwe OJ, Kim DC, Seybold VS, and Larson AA (1990) Specific binding of substance P aminoterminal heptapeptide [SP(1–7)] to mouse brain and spinal cord membranes. J Neurosci 10:3653–3663.
- Ikeda T, Minakata H, Nomoto K, Kubota I, and Muneoka Y (1993) Two novel tachykinin-related neuropeptides in the echiuroid worm *Urechis unicinctus*. Biochem Biophys Res Commun 192:1-6.
- Impicciatore M, Maraini P, and Bertaccini G (1973) Action of eledoisin on human lacrimal secretion in normal and pathological conditions. *Naunyn-Schmiedeberg's Arch Pharmacol* **279**:127–131.
- Improta G and Broccardo M (1990) Tachykinins: effects on gastric secretion and emptying in rats. Pharmacol Res 22:605-610.
- Improta G and Broccardo M (2000) Effects of supraspinal administration of PG-SPI and PG-KII, two amphibian tachykinin peptides, on nociception in the rat. Peptides 21:1611–1616.
- Improta G, Broccardo M, Severini C, and Erspamer V (1996) In vitro and in vivo biological activities of PG-KII, a novel kassinin-like peptide from the skin of the Australian frog *Pseudophryne güntheri*. *Peptides* 17:1003–1008.
- Inagaki S, Senba E, Shiosaka S, Takagi H, Kawai Y, Takatsuki K, Sakanaka M, Matsuzaki T, and Tohyama M (1981) Regional distribution of substance P-like immunoreactivity in the frog brain and spinal cord: immunohistochemical analysis. J Comp Neurol 201:243–254.
- Inagaki S, Sakanaka M, Shiosaka S, Senba E, Takatsuki K, Takagi H, Kawai Y, Minagawa H, and Tohyama M (1982) Ontogeny of substance P-containing neuron system of the rat: immunohistochemical analysis: I. Forebrain and upper brainstem. Neuroscience 7:251–277.
- Inoue M, Kobayashi M, Kozaki S, Zimmer A, and Ueda H (1998) Nociceptin/orphanin FQ-induced nociceptive responses through substance P release from peripheral nerve endings in mice. *Proc Natl Acad Sci USA* **95**:10949–10953.
- Ireland SJ, Bailey F, Cook A, Hagan RM, Jordan CC, and Stephens-Smith ML (1991) Receptors mediating tachykinin-induced contractile responses in guinea-pig trachea. Br J Pharmacol 103:1463–1469.
- Iversen L (1998) Substance P equals pain substance. Nature (Lond) 392:334-335.
- Iwabuchi Y, Masuhara T, and Sofuku S (1992) Sialogogic effects on rat submandibular gland of analogs of the C-terminal hexapeptide of substance P. Jpn J Pharmacol 58:325–328.
- Jaeger W (1988) Treatment of a severe case of cheratoconjunctivitis sicca with eledoisin. Klin Monatsbl Augenheilkd 192:163–166.
- Jaeger W, Gotz ML, and Kaercher T (1985) Eledoisin: a successful therapeutic concept for filamentary keratitis. Trans Ophthalmol Soc U K 104:496.
- Jancso N, Jancso-Gabor A, and Szolcsanyi J (1967) Direct evidence for neurogenic inflammation and its prevention by denervation and by pretreatment with capsaicin. Br J Pharmacol 31:138-151.
- Jensen J (1997) Co-release of substance P and neurokinin A from the Atlantic cod stomach. Peptides 18:717–722.
- Jensen J and Conlon JM (1992) Substance P-related and neurokinin A-related peptides from the brain of the cod and trout. Eur J Biochem 206:659-664.
- Jensen J, Olson KR and Conlon JM (1993) Primary structures and effects on gastrointestinal motility of tachykinins from the rainbow trout. Am J Physiol 265:R804–R810.
- Jensen RT and Gardner JD (1979) Interaction of physalaemin, substance P and eledoisin with specific membrane receptors on pancreatic acinar cells. *Proc Natl Acad Sci USA* 76:5679–5683
- Joos G, Kips J, Pauwels R, and van der Straeten M (1986) The effect of tachykinins on the conducting airways of the rat. Arch Int Pharmacodyn Ther 280:176–190.
- Kage R, McGregor GP, Thim L, and Conlon M (1988) Neuropeptide gamma: a peptide isolated from rabbit intestine that is derived from gamma-preprotachykinin. J Neurochem 50:1412–1417.
- Kagstrom J, Axelsson M, Jensen J, Farrell AP, and Holmgren S (1996) Vasoactivity and immunoreactivity of fish tachykinins in the vascular system of the spiny dogfish. Am J Physiol 270:R585–R593.
- Kaloustian KV and Edmands JA (1986) Immunochemical evidence for substance P-like peptide in tissues of the earthworm *Lumbricus terrestris*: action on intestinal contraction. *Comp Biochem Physiol C* 83:329–333.
- Kanazawa I and Jessell T (1976) Post mortem changes and regional distribution of substance P in the rat and mouse nervous system. Brain Res 117:362–367.
- Kanazawa I, Ogawa T, Kimura S, and Munekata E (1984) Regional distribution of substance P, neurokinin α and neurokinin β in rat central nervous system. Neurosci Res 2:111–120.
- Kangawa K, Minamino N, Fukada A, and Matsuo H (1983) Neuromedin K: a novel mammalian tachykinin identified in porcine spinal cord. Biochem Biophys Res Commun 114:533-540.
- Kantor TG, Jarvik ME, and Wolff BB (1967) Bradykinin as a mediator of human pain. Proc Soc Exp Biol Med 126:505–507.
- Karila P, Shahbazi F, Jensen J, and Holmgren S (1998) Projections and actions of tachykininergic, cholinergic and serotoninergic neurones in the intestine of the Atlantic cod. Cell Tissue Res 291:403–413.
- Kelley AE and Iversen SD (1979) Substance P infusion into substantia nigra of the

rat: behavioural analysis and involvement of striatal dopamine. $Eur\ J\ Pharmacol\ 60:171-179.$

- Kelley AE, Stinus L, and Iversen SD (1979) Behavioural activation induced in the rat by substance P infusion into ventral tegmental area: implication of dopaminergic A10 neurones. Neurosci Lett 11:335–339.
- Khan S, Sandhu J, Whelpton R, and Michael-Titus AT (1998) Substance P fragments and striatal endogenous dopamine outflow: interaction with substance P. Neuropeptides 32:519–526.
- Kimura H and Schubert D (1993) Amyloid β-protein activates tachykinin receptors and inositol triphosphate accumulation by synergy with glutamate. Proc Natl Acad Sci USA 90:7508-7512.
- Kimura S, Okada M, Sugita Y, Kanazawa I, and Munekata E (1983) Novel neuropeptides, neurokinin a and b, isolated from porcine spinal cord. Proc Jpn Acad B 59:101–104.
- Kirkwood KS, Kim EH, He XD, Calaustro EQ, Domush C, Yoshimi SK, Grady EF, Maa J, Bunnett NW, and Debas HT (1999) Substance P inhibits pancreatic exocrine secretion via a neural mechanism. Am J Physiol 277:G314–G320.
- Konishi S and Otsuka M (1974) The effects of substance P and other peptides on spinal neurons of the frog. Brain Res 65:397–410.
- Konturek SJ, Jaworek J, Tasler J, Cieszkowski M, and Pawlik W (1981) Effect of substance P and its C-terminal hexapeptide on gastric and pancreatic secretion in the dog. Am J Physiol 241:G74–G81.
- Kotani H, Hoshimaru M, Nawa H, and Nakanishi S (1986) Structure and gene organization of bovine neuromedin K precursor. Proc Natl Acad Sci USA 83:7074– 7078.
- Kozawa H, Hino J, Minamino N, Kangawa K, and Matsuo H (1991) Isolation of four novel tachykinins from frog (Rana catesbeiana) brain and intestine. Biochem Biophys Res Commun 177:588-595.
- Kramer MS, Cutler N, Feighner J, Shrivastava R, Carman J, Sramek JJ, Reines SA, Liu G, Snavely D, and Wyatt-Knowles E (1998) Distinct mechanism for antidepressant activity by blockade of central substance P receptors. Science (Wash DC) 281:1640-1645
- Krause JE, Chirgwin JM, Carter MS, Xu ZS, and Hershey AD (1987) Three rat preprotachykinin mRNAs encode the neuropeptides substance P and neurokinin A. Proc Natl Acad Sci USA 84:881–885.
- Kuwahara A and Yanaihara N (1987) Action of the newly discovered mammalian tachykinins substance k and neuromedin k on gastroduodenal motility of anaesthetized dogs. Regul Pept 17:221–228.
- Lambert GA and Lang WJ (1970) The effects of bradykinin and eledoisin injected into the cerebral ventricles of conscious rats. Eur J Pharmacol 9:383–386.
- Lazarus LH and Di Augustine RP (1980) Radioimmunoassay for the tachykinin peptide physalaemin: detection of a physalaemin-like substance in rabbit stomach. Anal Biochem 107:350–357.
- Lazarus LH, Di Augustine RP, Jahnke GD, and Hernandez O (1983) Physalaemin: an amphibian tachykinin in human lung small-cell carcinoma. *Science (Wash DC)* **219**:79–81.
- Lazarus LH, Linnoila RI, Hernandez O, and Di Augustine RP (1980) A neuropeptide in mammalian tissues with physalaemin-like immunoreactivity. *Nature (Lond)* 287:555–558.
- Lembeck F, Bernatzky G, Gamse R, and Saria A (1985) Characterization of substance P-like immunoreactivity in submammalian species by high performance liquid chromatography. *Peptides* **6** (Suppl 3):231–236.
- Lembeck F and Holzer P (1979) Substance P as neurogenic mediator of antidromic vasodilation and neurogenic plasma extravasation. Naunyn-Schmiedeberg's Arch Pharmacol 310:175–183.
- Lembeck F and Starke K (1968) Substance P and salivary secretion. Naunyn-Schmiedeberg's Arch Exp Pathol Pharmakol 259:375–385.
- Lembeck F, Starke K, and Weiss U (1968) Sialogenic action of substance P and physalaemin-like octapeptide upon infusion into various vessels. *Naunyn-Schmiedeberg's Arch Exp Pathol Pharmakol* **261**:329–337.
- Lidberg P, Dahlstrom A, Lundberg JM, and Ahlman H (1983) Different modes of action of substance P in the motor control of the feline stomach and pylorus. Regul Pept 7:41–52.
- Lieberman DN and Mody I (1998) Substance P enhances NMDA channel function in hippocampal dentate gyrus granule cells. J Neurophysiol 80:113–119.
- Lin XW and Peter RE (1997) Goldfish gamma-preprotachykinin mRNA encodes the neuropeptides substance P, carassin and neurokinin A. *Peptides* **18**:817–824.
- Lindefors N, Brodin E, Theodorsson-Norheim E, and Ungerstedt U (1985) Regional distribution and in vivo release of tachykinin-like immunoreactivities in rat brain: evidence for regional differences in relative proportions of tachykinins. Regul Pept 10:217–230.
- Linnik MD and Moskowitz MA (1989) Identification of immunoreactive substance P in human and other mammalian endothelial cells. *Peptides* **10:**957–962. Liu H, Mazarati AM, Katsumori H, Sankar R, and Wasterlain CG (1999a) Substance
- Liu H, Mazarati AM, Katsumori H, Sankar R, and Wasterlain CG (1999a) Substance P is expressed in hippocampal principal neurons during status epilepticus and plays a critical role in the maintenance of status epilepticus. Proc Natl Acad Sci USA 96:5286-5291.
- Liu L, Warner FJ, Conlon JM, and Burcher E (1999b) Pharmacological and biochemical investigation of receptors for the toad and the gut tachykinin peptide bufokinin, in its species of origin. Naunyn Schmiedeberg's Arch Pharmacol 360:187–195.
- Liu Chen LY, Liszczak TM, King JC, and Moskowitz MA (1986) Immunoelectron microscopic study of substance P-containing fibers in feline cerebral arteries. Brain Res 369:12–20.
- Lochner W and Parratt JR (1966) A comparison of the effects of locally and systematically administered kinins on coronary blood flow and myocardial metabolism. $Br\ J\ Pharmacol\ 26:17-26.$
- Lordal M, Hallgren A, Nylander O, and Hellstrom PM (1996) Tachykinins increase vascular permeability in the gastrointestinal tract of the rat. *Acta Physiol Scand* **156**:489–494.
- Losay J, Mroz E, Tregear GW, Leeman SE, and Gamble WJ (1977) Action of

- substance P on the coronary blood flow in the isolated dog heart, in *Substance P* (von Euler US and Pernow B eds) pp 287–293, Raven Press, New York.
- Lu YA, Peng JL, Zhu YQ, Wu SX, Tang YQ, Tian SH, and Zou G (1990) Synthesis and biological activity of a new frog skin peptide, ranamargarin. Sci China Ser B Chem Life Sci Earth Sci 33:170–177.
- Lundberg JM, Brodin E, and Saria A (1983) Effects and distribution of vagal capsaicin-sensitive substance P neurons with special reference to the trachea and lungs. *Acta Physiol Scand* 119:243–252.
- Lundquist CT, Clottens FL, Holman GM, Nichols R, Nachman RJ, and Nassel DR (1994) Callitachykinin I and II, two novel myotropic peptides isolated from the blowfly Calliphora vomitoria, that have resemblances to tachykinins. Peptides 15:761–768
- Ma QP and Woolf CJ (1997) Tachykinin NK1 receptor antagonist RP 67580 attenuates progressive hypersensitivity of flexor reflex during experimental inflammation in rats. Eur. J. Pharmacol. 322:165–171.
- Magee RM, Fairweather I, Johnston CF, Halton DW, and Shaw C (1989) Immunocytochemical demonstration of neuropeptides in the nervous system of the liver fluke Fasciola hepatica (Trematoda, Digenea). Parasitology 98:227–238.
- Maggi CA (1991) The role of peptides in the regulation of the micturition reflex: an update. Gen Pharmacol 22:1–24.
- Maggi CA (1997) The effects of tachykinins on inflammatory and immune cells. Regul Pept 70:75–90.
- Maggi CA, Catalioto RM, Criscuoli M, Cucchi P, Giuliani S, Lecci A, Lippi A, Meini S, Patacchini R, Renzetti AR, et al. (1997) Tachykinin receptors and intestinal motility. Can J Physiol Pharmacol 75:696–703.
- Maggi CA, Geppetti P, Santicioli P, Frilli S, Giuliani S, Furio M, Theodorsson E, Fusco B, and Meli A (1988) Tachykinin-like immunoreactivity in the mammalian urinary bladder: correlation with the functions of the capsaicin-sensitive sensory nerves. Neuroscience 26:233-242.
- Maggi CA, Giuliani S, Santicioli P, Abelli L, Regoli D, and Meli A (1987) Further studies on the mechanisms of the tachykinin-induced activation of micturition reflex in rats: evidence for the involvement of capsaicin-sensitive bladder mechanoreceptors. Eur J Pharmacol 136:189–205.
- Maggi CA and Meli A (1986) The role of neuropeptides in the regulation of the micturition reflex. J Auton Pharmacol 6:133–162.
- Maggi CA and Meli A (1988) The sensory-efferent function of capsaicin-sensitive sensory neurons. Gen Pharmacol 19:1–43.
- Maggi CA, Patacchini R, Quartara L, Rovero E, and Santicioli P (1991) Tachykinin receptors in the guinea-pig isolated bronchi. Eur J Pharmacol 197:167–174.
- Maggi CA, Patacchini R, Rovero P, and Giachetti A (1993) Tachykinin receptors and tachykinin receptor antagonists. *J Auton Pharmacol* 13:23–93.
- Maggi CA, Santicioli P, Borsini F, Giuliani S, and Meli A (1986a) The role of the capsaicin-sensitive innervation of the rat urinary bladder in the activation of micturition reflex. Naunyn-Schmiedeberg's Arch Pharmacol 332:276-283.
- Maggi CA, Santicioli P, Giuliani S, Regoli D, and Meli A (1986b) Activation of micturition reflex by substance P and substance K: indirect evidence for the existence of multiple tachykinin receptors in the rat urinary bladder. J Pharmacol Exp Ther 238:259-266.
- Maggio JE (1985) "Kassinin" in mammals: the newest tachykinins. Peptides 6 (Suppl 3):237–243.
- Maggio JE (1988) Tachykinins. Annu Rev Neurosci 11:13-28.
- Makridis C, Theodorsson E, Akerstrom G, Oberg K, and Knutson L (1999) Increased intestinal non-substance P tachykinin concentrations in malignant midgut carcinoid disease. J Gastroenterol Hepatol 14:500-507.
- Malick JB and Goldstein JM (1978) Analgesic activity of substance P after intracerebral administration in rats. *Life Sci* 23:835–844.
- Malthe-Sphirenssen D, Cheney DL, and Costa E (1978) Modulation of acetylcholine metabolism in the hippocampal cholinergic pathway by intraseptally injected substance P. J Pharmacol Exp Ther 206:21–28.
- Mancillas JR, McGinty JF, Selverstone AI, Karten H, and Bloom FE (1981) Immunocytochemical localization of enkephalin and substance P in retina and eyestalk neurones of lobster. *Nature (Lond)* **293:**576–578.
- Mancillas JR and Selverstone AI (1985) Substance P-like immunoreactivity is present in the central nervous system of *Limulus polyphemus*. J Comp Neurol 238:38-52
- Mantovani P, Piccin GL, and Bertaccini G (1969) Activity ratio between intestinal and cardiovascular actions of caerulein and related substances in the anaesthetized dog. *Pharmacol Res* 1:172–174.
- Manzini S, Conti S, Maggi CA, Abelli L, Somma V, Del Bianco E, and Geppetti P (1989) Regional differences in the motor and inflammatory responses to capsaicin in guinea pig airways. Correlation with content and release of substance P-like immunoreactivity. Am Rev Respir Dis 140:936-941.
- Marcinkiewicz N, Seidah NG, and Chretien M (1993) Les convertases des prohormones et le systéme nerveus. Médecine/Science (Wash DC) 9:553-561.
- Mason GS, Graham EA, and Elliott PJ (1992) NK1 receptors in the guinea-pig mediate locomotor hyperactivity Br J Pharmacol 105:251P.
- Massi M, de Caro G, Perfumi M, and Venturi F (1988) Mapping of brain sites sensitive to the antidipsogenic effect of tachykinins. *Peptides* 9:347–356.
- Massi M and Epstein AN (1989) Suppression of salt intake in the rat by neurokinin A: comparison with the effect of kassinin. Regul Pep 24:233–244.
- Massi M, Gentili L, Perfumi M, de Caro G, and Schulkin J (1990) Inhibition of salt appetite in the rat following injection of tachykinins into the medial amygdala. Brain Res 513:1–7.
- Massi M, Micossi LG, de Caro G, and Epstein AN (1986) Suppression of drinking but not feeding by central eledoisin and physalaemin in the rat. Appetite 7:63–70.
- Massi M, Perfumi M, Polidori C, and de Caro G (1987) Effect of kassinin, neurokinin A and neurokinin B on drinking behaviour in the pigeon. *Regul Pept* 17:85–97. Massi M, Saija A, Polidori C, Perfumi M, Gentili L, Costa G, and de Caro G (1991)
- Massi M, Saija A, Polidori C, Perfumi M, Gentili L, Costa G, and de Caro G (1991) The hypothalamic paraventricular nucleus is a site of action for the central effect of tachykinins on plasma vasopressin. Brain Res Bull 26:149–154.
- Mastrangelo D, Mathison R, Huggel HJ, Dion S, D'Orléans-Juste P, Rhaleb NE,

- Drapeau G, Rovero P, and Regoli D (1987) The rat isolated portal vein: a preparation sensitive to neurokinins, particularly neurokinin B. Eur J Pharmacol 134: 321–326
- Matheson MJ, Rynell AC, McClean NA, and Berend N (1997) Tachykinins do not cause plasma leakage in the rabbit trachea. Respir Physiol 108:165-170.
- Matsumoto S, Takeda M, Saiki C, Takahashi T, and Ojima K (1997) Effects of tachykinins on rapidly adapting pulmonary stretch receptors and total lung resistance in anaesthetized artificially ventilated rabbits. J Pharmacol Exp Ther 283: 1026–1031.
- Matsuto T, Yanagisawa M, Otsuka M, Kanazawa I, and Munekata E (1984) The excitatory action of the newly-discovered mammalian tachykinins, neurokinin alpha and neurokinin beta, on neurones of the isolated spinal cord of the newborn rat. Neurosci Res 2:105–110.
- Maubach KA, Cody C, and Jones RS (1998) Tachykinins may modify spontaneous epileptiform activity in the rat entorhinal cortex in vitro by activating GABAergic inhibition. Neuroscience 83:1047–1062.
- Maule AG, Shaw C, Halton DW, Johnston CF, Fairweather I, and Buchanan KD (1989) Tachykinin immunoreactivity in the parasitic flatworm *Diclidophora merlangi* and its fish host the whiting (*Merlangius merlangus*): radioimmunoassay and chromatographic characterization using region-specific substance P and neurokinin A antisera. *Comp Biochem Physiol C* **94:**533–541.
- May RJ, Conlon TP, Erspamer V, and Gardner JD (1978) Actions of peptides isolated from amphibian skin on pancreatic acinar cells. Am J Physiol 235:E112–E118.
- Mazurek N, Pecht I, Teichberg VI, and Blumberg S (1981) The role of N-terminal tetrapeptide in the histamine-releasing action of substance P. Neuropharmacology 20:1025–1027.
- McFadden D, Zinner MJ, and Jaffe BM (1986) Substance P-induced intestinal secretion of water and electrolytes. *Gut* 27:267–272.
- McGillis JP, Mitsuhashi M, and Payan DG (1990) Immunomodulation by tachykinin neuropeptides. Ann NY Acad Sci 594:85–94.
- Melchiorri P, Tonelli F, and Negri L (1977) Comparative circulatory effects of substance P, eledoisin and physalaemin in the dog, in *Substance P* (Euler von US and Pernow B eds) pp 311–319, Raven Press, New York.
- Merchenthaler I, Maderdrut JL, O'Harte F, and Conlon JM (1992) Localization of neurokinin B in the central nervous system of the rat. *Peptides* 13:815–829.
- Michael-Titus AT, Blackburn D, Connolly Y, Priestley JV, and Whelpton R (1999) Nand C-terminal substance P fragments: differential effects on striatal (3H)substance P binding and NK1 receptor internalization. *Neuroreport* 10:2209–2213.
- Mignogna G, Severini C, Erspamer GF, Siciliano R, Kreil G, and Barra D (1997) Tachykinins and other biologically active peptides from the skin of the Costa Rican phyllomedusid frog Agalychnis callidryas. Peptides 18:367–372.
- Mohrland JS and Gebhart GF (1979) Substance P-induced analgesia in the rat. Brain Res 171:556–559.
- Moochhala SM and Sawynok J (1984) Hyperalgesia produced by intrathecal substance P and related peptides: desensitization and cross desensitization. Br J Pharmacol 82:381–388.
- Muren JE and Nassel DR (1996) Isolation of five tachikinin-related peptides from the midgut of the cockroach *Leucophaea maderae*: existence of N-terminally extended isoforms. *Regul Pept* **65**:185–196.
- Nagashima AY, Takano Y, Tateishi K, Matsuoka Y, Hamaoka T, and Kamiya H (1989) Cardiovascular roles of tachykinin peptides in the nucleus tractus solitarii of rats. Brain Res 487:392–396.
- Nakajima T, Yasuhara T, Erspamer V, Erspamer GF, Negri L, and Endean R (1980) Physalaemin- and bombesin-like peptides in the skin of the Australian leptodactylid frog *Uperuleia rugosa. Chem Pharm Bull (Tokyo)* **28**:689–695. Nakajima Y, Tsuchida K, Negishi M, Ito S, and Nakanishi S (1992) Direct linkage of
- Nakajima Y, Tsuchida K, Negishi M, Ito S, and Nakanishi S (1992) Direct linkage of three tachykinin receptors to stimulation of both phosphatidylinositol hydrolysis and cyclic AMP cascades in transfected Chinese hamster ovary cells. J Biol Chem 267:2437-2442
- Nakamura M, Chitama T, and Nishida T (1999) Synergistic effect with Phe-Gly-Leu-Met-NH2 of the C-terminal of substance P and insulin-like growth factor-1 on epithelial wound healing of rabbit cornea. Br J Pharmacol 127:489-497.
- Nakamura M, Nishida T, Ofuji K, Reif TW, Mannis MJ, and Murphy CJ (1997) Synergistic effect of substance P with epidermal growth factor on epithelial migration in rabbit cornea. Exp Eye Res 65:321–329.
- Nakanishi S (1987) Substance P precursor and kininogen: their structures, gene organizations and regulation. *Physiol Rev* **67:**1117–1142.
- Nakano J (1964) Studies on the cardiovascular effects of synthetic eledoisin. J Pharmacol Exp Ther 145:71–77.
- Nakano J (1965) Effects of eledoisin on the systemic venous return in dogs. Proc Soc Exp Biol Med 118:108—110.
- Nakano J, Darrow BA, and McCurdy JR (1968) Cardiovascular effects of synthetic physalaemin. Arch Int Pharmacodyn 172:429-434.
- Nassel DR (1999) Tachykinin-related peptides in invertebrates: a review. *Peptides* 57:141–158.
- Nawa H, Hirose T, Takashima H, Inayaka S, and Nakanishi S (1983) Nucleotide sequences of cloned cDNA for two types of bovine brain substance P precursor. *Nature (Lond)* **306**:32–36.
- Nawa H, Kotani H, and Nakanishi S (1984) Tissue-specific generation of two preprotachykinin mRNAs from one gene by alternative RNA splicing. *Nature (Lond)* 312:729–734
- Nieto J, Veelaert D, Derua R, Waelkens E, Cerstiaens A, Coast G, Devreese B, Van Beeumen J, Calderon J, De Loof A, and Schoofs L (1998) Identification of one tachykinin- and two kinin-related peptides in the brain of the white shrimp Penaeus vannamei. Biochem Biophys Res Commun 248:406-411.
- Nilsson G, Dahlberg K, Brodue E, Sundler F, and Strandberg K (1977) Distribution and constrictor effect of substance P in guinea pig tracheobronchial tissue, in Substance P (von Euler US and Pernow B eds) pp 75–82, Raven Press, New York.
- Norheim I, Theodorsson-Norheim E, Brodin E, Oberg K, Lundqvist G, and Rosell S (1984) Antisera raised against eledoisin and kassinin detect elevated levels of

- immunoreactive materials in plasma and tumor tissues from patients with carcinoid tumors. Regul Pept 9:245–257.
- Norheim I, Wilander E, Öberg K, Theodorsson-Norheim E, Lundqvist ML, Lindgren P, and Bergh J (1987) Tachykinin production by carcinoid tumours in culture. *Eur J Cancer Clin Oncol* **23**:689–695.
- Ogawa T, Kanazawa I, and Kimura S (1985) Regional distribution of substance P, neurokinin alpha and neurokinin beta in rat spinal cord, nerve roots and dorsal root ganglia and the effects of dorsal root section or spinal transection. *Brain Res* 359:152–157.
- O'Harte F, Burcher E, Lovas S, Smith DD, Vaudry H, and Conlon JM (1991) Ranakinin: a novel NK1 tachykinin receptor agonist isolated with neurokinin B from the brain of the frog Rana ridibunda. J Neurochem 57:2086–2091.
- O'Neil GS, Conlon JM, Deacon CF, and Thorndyke MC (1987) Tachykinins in the central and peripheral nervous system of the ascidian *Ciona Intestinalis*. Gen Comp Endocrinol **66**:314–322.
- Ormas P, Castelli S, Beretta CM, Nilsson I, Galbiati A, Beretta C, and Faustini R (1977) The effects of eledoisin on intestinal smooth muscles of ruminants. Folia Vet Lat 7:252–257
- Ormas P, Pompa G, Beretta C, and Faustini R (1975) The effects of some polypeptides on the systemic blood pressure of sheep. Folia Vet Lat 5:45-54.
- Otsuka M and Yoshioka K (1993) Neurotransmitter functions of mammalian tachykinins. *Physiol Rev* **73:**229–308.
- Pearse AG and Polak JM (1975) Immunocytochemical localization of substance P in mammalian intestine. *Histochemistry* 41:373–375.
- Pearson L, Lambert GA, and Lang WJ (1969) Centrally mediated cardiovascular and EEG responses to bradykinin and eledoisin. Eur J Pharmacol 8:153–158.
- Pernow B (1983) Substance P. Pharmacol Rev 35:85-141.
- Pernow B (1985) Role of tachykinins in neurogenic inflammation. J Immunol 135: 812s–815s.
- Pernow B and Rosell S (1975) Effect of substance P on blood flow in canine adipose tissue and skeletal muscle. *Acta Physiol Scand* **93**:139–141.
- Phillis JW and Limacher JJ (1974) Excitation of cerebral cortical neurons by various polypeptides. Exp Neurol 43:414–423.
- Pierobon P, Kemali M, and Milici N (1989) Substance P and Hydra: an immunohistochemical and physiological study. Comp Biochem Physiol C 92:217–221.
- Pietrowski W, Devoy MAB, Jordan CC, and Foreman JC (1984) The substance P receptor on rat mast cells and in human skin. Agents Actions 14:425–428.
- Polidori C, Saija A, Perfumi M, Costa G, De Caro G, and Massi M (1989) Vasopressin release induced by intracranial injection of tachykinins is due to activation of central neurokinin-3 receptors. *Neurosci Lett* 103:320–325.
- Quartara L and Maggi CA (1997) The tachykinin NK1 receptor. Part I: ligands and mechanisms of cellular activation. Neuropeptides 31:537–563.
- Quartara L and Maggi CA (1998) The tachykinin NK1 receptor. Part II: distribution and pathophysiological roles. Neuropeptides 32:1–49.
- Raffa RB (1998) Possible role(s) of neurokinins in CNS development and neurodegenerative or other disorders. Neurosci Biobehav Rev 22:789-813.
- Ralevic V, Milner P, Hudlicka O, Kristek F, and Burnstock G (1990) Substance P is released from the endothelium of normal and capsaicin-treated rat hind limb vasculature, in vivo, by increased flow. Circ Res 66:1178–1183.
- Regoli D, Boudon A, and Fauchère GL (1994a) Receptors and antagonists for substance P and related peptides. *Pharmacol Rev* 46:551–599.
- Regoli D, Drapeau G, Dion S, and D'Orléans-Juste P (1987) Pharmacological receptors for substance P and neurokinins. *Life Sci* **40:**109–117.
- Regoli D, Nguyen QT, and Jukic D (1994b) Neurokinin receptor subtypes characterized by biological assays. Life Sci 54:2035–2047.
- Rogers DF, Belvisi MG, Aurusdkij B, Dijk S, Evans TW, and Barnes PJ (1988) Effects and interactions of sensory neuropeptides on airway microvascular leakage in guinea-pigs. Br J Pharmacol 95:1109–1116.
- Roth KA, Makk G, Beck O, Faull K, Tatemoto K, Evans CJ, and Barchas JD (1985) Isolation and characterization of substance P, substance P5–11 and substance K from two metastatic ileal carcinoids. *Regul Pept* 12:185–199.
- Rudlich L and Butcher FR (1976) Effect of substance P and eledoisin on K⁺ efflux, amylase release and cyclic nucleotide levels in slices of rat parotid gland. *Biochem Biophys Acta* 444:704–711.
- Rupniak NM and Williams AR (1994) Differential inhibition of foot tapping and chromodacryorrhoea in gerbils by CNS penetrant and non-penetrant NK1 receptor antagonists. Eur J Pharmacol 265:179–183.
- Sakanaka M, Inagaki S, Shiosaka S, Senba E, Takagi H, Takatsuki K, Kawai Y, Iida H, Hara Y, and Tohyama M (1982) Ontogeny of substance P-containing neuron system of the rat: immunohistochemical analysis: II. Lower brain stem. Neuroscience 7:1097–1126.
- Sakurada T, Tan-No K, Yamada T, Sakurada S, and Kisaka K (1990a) Phosphoramidon potentiates mammalian tachykinin induced biting, licking and scratching behaviour in mice. *Pharmacol Biochem Behav* 37:779–783.
- Sakurada T, Tan-No K, Yamada T, Sakurada S, Kisaka K, Ohba M, and Terenius L (1990b) N-terminal substance P fragments inhibit the spinally induced NK1 receptor mediated behavioural response in mice. *Life Sci* 47:PL109–PL113.
- Salonen RO, Webber SE, and Widdicombe JG (1988) Effects of neuropeptides and capsaicin on the canine tracheal vasculature in vivo. Br J Pharmacol 95:1262–1270.
- Samnegard H, Thulin L, Thydèn G, Johansson C, Muhrbeck O, and Bjorklund C (1978) Effect of synthetic substance P on internal carotid blood flow in man. *Acta Physiol Scand* **104**:491–495.
- Saria A, Martling CR, Dalsgaard CJ, and Lundberg JM (1985) Evidence for substance P-immunoreactive spinal afferents that mediate bronchoconstriction. *Acta Physiol Scand* **125**:407–414.
- Schildein S, Agmo A, Huston JP, and Schwarting RK (1998) Intraaccumbens injections of substance P, morphine and amphetamine: effects in conditioned place preference and behavioral activity. *Brain Res* **790:**185–194.
- Schlesinger K, Lipsitz DU, Peck PL, Pelleymounter MA, Stewart JM, and Chase TM

- (1983) Substance P enhancement of passive and active avoidance conditioning in mice. Pharmacol Biochem Behav 19:655-661.
- Schlesinger K, Pelleymounter MA, van de Kamp I, Bader DL, Stewart JM, and Chase TM (1986) Substance P facilitation of memory: effects in an appetitively motivated learning task. Behav Neural Biol 45:230–239.
 Schmidt PT, Rickelt LF, and Holst JJ (1999) Tachykinins stimulate release of
- peptide hormones (glucagon-like peptide-1 and paracrine somatostatin) and neurotransmitter (vasoactive intestinal polypeptide) from porcine ileum through NK1 receptors. Dig Dis Sci 44:1273-1281.
- Schneyer CA and Hall HD (1968) Characterization of physalaemin-evoked rat saliva and failure of autonomic blocking agents to modify composition. Proc Soc Exp Biol Med 127:1245-1248.
- Schoofs L, Hollman GM, Hayes TK, Kochansky IP, Nachman RI, and De Loof A (1990a) Locustatachykinins III and IV: two additional insect neuropeptides with homology to peptides of the vertebrate tachykinin family. Regul Pept 31:199-212.
- Schoofs L, Hollman GM, Hayes TK, Nachman RI, and De Loof A (1990b) Locustatachykinin I and II, two novel insect neuropeptides with homology to peptides of the vertebrate tachykinin family. FEBS Lett 261:397-401.
- Severini C, Salvadori S, Guerrini R, Falconieri-Erspamer G, Mignogna G, and Erspamer V (2000) Parallel bioassay of 39 tachykinins on 11 smooth muscle preparations, Structure and receptor selectivity/affinity relationship, Peptides 21: 1587-1595
- Seybold VS, Hylden JLK, and Wilcox GL (1982) Intrathecal substance P and somatostatin in rats: behaviors indicative of sensation. Peptides 3:49-54.
- Sharkey KA, Williams RG, Schultzberg M, and Dockray GJ (1983) Sensory substance P-innervation of the urinary bladder: possible site of action of capsaicin in causing urine retention in rats. *Neuroscience* **10:**861–868.
 Sherwood JE, Mauser PJ, and Chapman RW (1977) Bronchoconstrictor and respi-
- ratory effects of neurokinin A in dogs. J Pharmacol Exp Ther 283:788-793.
- Shibata C, Sasaki I, Naito H, Ohtani N, Matsuno S, Mizumoto A, Iwanaga Y, and Itoh Z (1994) Effects of substance P on gastric motility differ depending on the site and vagal innervation in conscious dogs. Tohoku J Exp Med 174:119-128.
- Sicuteri F, Fanciullacci M, Franchi G, and Michelacci S (1963) The endecapeptide eledoisin is a powerful vasodilating and hypotensive agent in man. Experientia 19:44-47
- Simmaco M, Severini C, De Biase D, Barra D, Bossa F, Roberts J, Melchiorri P, and Erspamer V (1990) Six novel tachykinin- and bombesin-related peptides from the skin of the Australian frog Pseudophryne guntheri. Peptides 11:299-304
- Simon C, Portalier P, Chamoin MC, and Temaux JP (1992) Substance P-likeimmunoreactivity release from enterochromaffin cells of rat caecum mucosa. In-
- hibition by serotonin and calcium-free medium. *Neurochem Int* **20:**529–536. Siyasubramanian P (1990) Substance P-like immunoreactive neurons in the adult nervous system of the fly Sarcophaga bullata. Comp Biochem Physiol C 96:235-
- Skrabanek P, Cannon D, Dempsey J, Kirrane J, Neligan M, and Powell D (1979) Substance P in medullary carcinoma of the thyroid. Experientia 35:1259-1260.
- Skrabanek P, Dervan P, Čannon D, and Powell D (1980) Substance P in ovarian carcinoid. J Clin Pathol 33:160-162.
- Sokolski KN and Lechago J (1984) Human colonic substance P-producing cells are a separate population from the serotonin producing enterochromaffin cells. J Histochem Cytochem 32:1066-1074.
- Stacey RS (1966) Clinical aspects of cerebral and extracerebral 5-hydroxytryptamine, in 5-Hydroxytryptamine and Related Indolealkylamines. Handbook of Exerimental Pharmacology, XIX (Erspamer V ed) pp 745-786, Springer Verlag,
- Berlin-Heidelberg, New York. Starke K, Lembeck F, Lorenz W, and Weiss U (1968) Biliary and pancreatic secretion under the influence of substance P and a physalaemin derivative. Naunyn-Schmiedeberg's Arch Exp Pathol Pharmakol 260:269-274.
- Starr M, James T, and Gaytten D (1978) Behavioural depressant and antinociceptive properties of substance P in the mouse: possible implication of brain monoamines. Eur J Pharmacol **48:**203–212.
- Steiner DF, Smeekens SP, Ohagi S, and Chan SJ (1992) The new enzymology of precursor processing endoproteases. J Biol Chem 267:23435-23438.
- Stewart JM, Getto C, Neldner K, Reece E, Krivoy W, and Zimmermann E (1976) Substance P and analgesia. Nature (Lond) 262:784-785.
- Stewart JM, Hall ME, Harkins J, Frederickson RCA, Terenius L, Hokfelt T, and Krivoy WA (1982) A fragment of substance P with specific central activity: SP(1-7). Peptides 3:851-857.
- Strodel WE, Vinik AI, Jaffe BM, Eckhauser FE, and Thompson NW (1984) Substance P in the localization of a carcinoid tumor. J Surg Oncol 27:106-111.
- Studer ROO, Trzeciak A, and Lergier W (1973) Isolierung und Aminosauren Sequenz von Substanz P aus Pferdedarm. Helv Chim Acta 56:860-866.
- Szam I, Kusztos D, and Csapo G (1966) The effect of eledoisin on the rheogram in arteriosclerosis obliterans. Arzneimittelforschung 16:1671-1673.
- Taban CH and Cathieni M (1979) Localization of substance P-like immunoreactivity in Hydra. Experientia 35:811–812.
- Takano Y, Nakashima A, Hagio T, Tateishi K, and Kamiya H (1990) Role of central
- tachykinin peptides in cardiovascular regulation in rats. Brain Res 528:231–237. Takaori K, Inoue K, Kogire M, Doi R, Sumi S, Yun M, Fujii N, Yajima H, and Tobe T (1989) Effect of synthetic physalaemin on splanchnic circulation in dogs. Life Sci 44:667-672
- Takeuchi H, Matsumoto M, and Mori A (1977a) Modification of effects of biologically active peptides, caused by enzyme treatment, on the excitability of identifiable giant neurones of an African giant snail (Achatina fulica Férussac). Experientia 33:249-251.
- Takeuchi H, Morimasa T, and Matsumoto M (1977b) Inhibitory tripeptide, Lys-Phe-Tyr-, as a fragment of physalaemin. Experientia 33:938-939.
- Takeuchi H and Sakai A (1977) Effects of some oligopeptides, consisting of aromatic amino acids, on the excitability of an identifiable giant neurone of the African giant snail (Achatina fulica Férussac). Experientia 33:1348-1350.
- Takeuchi H, Yokoi I, and Mori A (1976) Effects of physalaemin, a vaso-active peptide

- from amphibian skin, on the excitability of an identified molluscan giant neurone of Achatina fulica Férussac. Experientia 32:606-608.
- Tan DP and Tsou K (1988) Differential effects of tachykinins injected intranigrally on striatal dopamine metabolism. J Neurochem 51:1333–1337.
- Tatemoto K, Lundberg JM, Jornvall H, and Mutt V (1985) Neuropeptide K: isolation, structure and biological activities of a novel brain tachykinin. Biochem Biophys Res Commun 128:947-953.
- Theodorsson-Norheim E, Norheim I, Oberg K, Brodin E, Lundberg JM, Tatemoto K, and Lindgren PG (1985) Neuropeptide K: a major tachykinin in plasma and tumor tissues from carcinoid patients. Biochem Biophys Res Commun 131:77-83.
- Thulin A (1976) Secretory and motor effects in the submaxillary gland of the rat by intraarterial administration of some polypeptides and autonomic drugs. Acta Physiol Scand 97:343-348.
- Thulin L and Holm I (1977) Effect of substance P on the flow of hepatic bile and pancreatic juice, in Substance P (von Euler US and Pernow B eds) pp 247-251, Raven Press. New York.
- Tobin G and Ekstrom J (1992) Parasympathetic NANC-secretion of saliva in the mink and effects of substance P and VIP. Regul Pept **39:**95–101.
- Unger T. Carolus S. Demmert G. Ganten D. Lang RE. Maser-Gluth C. Steinberg H. and Veelken R (1988) Substance P induces a cardiovascular defense reaction in the rat: pharmacological characterization. Circ Res 63:812-820.
- Verhaert P and De Loof A (1985) Substance P-like immunoreactivity in the central nervous system of the blattarian insect Periplaneta americana L. revealed by a monoclonal antibody. Histochemistry 83:501-507.
- Vialli M and Erspamer V (1933) Cellule enterocromaffini e cellule basigranulose acidofile nei Vertebrati. Z Zellforschg mikr Anatomie 19:743-773
- Wahlestedt C (1998) Reward for persistence in substance P research. Science (Wash DC) 281:1624-1625.
- Wallace JL, McCafferty DM, and Sharkey KA (1998) Lack of beneficial effect of a tachykinin receptor antagonist in experimental colitis. Regul Pept 73:95-101.
- Wang Y, Badgery-Parker T, Lovas S, Chartrel N, Vaudry H, Burcher E, and Conlon JM (1992a) Primary structure and receptor-binding properties of a neurokinin A-related peptide from frog gut. Biochem J 287:827-832.
- Wang Y, Barton BA, Thim L, Nielsen PF, and Conlon JM (1999) Purification and characterization of galanin and schyliorhinin I from the hybrid sturgeon Scaphirhynchus platorynchus and Scaphirhynchus albus (Acipenseriformes). Gen Comp Endocrinol 113:38-45
- Wang Y, O'Harte F, and Conlon JM (1992b) Structural characterization of tachykinins (neuropeptide gamma, neurokinin A and substance P) from a reptile Alligator mississipiensis. Gen Comp Endocrinol 88:277-286.
- Waugh D, Bondareva V, Rusakov Y, Bjenning C, Nielsen PF, and Conlon JM (1995a) Tachykinins with unusual structural features from a urodele, the amphiuma, an elasmobranch, the hammerhead shark and an agnathan, the river lamprey. Peptides 16:615-621
- Waugh D, Groff KE, Platzack B, Youson JH, Olson KR, and Conlon JM (1995b) Isolation, localization and cardiovascular activity of tachykinins from the stomach of the bowfin Amia calva. Am J Physiol 269:R565-R571.
- Waugh D, Sower S, Bjenning C, and Conlon JM (1994) Novel tachykinins from the brain of the sea lamprey, Petromyzon marinus and the skate, Raja rhina. Peptides
- Waugh D, Wang Y, Hazon N, Balment RJ, and Conlon JM (1993) Primary structures and biological activities of substance P-related peptides from the brain of the dogfish Scyliorhinus canicula. Eur J Biochem 214:469-474.
- Wilander E, Grimelius L, Portela-Gomes G, Lundquist G, Skoog V, and Westermark P (1979) Substance P and enteroglucagon-like immunoreactivity in argentaffin and argyrophil midgut carcinoid tumours. Scand J Gastroenterol Suppl 53:19-25.
- Wilander E, Portela-Gomes G, Grimelius L, Lundqvist G, and Skoog V (1977) Enteroglucagon and substance P-like immunoreactivity in argentaffin and argyrophil rectal carcinoids. Virchows Arch B Cell Phatol 25:117-124.
- Winther AM, Muren JE, Ahlborg N, and Nassel DR (1999) Differential distribution of isoforms of Leucophaea tachykinin-related peptides (LemTRPs) in endocrine cells and neuronal processes of the cockroach midgut. J Comp Neurol 406:15-28.
- Winther AM, Muren JE, Lundquist CT, Osborne RH, and Nassel DR (1998) Characterization of actions of Leucophaea tachykinin-related peptides (LemTRPs) and proctolin on cockroach hindgut contractions. Peptides 19:445-458.
- Yaksh TL, Jessel TM, Gamse R, Mudge AW, and Leeman SE (1980) Intrathecal morphine inhibits substance P release from mammalian spinal cord in vivo. Nature (Lond) 286:155-157.
- Yamamoto Y and Lagercrantz H (1985) Some effects of substance P on central respiratory control in rabbit pups. Acta Physiol Scand 124:449-455.
- Yanaihara N, Yanaihara C, Hirohashi M, Sato H, Iizuka Y, Hashimoto T, and Sakagami M (1977) Substance P analogs: synthesis and biological and immunological properties, in Substance P (von Euler US and Pernow B eds) pp 27–33, Raven Press, New York.
- Yankner BA, Duffy LK, and Kirschner DA (1990) Neurotrophic and neurotoxic effects of amyloid beta protein: reversal by tachykinin neuropeptides. Science (Wash DC) 250:279-282
- Yasuhara T, Nakajima T, Falconieri Erspamer G, and Erspamer V (1981) New tachykinins Glu2, Pro5-kassinin (Hylambates-kassinin) and hylambatin in the skin of the African rhacophorid frog Hylambates maculatus. Biomed Res 2:613-
- Zhao X, Valantas JA, Vyas S, and Duffy LK (1993) Comparative toxicity of amyloid beta-peptide in neuroblastoma cell lines; effects of albumin and physalaemin. Comp Biochem Physiol C 106:165–170.
- Zhou Q, Liu Z, Huang W, Karlsson K, and Nyberg F (1998) Alteration in the brain content of substance P(1-7) during withdrawal in morphine-dependent rats. Neuropharmacology 37:1545-1552.
- Zimmer A, Zimmer AN, Baffi J, Usdin T, Reynolds K, Konig M, Palkovits M, and Mezey E (1998) Hypoalgesia in mice with a targeted deletion of the tachykinin 1 gene. Proc Natl Acad Sci USA 95:2630-2635.