

Renal Uptake of Circulating Angiotensin II in Val⁵-Angiotensin II Infused Rats Is Mediated by AT₁ Receptor

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Previous studies have demonstrated that augmentation of intrarenal angiotensin II (ANG II) levels during ANG II induced hypertension involves both endogenous formation and accumulation of circulating ANG II. The present work extends these findings and determines whether accumulation of infused ANG II in the kidney requires AT₁ receptor activation by using Val⁵-ANG II as the infused peptide. Male Sprague-Dawley rats were uninephrectomized and divided into three groups: control (n = 6), Val⁵-ANG II (exogenous form) infused (n = 8), and Val⁵-ANG II infused rats treated with losartan (n = 8). Val⁵-ANG II, which has the same biological and immunoreactive properties as endogenous ANG II, was infused at 40 ng/min via an osmotic minipump implanted subcutaneously. By day 12, systolic blood pressure (SBP) increased significantly in Val⁵-ANG II infused rats (197 ± 7 mm Hg). As previously shown, the development of hypertension in ANG II infused rats was

prevented by losartan treatment. Blood and kidney samples were harvested, subjected to HPLC to separate Val⁵-ANG II (exogenous) from Ile⁵-ANG II (endogenous) and the fractions were measured by radioimmunoassay. In the Val⁵-ANG II infused rats treated with losartan, total plasma ANG II levels were elevated to a greater extent than in rats not treated with losartan (289 ± 20 v 119 ± 14 fmol/mL). However, losartan markedly decreased by 88% the enhancement of intrarenal Val⁵-ANG II content that occurred in the rats infused with Val⁵-ANG II alone. These results demonstrate that AT₁ receptor blockade markedly reduces the intrarenal uptake of circulating ANG II that occurs in ANG II induced hypertension. *Am J Hypertens* 1998; 11:570-578 © 1998 American Journal of Hypertension, Ltd.

KEY WORDS: Val⁵-angiotensin II, Ile⁵-angiotensin II, angiotensin II induced hypertension, renin-angiotensin system.

Previous studies have indicated that increased angiotensin II (ANG II) levels in the contralateral, nonclipped kidney may contribute to the pathogenesis of two-kidney, one-clip (2K1C) hypertension.¹⁻³ However, the mechanisms of intrarenal ANG II augmentation remain unclear. Al-

though intrarenal ANG II is augmented in the non-clipped kidney of 2K1C rats, there is marked depletion of renal renin content and reduction of renal renin mRNA levels.^{2,4-8} These findings are consistent with the possibility that there is a non-renin-dependent mechanism responsible for augmenting intrarenal

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ANG II levels in the nonclipped kidney of 2K1C rats. This possibility has received support from studies indicating that chronic infusion of subpressor doses of ANG II into uninephrectomized rats mimics 2K1C hypertension, increases intrarenal ANG II levels, and depletes renal renin content and renin mRNA levels in the remaining kidney, as has been observed in the nonclipped kidney of 2K1C rats.^{2,5,9}

A recent study using Val⁵-ANG II as the infused peptide demonstrated that a substantial fraction of the elevated intrarenal ANG II found in the kidneys consisted of exogenously infused Val⁵-ANG II.¹⁰ These data suggest that the kidney has the capability to take up circulating ANG II into intrarenal sites that protect against degradation and metabolism. Quantitatively, the tissue Val⁵-ANG II contents were much greater than could be explained simply by nonspecific trapping of circulating ANG II. The observation that the AT₁ receptor antagonist, losartan, prevents intrarenal ANG II augmentation during chronic low dose ANG II infusion¹¹ also provides evidence that the renal uptake of ANG II is an active process and does not represent nonspecific intrarenal sequestration of circulating ANG II. In this previous study, heart and adrenal ANG II levels increased in parallel with the plasma ANG II levels during losartan treatment, suggesting that receptor mediated augmentation of renal ANG II is tissue specific. However, the available data did not allow us to determine whether the losartan was preventing uptake of circulating ANG II or reducing intrarenal ANG II formation. The present study was therefore designed to determine the involvement of AT₁ receptors in the renal uptake of circulating ANG II. Rats were infused with Val⁵-ANG II, which is not endogenously produced in the rat but has the same biological and immunoreactive properties as endogenous Ile⁵-ANG II and can be separated by high performance liquid chromatography (HPLC).¹⁰ As previously described,¹⁰ this approach allowed us to distinguish renal uptake of exogenously infused Val⁵-ANG II from that of endogenously formed ANG II. In the present study, the extent to which this process involves AT₁ receptor activation was tested by treating a separate group of Val⁵-ANG II infused rats with the AT₁ receptor antagonist, losartan.

METHODS

Experimental Design Male Sprague-Dawley rats (Charles River, Wilmington, MA) were housed in wire cages and maintained in a temperature and light controlled room. Throughout the experiments, animals had free access to standard rat chow (Ralston-Purina, St. Louis, MO). All experiments were approved by the Tulane University Animal Care and Use Committee. Rats (180 to 200 g body weight) were anesthetized with pentobarbital anesthesia (50 mg/kg intraperito-

neally) and the right kidney was removed. An osmotic minipump (Model 2002, Alza Corp, Palo Alto, CA) containing Val⁵-ANG II was implanted subcutaneously in each rat. Val⁵-ANG II (Novabiochem, San Diego, CA) was delivered continuously at a rate of 40 ng/min. Losartan (Du Pont-Merck Pharmaceutical Co, Wilmington, DE) was administered in the drinking water at a dose of 30 mg/kg/day to allow chronic treatment throughout the period of ANG II infusion. The results obtained in this group were compared to results obtained in other rats prepared in a similar manner including rats infused with vehicle or Val⁵-ANG II but not treated with losartan.¹⁰

Systolic blood pressure (SBP) was measured in conscious rats by tail-cuff plethysmography (Harvard Apparatus, South Natick, MA) to monitor the progression of hypertension. For measurement of plasma renin activity (PRA), plasma and renal ANG I, Ile⁵-ANG II, and Val⁵-ANG II levels, the conscious rats were decapitated on day 13. Approximately 4 mL of trunk blood were collected in about 15 to 20 sec and the kidneys were immediately removed, quickly weighed, and homogenized in methanol. The time delay between decapitation and homogenization of the kidney did not exceed 60 sec.

Measurement of Val⁵-ANG II and Ile⁵-ANG II Levels in Plasma and Kidney

Collection and Extraction of Blood and Kidney Trunk blood was collected in chilled tubes containing a mixed inhibitor solution (5 mmol/L EDTA, 10 μ mol/L pepstatin, 20 μ mol/L enalaprilat, and 1.25 mmol/L 1,10-phenanthroline) and kept at 4°C. To minimize in vitro generation of the peptides, blood samples were immediately centrifuged at 4°C for 10 min at 1000 g. After centrifugation, plasma was separated and immediately extracted by adsorption to and elution from a phenyl bonded, solid phase extraction column (Bond-Elut; Varian, Harbor City, CA). The eluants were collected, evaporated to dryness under vacuum, and then subjected to HPLC to separate Val⁵-ANG II from Ile⁵-ANG II. One-half of each kidney was immersed in cold methanol (100%) and homogenized with a glass homogenizer immediately upon harvest. The supernatants from the kidney homogenates were dried overnight in a vacuum centrifuge. The dried residue was reconstituted in 4 mL assay diluent (50 mmol/L sodium phosphate buffer, pH 7.4, containing 0.1 mg human serum albumin/mL). These samples were extracted and evaporated as described above for plasma, and subjected to HPLC.

Separation of Val⁵-ANG II from Ile⁵-ANG II by HPLC

The HPLC and radioimmunoassay methodology for the measurement of angiotensin peptides has been reported.^{10,12} Briefly, the extract residue from each plasma and kidney sample was redissolved in 150- μ L HPLC column equilibration solvent (35%

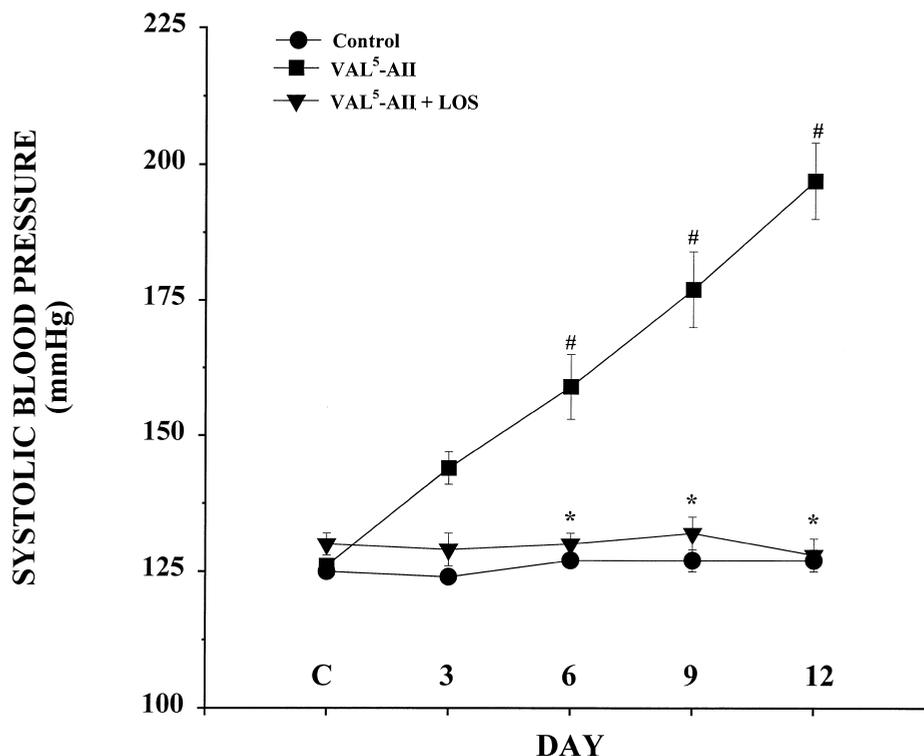


FIGURE 1. Comparison of systolic blood pressure (SBP) in control (circles, $n = 6$), Val⁵-ANG II infused (squares, $n = 8$), and Val⁵-ANG II infused plus losartan treated (triangles, $n = 8$) groups before (control, C) and for 12 days after initiation of ANG II infusions. Values are mean \pm SE. # $P < .05$ v control group; * $P < .05$ v Val⁵-ANG II infused group.

methanol, 65% water, 0.1% H₃PO₄) and chromatographed at 1 mL/min on a 25 \times 0.46 cm, 5 μ m Vydac C18 reversed-phase HPLC column (Separations Group, Hesperia, CA). To shorten the duration of the chromatography run, a combination of isocratic and step-gradient elution modes was used. After 12 min of isocratic elution with the equilibration solvent, the solvent was changed to 40% methanol, 60% water, and 0.1% H₃PO₄ and then the new solvent composition was continued for an additional 18 min. As previously described,¹⁰ Val⁵-ANG II eluted at 6 min (fractions 11 to 14) and Ile⁵-ANG II had an elution peak of 9.5 min (fractions 18 to 22). Fractions were collected every 30 sec, evaporated to dryness, reconstituted in assay diluent, and measured directly by RIA.

Quantitation of Val⁵-ANG II and Ile⁵-ANG II by RIA The reconstituted plasma and kidney fractions were incubated with rabbit anti-ANG II antisera (Arnel, New York, NY), and ¹²⁵I-radiolabeled ANG II (Sigma Chemical Co., St Louis, MO) for 48 h at 4°C. Bound and free ANG peptides were separated by dextran coated charcoal and the supernatants were counted by a computer linked gamma counter for 3 min. As previously shown, the immunoreactivity of the antibodies for Val⁵-ANG II and Ile⁵-ANG II were virtually

identical. Results are reported in femtomoles/gram kidney weight or femtomoles/milliliter plasma. The sensitivity of the ANG II assay was 1.46 ± 0.33 fmol. For the ANG II assays, the specific binding was $43.4\% \pm 2.3\%$, with a nonspecific binding of $1.3\% \pm 0.1\%$.

Plasma Renin Activity and ANG I Assays For renin determination, trunk blood was collected in chilled tubes containing EDTA (5 mmol/L). Plasma was separated and stored at -20°C until assayed using a commercially available ANG I RIA kit (Incstar, Stillwater, MN) as described previously.² For ANG I measurement, blood and kidney samples were collected, extracted, and quantitated by RIA as reported previously.²

Data Analysis All data are presented as mean \pm SEM. The statistical analyses for plasma and kidney levels were performed using the one way analysis of variance (ANOVA) and Fisher's least significant difference (FLSD) post hoc test. Differences between and within groups for systolic blood pressure measurements were analyzed by two way ANOVA with repeated measures on one factor and the FLSD post hoc test. A value of $P < .05$ was considered statistically significant.

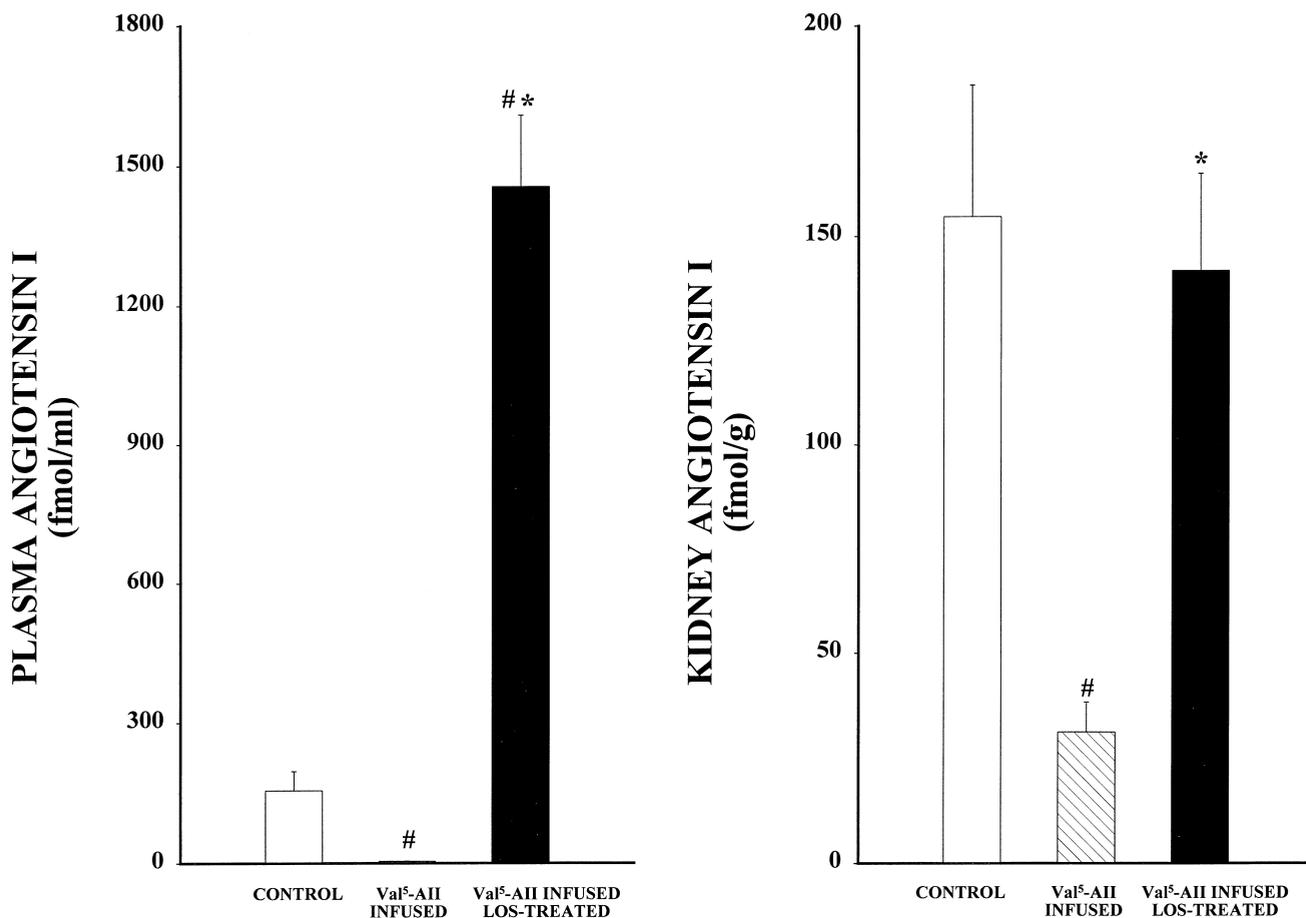


FIGURE 2. Comparison of plasma and kidney angiotensin I levels in control (open bar, $n = 6$), Val⁵-ANG II infused (striped bar, $n = 8$), and Val⁵-ANG II infused plus losartan treated (solid bar, $n = 8$) groups. Values are mean \pm SE. #P < .05 v control group; *P < .05 v Val⁵-ANG II infused group.

RESULTS

Influence of Losartan on Arterial Pressure During Val⁵-ANG II Infusion (Figure 1) Before surgical manipulations, SBP values were normotensive with an average of 127 ± 1 mm Hg. As previously shown,¹⁰ SBP in the Val⁵-ANG II infused rats exhibited progressive increases over a 12 day period reaching a value of 197 ± 7 mm Hg at 12 days. This increase was similar to or slightly greater than that seen in rats infused with Ile⁵-ANG II.¹⁰ As shown for rats infused with Ile⁵-ANG II,¹¹ losartan treatment prevented the development of hypertension in the Val⁵-ANG II infused group and SBP remained at normal levels for the duration of the study.

Effects of Losartan on Plasma Renin and Plasma and Kidney ANG I During Val⁵-ANG II Infusions As previously reported,¹⁰ in vehicle infused controls, PRA averaged 4.96 ± 1.17 ng ANG I/mL/h, and the Val⁵-ANG II infused rats had almost complete suppression of PRA with values of 0.21 ± 0.06 ng ANG

I/mL/h. In contrast, losartan treatment in the Val⁵-ANG II infused group exhibited a marked sevenfold increase in PRA at 13 days (36.35 ± 5.24 ng ANG I/mL/h).

As previously reported,¹⁰ Plasma and kidney ANG I levels in the Val⁵-ANG II infused rats were reduced markedly. The rats receiving losartan treatment, however, showed a ninefold increase in plasma ANG I. Nevertheless, intrarenal ANG I content was restored only to that found in control rats but was not significantly elevated beyond these values (Figure 2).

Effects of Losartan on Plasma and Kidney ANG II During Val⁵-ANG II Infusions As shown in Figure 3 and previously reported,¹⁰ total plasma ANG II levels were elevated in the Val⁵-ANG II infused group compared with controls. About half of the plasma ANG II was in the form of Val⁵-ANG II, whereas Ile⁵-ANG II levels were maintained at concentrations similar to those found in control rats in spite of marked renin depletion and reduction in ANG I con-

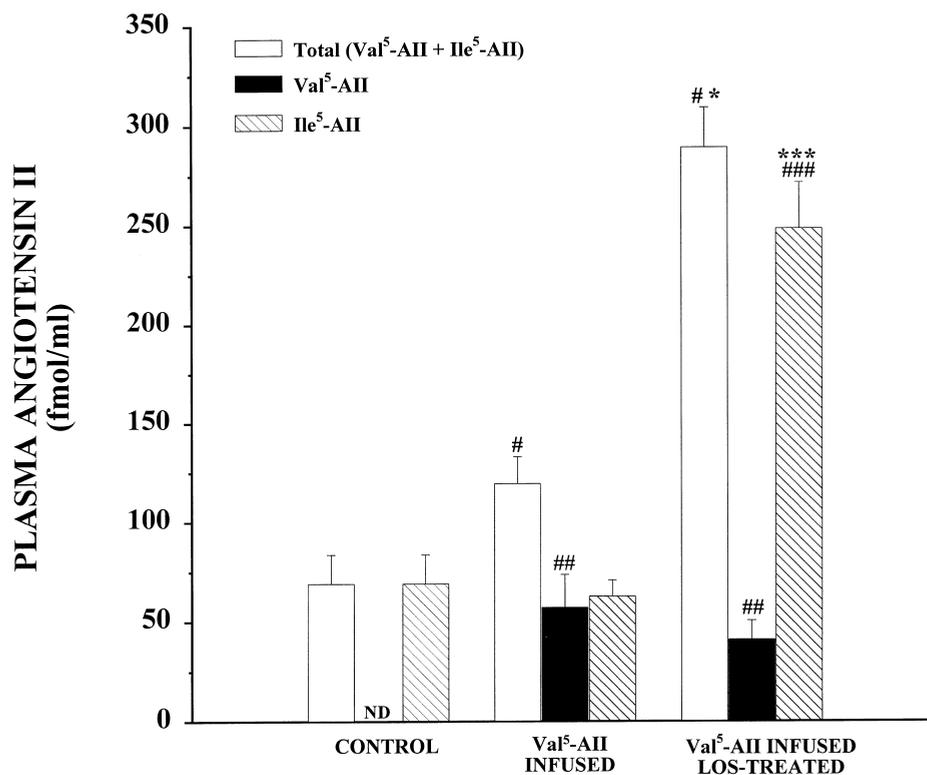


FIGURE 3. Comparison of plasma Ile⁵-ANG II (striped bar) and Val⁵-ANG II (solid bar) levels in control ($n = 6$), Val⁵-ANG II infused ($n = 8$), and Val⁵-ANG II infused plus losartan treated ($n = 8$) groups. Values are mean \pm SE. # $P < .05$ v total ANG II levels of controls; ## $P < .05$ v Val⁵-ANG II levels of controls; ### $P < .05$ v Ile⁵-ANG II levels of controls; * $P < .05$ v total ANG II levels of Val⁵-ANG II infused group; *** $P < .05$ v Ile⁵-ANG II levels of Val⁵-ANG II infused group.

centration. Losartan treatment caused substantial increases in total plasma ANG II levels in the Val⁵-ANG II infused group. Total ANG II levels were 2.4-fold higher than in rats infused with Val⁵-ANG II but not treated with losartan and fourfold higher than in the control group. A 3.6-fold elevation of Ile⁵-ANG II was responsible for the elevated plasma ANG II levels, whereas the plasma Val⁵-ANG II levels were not influenced by losartan treatment.

The intrarenal ANG II contents, however, showed a distinctly different pattern (Figure 4). As already reported,¹⁰ Val⁵-ANG II infused rats exhibited a marked increase in total intrarenal ANG II content (371 ± 57 v 116 ± 11 fmol/g controls). Interestingly, 70% of total intrarenal ANG II in the Val⁵-ANG II infused rats was in the form of Val⁵-ANG II and renal Val⁵-ANG II content in the kidney was 256 ± 44 fmol/g, which greatly exceeded plasma concentrations of 57 ± 17 fmol/mL. The ratio of kidney to plasma Val⁵-ANG II is 6.78 ± 1.73 (Figure 5). Additionally, intrarenal Ile⁵-ANG II contents in the Val⁵-ANG II infused rats were maintained compared with controls even though plasma renin activity and plasma and renal ANG I levels were markedly suppressed. In contrast to the changes observed for plasma ANG II levels, total in-

trarenal ANG II contents were decreased in the losartan treated rats. In particular, losartan treatment markedly reduced renal Val⁵-ANG II content by 88% but did not significantly decrease renal Ile⁵-ANG II contents in the Val⁵-ANG II infused rats. The ratio of kidney to plasma Ile⁵-ANG II was used as a reflection of the degree of intrarenal ANG II accumulation relative to the plasma levels. In the Val⁵-ANG II infused rats this ratio was significantly reduced by losartan from 2.18 ± 0.49 to 0.37 ± 0.03 (Figure 5). Thus, the intrarenal levels of Ile⁵-ANG II did not increase concomitantly with the plasma levels and were significantly reduced to ratios below 1 in the losartan treated rats, indicating that the intrarenal contents of Ile⁵-ANG II could be attributed primarily to the extracellular fluid concentration.

DISCUSSION

Previous studies demonstrating that intrarenal ANG II levels are augmented during ANG II-induced hypertension.¹⁻² have led to the hypothesis that chronic increases in circulating ANG II lead to progressive augmentation of intrarenal ANG II content. Our recent observation that 70% of the elevated intrarenal ANG II found in the Val⁵-ANG II infused rats was derived

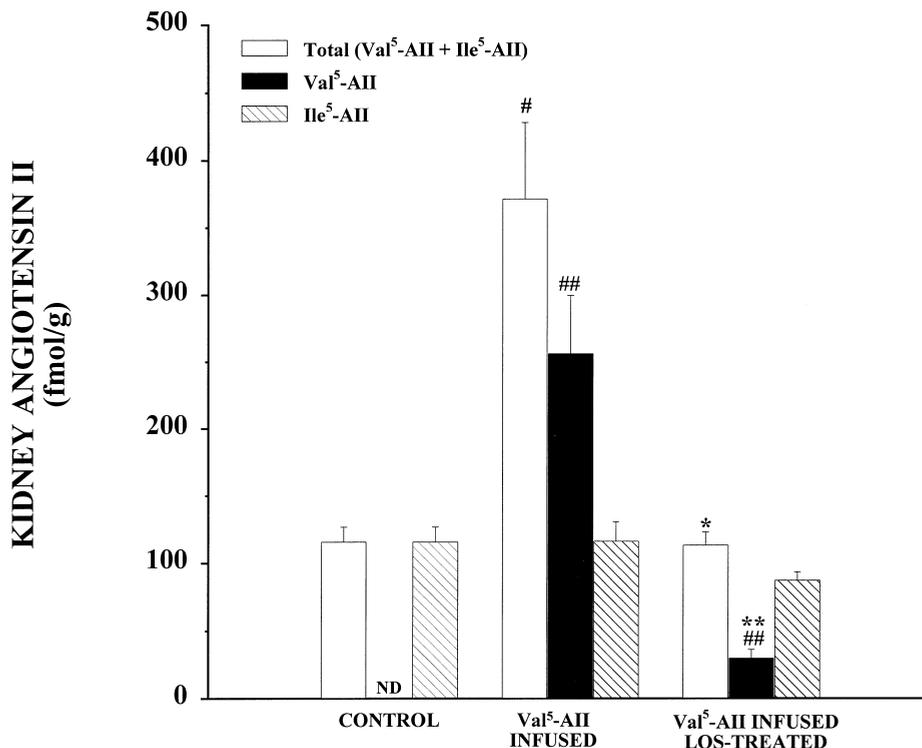


FIGURE 4. Comparison of kidney Ile⁵-ANG II (striped bar) and Val⁵-ANG II (solid bar) levels in control ($n = 6$), Val⁵-ANG II infused ($n = 8$), and Val⁵-ANG II infused plus losartan treated ($n = 8$) groups. Values are mean \pm SE. #P < .05 v total ANG II levels of controls; ##P < .05 v Val⁵-ANG II levels of controls; ###P < .05 v Ile⁵-ANG II levels of controls; *P < .05 v total ANG II levels of Val⁵-ANG II infused group; **P < .05 v Val⁵-ANG II levels of Val⁵-ANG II infused group.

from the exogenously infused Val⁵-ANG II¹⁰ indicates that augmentation of intrarenal ANG II is due, in large part, to uptake of circulating ANG II. In addition, the finding that endogenous Ile⁵-ANG II is maintained in the kidneys of Val⁵-ANG II infused rats in a setting where renin and ANG I levels are markedly reduced¹⁰ suggests that endogenous production of ANG II is maintained under conditions of renin depletion. Additional recent results¹¹ demonstrate that intrarenal ANG II augmentation involves activation of AT₁ receptors, which may be responsible for renal uptake of ANG II or renal enhanced formation of ANG II. By using a different form of ANG II in the minipumps, the current study extends previous findings and specifically demonstrates the role of AT₁ receptor activation in mediating renal uptake of ANG II during ANG II infusion.

Losartan treatment to rats infused with Val⁵-ANG II for 13 days prevented the progressive increases in systolic blood pressure and markedly reduced the intrarenal augmentation of ANG II. As expected, the PRA levels, which were almost completely suppressed in the Val⁵-ANG II infused rats, were markedly elevated during losartan treatment. These results are consistent with an important role of AT₁ receptor activation in the ANG II-induced negative feedback effect

on renin release.^{2,5,13-16} Chronic administration of losartan was also associated with elevated plasma ANG I and ANG II levels, which can be accounted for by the marked elevation of circulating renin activity leading to increased formation of ANG I from angiotensinogen and conversion to Ile⁵-ANG II.

The significant finding of this study is that losartan treatment markedly diminished the increase in intrarenal Val⁵-ANG II content in the Val⁵-ANG II infused rats. These studies indicate that intrarenal ANG II augmentation is due, in large part, to AT₁ receptor mediated binding and internalization of circulating ANG II. These data are consistent with the findings by Anderson and Peach,¹⁷ who demonstrated that ANG II is internalized via AT₁ receptors and that losartan can block ANG II internalization in cultured explant derived rat aortic vascular smooth muscle cells. Likewise, Wang et al¹⁸ reported receptor mediated internalization of ANG II in primary cultures of bovine adrenal medullary chromaffin cells. Moreover, Ullian and Linas¹⁹ showed the importance of receptor cycling in the regulation of angiotensin II surface receptor number and angiotensin II uptake in rat vascular smooth muscle cells. Our present observation provides complementary *in vivo* data that the ANG II receptor complex is internalized in the kidney. This

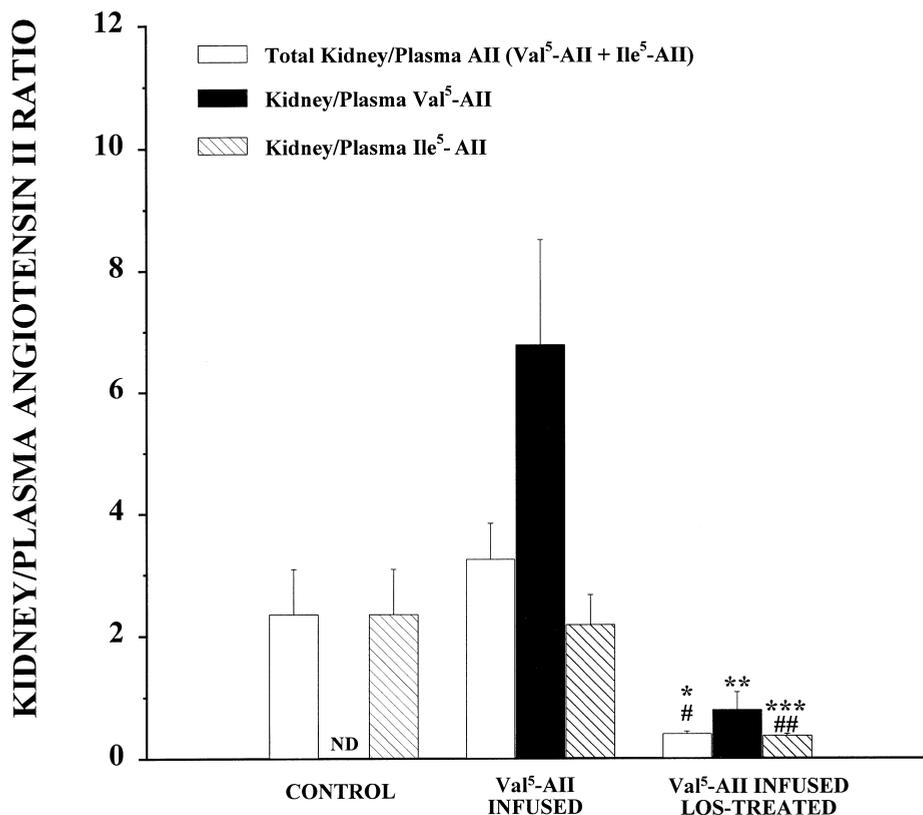


FIGURE 5. Comparison of ratio of kidney ANG II to plasma ANG II levels in control ($n = 6$), Val⁵-ANG II infused ($n = 8$), and Val⁵-ANG II infused plus losartan treated ($n = 8$) groups. Values are mean \pm SE. #P < .05 v total kidney/plasma ANG II ratio of controls; ##P < .05 v kidney/plasma Ile⁵-ANG II ratio of controls; *P < .05 v total kidney/plasma ANG II ratio of Val⁵-ANG II infused rats; **P < .05 v kidney/plasma Val⁵-ANG II ratio of Val⁵-ANG II infused rats; ***P < .05 v kidney/plasma Ile⁵-ANG II ratio of Val⁵-ANG II infused rats.

possibility has received support from several studies showing that binding of ANG II to its receptor initiates internalization of the complex in hepatoma cells,²⁰ adrenocortical cells,²¹ and vascular smooth muscle cells.^{17,22} However, these previous studies did not determine whether the internalized ANG II was immediately degraded or accumulated. The approach used in the current study allowed us to detect increased amounts of intrarenal ANG II suggesting that some of the internalized ANG II is protected from degradation and is possibly used for some intracellular process. This hypothesis is supported by results from experiments in isolated or cultured cells showing intracellular localization of ANG II and ANG II specific binding sites.^{17,23–28}

Although the present studies demonstrate that receptor mediated internalization of ANG II occurs in the kidney, the specific cells responsible for renal internalization of ANG II remain undetermined. It is now clear that AT₁ receptors are abundant on rat proximal tubules.²⁹ Poggioli et al²⁵ provided further evidence by showing that AT₁ receptors are predominant in intact rat proximal tubule cells and are cou-

pled to both IP₃-Ca²⁺ and cAMP signaling pathways. Douglas and Hopfer²⁸ also found that AT₁ receptors of proximal tubular epithelial cells are linked to a complex combination of signal transduction pathways. In addition, it has been found that ANG II leads to increases in proximal tubule AT₁ receptor mRNA and receptor binding.³⁰ In regard to the functional properties of receptor mediated ANG II internalization, Schelling and Linas³¹ reported that ANG II dependent proximal tubule sodium transport involves receptor mediated endocytosis. Becker and Harris³² using LL-CPK epithelial cells demonstrated that ANG II binding to the apical AT₁ receptor and subsequent endocytosis is required for PLA₂ activation. In preliminary studies ANG II stimulated Na⁺ flux was inhibited by the calcium independent PLA₂ inhibitor HELSS, raising the possibility that internalization of the ANG II AT₁ receptor complex is required for full expression of biological activity.³² Moreover, a large body of studies in vivo have indicated that the blockade of AT₁ receptors by losartan is associated with decreases in fractional proximal reabsorption and proximal tubular transport.^{33–34} Collectively, these findings suggest that

renal proximal tubular cells may be involved in receptor mediated ANG II internalization and receptor mediated cellular signal transduction.

Because losartan also prevented the development of hypertension during the 2 weeks of ANG II infusion, it is also possible that the renal ANG II uptake in the ANG II infused rats may be secondary to renal injury occurring as a consequence of hypertension.³⁵ The ANG II infused rats clearly develop extensive renal injury to the vasculature, glomeruli, tubules, and interstitium.^{11,35} Thus, renal injury could serve as a stimulus for both ANG II uptake and intrarenal ANG II formation. Because losartan prevented hypertension, it is also possible that the reduction in intrarenal ANG II content in the losartan treated rats was due, in part, to the prevention of hypertension. At present, the relative contributions of the hypertension associated renal injury versus the increased circulating ANG II concentrations to the increased intrarenal uptake of ANG II cannot be established. However, it was recently shown that the increased intrarenal ANG II levels still occurred to the same extent in ANG II infused rats subjected to renal denervation, which markedly attenuated the magnitude of the hypertension.³⁶

The current data also showed that losartan treatment did not alter intrarenal Ile⁵-ANG II contents in the Val⁵-ANG II infused rats. Although this would seem to suggest that intrarenal handling of Ile⁵-ANG II was not influenced by losartan, it should be noted that losartan stimulated renin release and markedly increased the circulating ANG II concentrations. Thus, when considered as the ratio of intrarenal content to circulating ANG II concentration, it can be noted that the relative intrarenal content of Ile⁵-ANG II was reduced because it was a much smaller fraction of circulating Ile⁵-ANG II during losartan treatment than before. In all likelihood, losartan also prevented receptor mediated internalization of endogenous ANG II but it is difficult to determine how much of the intrarenal Ile⁵-ANG II was formed de novo as opposed to being derived from circulating ANG II.

In summary, the results of the present study indicate that losartan markedly reduced renal Val⁵-ANG II contents in the Val⁵-ANG II infused rats. These results indicate that circulating ANG II is bound and taken up into kidney cells primarily through an AT₁ receptor mediated process, which may play a major role in augmenting intrarenal ANG II levels in conditions of ANG II dependent hypertension caused by elevated circulating ANG II concentrations.

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REFERENCES

- Guan S, Fox J, Mitchell KD, Navar LG: Angiotensin and angiotensin converting enzyme tissue levels in two-kidney, one-clip hypertensive rats. *Hypertension* 1992; 20:763–767.
- Von Thun AM, Vari RC, El-Dahr SS, Navar LG: Augmentation of intrarenal angiotensin II levels by chronic angiotensin II infusion. *Am J Physiol* 1994;266:F120–F128.
- Navar LG, Von Thun AM, Zou LX, et al: Enhancement of intrarenal angiotensin II levels in 2 kidney 1 clip and angiotensin II induced hypertension. *Blood Press* 1995; 4(suppl 2):88–92.
- El-Dahr SS, Dipp S, Guan S, Navar LG: Renin, angiotensinogen and kallikrein gene expression in two kidney Goldblatt hypertensive rats. *Am J Hypertens* 1993; 6:914–919.
- Von Thun AM, El-Dahr SS, Vari RC, Navar LG: Modulation of renin-angiotensin and kallikrein gene expression in experimental hypertension. *Hypertension* 1994; 23:1131–1136.
- Moffett RB, McGowan RA, Gross KW: Modulation of kidney renin messenger RNA levels during experimentally induced hypertension. *Hypertension* 1986;8:874–882.
- Morishita R, Higaki J, Okunishi H, et al: Changes in gene expression of the renin-angiotensin system in two-kidney, one-clip hypertensive rats. *J Hypertens* 1991;9:187–192.
- Samani NJ, Godfrey NP, Major JS, et al: Kidney renin mRNA levels in the early and chronic phases of two-kidney, one-clip hypertension in the rat. *J Hypertens* 1989;7:105–112.
- van der Mark J, Kline RL: Altered pressure natriuresis in chronic angiotensin II hypertension in rats. *Am J Physiol* 1994;266:R739–R748.
- Zou LX, Hymel A, Imig JD, Navar LG: Renal accumulation of circulating angiotensin II in angiotensin II-infused rats. *Hypertension* 1996;27(part 2):658–662.
- Zou LX, Imig JD, Von Thun AM, et al: Receptor mediated intrarenal angiotensin II augmentation in angiotensin II infused rats. *Hypertension* 1996;28:669–677.
- Fox J, Guan S, Hymel AA, Navar LG: Dietary Na and ACE inhibition effects on renal tissue angiotensin I and II and ACE activity in rats. *Am J Physiol* 1992;262:F902–F909.
- Barrett GL, Morgan TO, Smith M: Influence of angiotensin II (AII) and angiotensin I (AI) on renal renin synthesis and release. *Clin Exp Hypertens [A]* 1988;10: 1297–1282.
- Johns DW, Peach MJ, Gomez RA, et al: Angiotensin II regulates renin gene expression. *Am J Physiol* 1990;259: F882–F887.
- Nakamura A, Iwao H, Fukui K, et al: Regulation of liver angiotensinogen and kidney renin mRNA levels by angiotensin II. *Am J Physiol* 1990;258:E1–E6.
- Schunkert H, Ingelfinger JR, Jacob H, et al: Reciprocal feedback regulation of kidney angiotensinogen and renin mRNA expressions by angiotensin II. *Am J Physiol* 1992;263:E863–E869.

17. Anderson KM, Peach MJ: Receptor binding and internalization of a unique biologically active angiotensin II-colloidal gold conjugate: morphological analysis of angiotensin II processing in isolated vascular strips. *J Vasc Res* 1994;31:10–17.
18. Wang JM, Llona I, De Potter WP: Receptor-mediated internalization of angiotensin II in bovine adrenal medullary chromaffin cells in primary culture. *Regul Pept* 1994;53:77–86.
19. Ullian ME, Linas SL: Role of receptor cycling in the regulation of angiotensin II surface receptor number and angiotensin II uptake in rat vascular smooth muscle cells. *J Clin Invest* 1989;84:840–846.
20. Schwartz AL, Fridovitch SE, Lodish HF: Kinetics of internalization and recycling of the asialoglycoprotein receptor in a hepatoma cell line. *J Biol Chem* 1982;257:4230–4237.
21. Crozat A, Penhoat A, Saze JM: Processing of angiotensin II (A-II) and (Sar¹,Ala⁸) A-II by cultured bovine adrenocortical cells. *Endocrinology* 1986;118:232–2318.
22. Griendling KK, Delafontaine P, Rittenhouse SE, et al: Correlation of receptor sequestration with sustained diacylglycerol accumulation in angiotensin II-stimulated cultured vascular smooth muscle cells. *J Biol Chem* 1987;262:14555–14562.
23. Eggena P, Zhu JH, Clegg K, Barrett JD: Nuclear angiotensin receptors induce transcription of renin and angiotensinogen mRNA. *Hypertension* 1993;22:496–501.
24. Hunyady L, Tian Y, Sandberg K, et al: Divergent conformational requirements for angiotensin II receptor internalization and signaling. *Kidney Int* 1994;46:1496–1498.
25. Poggioli J, Lazar G, Houillier P, et al: Effect of angiotensin II and nonpeptide receptor antagonists on transduction pathways in rat proximal tubule. *Am J Physiol* 1992;263:C750–C758.
26. Chaki S, Guo DF, Yamano Y, et al: Role of carboxyl tail of the rat angiotensin II type 1A receptor in agonist-induced internalization of the receptor. *Kidney Int* 1994;46:1492–1495.
27. Jan DA: Nuclear signaling pathways for polypeptide ligands and their membrane receptors. *FASEB J* 1994;8:841–847.
28. Douglas JG, Hopfer U: Novel aspect of angiotensin receptors and signal transduction in the kidney. *Annu Rev Physiol* 1994;56:649–669.
29. Zhuo J, Alcorn D, Harris PJ, Mendelsohn FAO: Localization and properties of angiotensin II receptors in rat kidney. *Kidney Int* 1993;44(suppl 42):S40–S46.
30. Cheng HF, Becker BN, Burns KD, Harris RC: Angiotensin II upregulates type-1 angiotensin II receptors in renal proximal tubule. *J Clin Invest* 1995;95:2012–2019.
31. Schelling JR, Linas SL: Angiotensin II-dependent proximal tubule sodium transport requires receptor-mediated endocytosis. *Am J Physiol* 1994;266:C669–C675.
32. Becker BN, Harris RC: A potential mechanism for proximal tubule angiotensin II-mediated sodium flux associated with receptor-mediated endocytosis and arachidonic acid release. *Kidney Int* 1996;50(suppl 57):S66–S72.
33. Zhuo J, Thomas D, Harris PJ, Skinner SL: The role of endogenous angiotensin II in regulation of renal hemodynamics and proximal fluid reabsorption. *J Physiol* 1992;453:1–13.
34. Xie MH, Liu FY, Wang PC, et al: Proximal nephron and renal effects of DuP 753, a nonpeptide angiotensin II receptor antagonist. *Kidney Int* 1990;38:473–479.
35. Johnson RJ, Alpers CE, Yoshimura A, et al: Renal injury from angiotensin II-mediated hypertension. *Hypertension* 1992;19:464–474.
36. Ichihara A, Inscho EW, Imig JD, et al: Role of renal nerves in afferent arteriolar reactivity in angiotensin-induced hypertension. *Hypertension* 1997;29(part 2):442–449.