

Lysosomotropic drugs inhibit maturation of transforming growth factor- β

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Abstract: Transforming growth factor- β (TGF β) is synthesized as a precursor protein, pro-TGF β , that must be cleaved by a furin-like proteinase before it becomes biologically active. We hypothesized that alkalization of the trans-Golgi network (TGN)/endosome system may suppress pro-TGF β processing and decrease TGF β secretion. This hypothesis was tested in human A549 alveolar epithelial and T98G glioblastoma cell lines and in C57BL/6 mice. Inhibition of furin-like activity with decanoyl-RVVR chloromethylketone suppressed pro-TGF β processing, thereby significantly reducing the levels of secreted TGF β . Brefeldin A, bafilomycin A1, ammonium chloride, and monensin also prevented pro-TGF β processing. The alkalizing lysosomotropic drugs chloroquine, hydroxychloroquine, amodiaquine, and azithromycin had a similar effect on the overall production of mature bioactive TGF β . Reduced levels of secreted TGF β were also associated with a decrease in Smad2 signaling. Mice treated with chloroquine showed a decrease in bronchoalveolar lavage fluid TGF β . We conclude that alkalizing lysosomotropic drugs inhibit pro-TGF β processing.

Key words: idiopathic pulmonary fibrosis, lysosomotropic drugs, glioblastoma, furin, epithelial cells, convertases.

Résumé : Le facteur de croissance transformant β (TGF β) est synthétisé en tant que protéine en amont : le pro-TGF β qui doit être scindé par une protéinase analogue à la furine avant de devenir biologiquement actif. Nous posons donc l'hypothèse selon laquelle l'alcalinisation de l'ensemble réseau trans-golgien/endosome stoppe la production de pro-TGF β et diminue la sécrétion de TGF β . Nous avons testé cette hypothèse dans une cellule épithéliale d'une alvéole A549, dans une lignée cellulaire de glioblastome T98G et chez des souris C57BL/6. L'inhibition de l'activité analogue à celle de la furine par la chlorométhylcétone de decanoyl-RVVR arrête la production de pro-TGF β et cause une diminution significative de la concentration de TGF β sécrétée. La brefeldine A, la bafilomycine A1, le chlorure d'ammonium et la monensine arrêtent aussi la production de pro-TGF β . La chloroquine, l'hydroxychloroquine, l'amodiaquine et l'azithromycine ont aussi un effet similaire sur la production globale de TGF β bioactif arrivé à terme. La diminution de la sécrétion de TGF β est aussi associée au fléchissement de la signalisation de Smad-2. Les souris traitées à la chloroquine révèlent une diminution de la concentration de TGF β dans le liquide de lavage bronchoalvéolaire. En conclusion, les drogues lysosomotropiques au pouvoir alcalinisant arrêtent la production de pro-TGF β .

Mots-clés : fibrose pulmonaire idiopathique, drogues lysosomotropiques, glioblastome, furine, cellules épithéliales, convertases.

[Traduit par la Rédaction]

Introduction

Transforming growth factor- β (TGF β) plays a central role in the pathophysiology of several diseases, particularly those associated with tissue fibrosis and neoplastic growth (Blobe

et al. 2000). Three isoforms of TGF β are relevant to human biology: TGF β 1, TGF β 2, and TGF β 3 (Sheppard 2006). Increased expression of TGF β 1 occurs in the alveolar epithelial cells of patients with idiopathic pulmonary fibrosis (IPF), whereas TGF β 1 and TGF β 2 are expressed in glioblastoma cells (de Martin et al. 1987; Khalil et al. 1991; Leitlein et al. 2001). Each TGF β isoform is encoded by a distinct gene encoding a large proprotein that is cleaved by the serine protease furin or a member of the proprotein convertase (PC) family (Dubois et al. 1995).

Furin (EC 3.4.21.75) is a ubiquitous subtilisin-like proprotein convertase that cleaves the TGF β proprotein to release the N-terminal propeptide known as the latency-associated peptide (LAP) and TGF β (Dubois et al. 1995, 2001; Thomas 2002). Other convertases of the PC family may also act as pro-TGF β convertases. Once pro-TGF β is cleaved, disulfide-linked dimers of LAP and TGF β associate noncovalently to form the small latency complex, which can be activated by proteases, oxidants, and other mechanisms (Sheppard 2006). Pro-TGF β , TGF β , and LAP can each be detected in the extracellular milieu of lung alveolar epithelial cells and glioblastoma cells, where activated TGF β

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binds to its receptor and initiates a signaling cascade that phosphorylates the Smad proteins leading to gene transcription (Massague et al. 2005).

Furin and the TGF β proprotein colocalize to the trans-Golgi network (TGN)/endosomal compartment, a mildly acidic compartment, where endoprotease cleavage of furin-like substrates has been shown to occur (Anderson et al. 2002). Since cleavage occurs in the TGN and since furin is active (or activated) in a mildly acidic compartment, we hypothesized that alkalinizing lysosomotropic drugs such as chloroquine and amodiaquine, which are antimalarial agents, hydroxychloroquine, an antirheumatic agent, and azithromycin, an antibiotic, can inhibit TGF β maturation by preventing PC-dependent proprotein maturation. This hypothesis was verified in relevant cell lines derived from human alveolar epithelium (A549) and glioblastoma tumor (T98G), as well as in C57BL/6 mice.

Materials and methods

Cell lines and reagents

The A549 pulmonary epithelial and the T98G glioblastoma cell lines were obtained from the American Type Culture Collection (Rockville, Md.), ATCC Nos. CCL 185 and CRL 1690, respectively. The A549 and T98G cells were grown in 100 millimetre petri dishes or in 6-well culture plates (Linbro Chemical, New Haven, Conn.) in RPMI medium (Gibco, Invitrogen Life Technologies, Burlington, Ont.) supplemented with 10% fetal bovine serum (FBS) and 2 mmol/L glutamine in 5% CO₂ at 37 °C. The A549 and T98G cells were trypsinised and seeded at a density of 1.5×10^6 cells/well in 6-well culture plates and incubated for 24 h before each experiment. The furin inhibitor decanoyl-Arg-Val-Lys-Arg-chloromethylketone (RVKRcmk) and the control peptide decanoyl-RVRK-chloromethylketone (RVRKcmk) were from Bachem (Torrance, Calif.). Brefeldin A, monensin, ammonium chloride, bafilomycin A1, azithromycin, amodiaquine, chloroquine, and hydroxychloroquine were purchased from Sigma-Aldrich (St. Louis, Mo.).

TGF β ELISA assay

A549 and T98G cells were incubated in 6-well culture plates in RPMI without FBS for 24 h either alone or with the following: 10 μ mol/L RVKRcmk, 10 μ mol/L RVRKcmk, 1 μ mol/L brefeldin A, 1 μ mol/L bafilomycin A1, 10 mmol/L NH₄Cl, 40 mmol/L NH₄Cl, and 1 μ mol/L monensin. In addition, both cell lines were incubated in RPMI without FBS for 24 h in the presence of 10, 50, 100, and 200 μ mol/L chloroquine, and A549 was also incubated with azithromycin, amodiaquine, and hydroxychloroquine. To determine the kinetics of chloroquine effects on TGF β 1 production, A549 cells were incubated with or without 100 μ mol/L chloroquine for 0.25, 0.5, 1, 2, 4, 12, and 24 h. After incubations, the cell supernatants were collected and treated with HCl to convert all mature TGF β 1 (that is, both active and inactive) into a form recognized by the antibody used in the assay. The concentration of HCl-treated human TGF β 1 was measured by an ELISA for human TGF β 1 (R&D Systems, Minneapolis, Minn.). It is important to note that this assay recognizes mature (active and inactive) TGF β but does not detect pro-TGF β .

Western blot analysis

Cell supernatants were concentrated on Amicon 5000 (Millipore, Billerica, Mass.) for 15 min for TGF β 1 Western blot analysis. To obtain cell lysates for phosphorylated Smad2 Western blot analysis, the cells were washed twice with PBS and then lysed in 0.5% (w/v) sodium deoxycholate (Sigma-Aldrich), 0.1% (w/v) sodium dodecyl sulfate (SDS) (Sigma-Aldrich), 1% (v/v) nonionic detergent (Igepal CA-630, Sigma-Aldrich), and protease inhibitor cocktail (1 tablet/50 mL) (Roche, Mannheim, Germany) before being placed on ice for 10 min. The lysates were centrifuged at 13 000g for 10 min at 4 °C, and the supernatants collected. The protein concentrations of the cell supernatants and lysates were estimated by using the Bio-Rad protein assay kit. Samples containing 75 μ g protein of cell lysates or 5 μ g protein of supernatant were resolved on 8% SDS – polyacrylamide gels. The separated proteins were electrotransferred to a nitrocellulose membrane (Bio-Rad Laboratories, Mississauga, Ont.) and analyzed by Western blotting procedures, using 5% (w/v) bovine serum albumin (BSA) as the blocking agent, and overnight incubation at 4 °C with goat anti-LAP (1:250, R&D Systems) or rabbit anti-Smad2 (pSer465/467) (1:1000, Calbiochem, San Diego, Calif.) polyclonal antibodies. The membranes were then washed and incubated 1 h with anti-rabbit or anti-goat IgG horseradish peroxidase-conjugated (1:3000, Calbiochem) and revealed by chemiluminescence (ECL kit, Amersham, Buckinghamshire, UK).

Chloroquine in mice

C57BL/6 mice (Charles River, St. Constant, Qué.) weighing 25 g were housed in accordance with guidelines from the Canadian Council on Animal Care. The protocol was approved by the institutional animal care committee of the University of Sherbrooke. Animals were treated with daily subcutaneous injections of 50 μ L 0.9% saline or 50 μ L of 20 mg/mL chloroquine for 7 days. On day 7, bronchoalveolar lavage (BAL) was performed with 3 aliquots (0.9 mL each) of 0.9% saline. BAL fluid was centrifuged, and the supernatant collected to measure TGF β 1 by ELISA and proteins with the Bradford protein assay (Bradford 1976).

Statistics

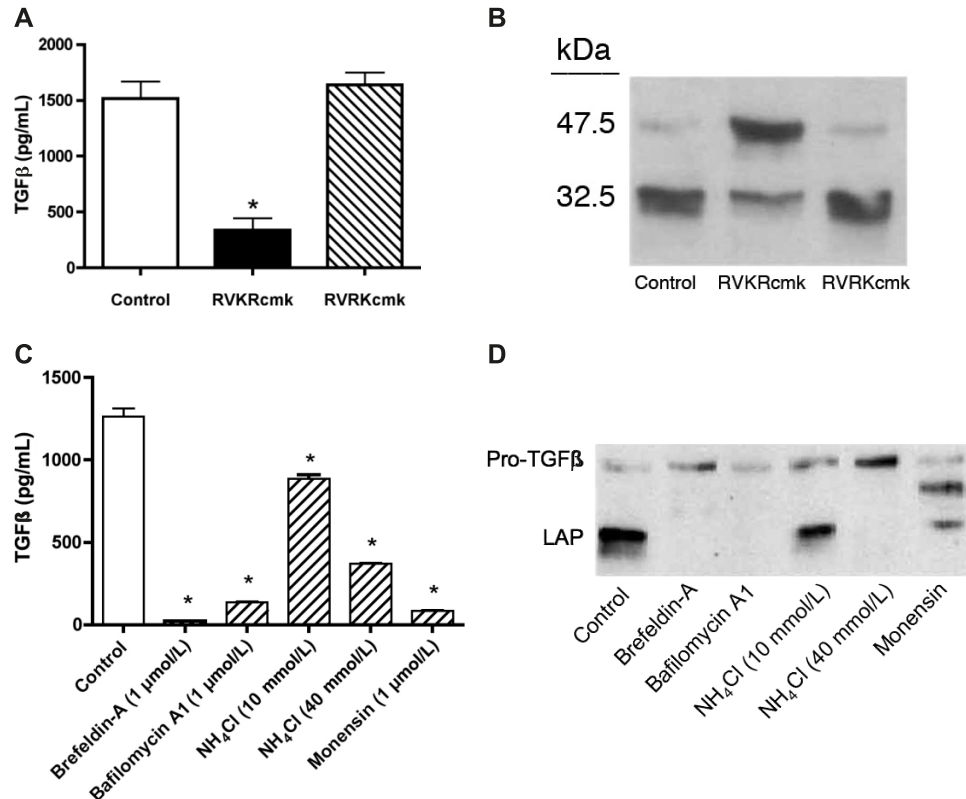
The results are presented as means $\pm$ SE. Data with multiple group comparisons were analyzed by using ANOVA and the post hoc Bonferroni test (GraphPad Prism version 4.0, San Diego, Calif.).

Results

Pro-TGF β processing by furin-like proteinase activity in A549 and T98G cells

TGF β 1 levels in the supernatant from A549 cells were significantly reduced following treatment with RVKRcmk (336.3 ± 108.6 pg/mL) compared with media alone ($1,518.0 \pm 152.0$ pg/mL) or the control peptide RVRKcmk (1639 ± 113.9 pg/mL), as determined by ELISA (Fig. 1A). The cell supernatants were also analyzed by Western blot to determine the apparent molecular ratio of the LAP as an index of TGF β proprotein processing (shown in Fig. 1B is one representative result of 4 separate experiments). As can be

Fig. 1. Release of TGF β 1 and maturation of cellular pro-TGF β in A549 cells treated with RVKRCmk, an inhibitor of furin-like proteinase activity (A, B), and agents that disrupt or alkalinize the acidic trans-Golgi network (TGN)/endosomal compartment (C, D). Cells were incubated for 24 h in media alone (control), or with 10 μ mol/L RVKRCmk, 10 μ mol/L control peptide RVRKcmk, or the indicated concentrations of TGN/endosome modifiers. ELISA determination of TGF β 1 production was performed on the cell supernatant media (A, C), and Western blots were used to identify the migration of the latency-associated peptide (LAP) in its native pro-TGF β (upper bands) and cleaved LAP (lower bands) forms (B, D). Blots are representative of 4 separate experiments. Asterisk (*) indicates significant difference ($p < 0.001$) from control and RVRKcmk (A) or from control (B), $n = 4$ per condition.



seen, we detected an increase in the intensity of the bands migrating at 47.5 kDa, corresponding to pro-TGF β and a decrease in the intensity of the bands migrating at 32.5 kDa, corresponding to the LAP. The control peptide RVRKcmk had no effect on pro-TGF β processing.

Pro-TGF β processing depended on the TGN/endosome system and its acidic milieu

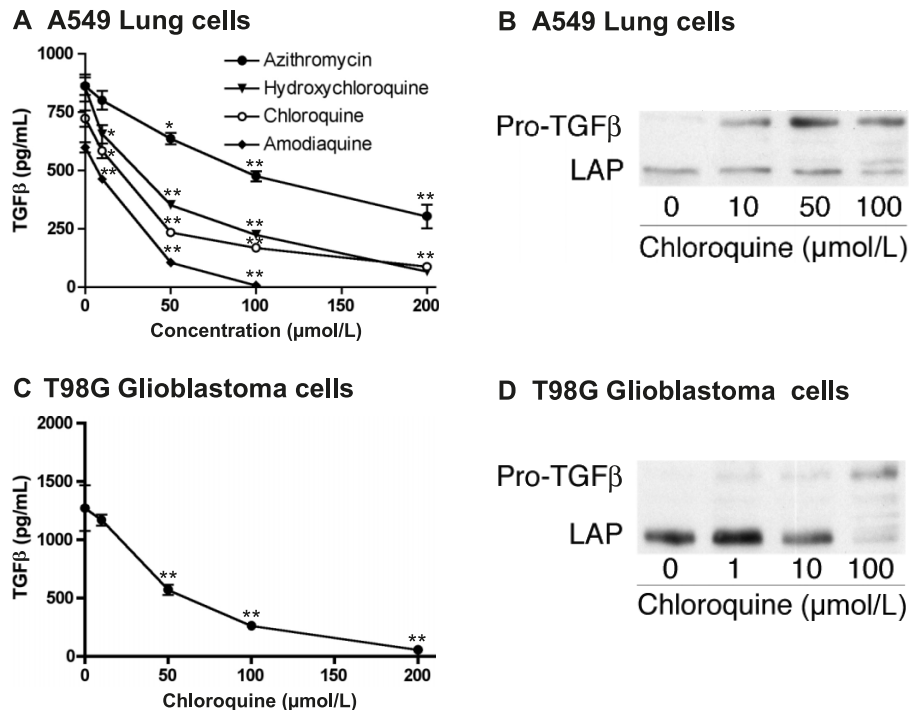
To verify whether the acidic milieu in the TGN/endosome is important for pro-TGF β maturation, we measured secreted TGF β 1 after incubating cells with several agents known to alkalinize TGN/endosomal pH (Fig. 1C). Extremely low levels of mature TGF β 1 were detected in the supernatants of A549 cells incubated with the TGN-disrupting brefeldin A at 1 μ mol/L (21.8 ± 1.5 pg/mL) compared with control (1264.0 ± 48.5 pg/mL). In addition, the other agents also significantly decreased levels of TGF β 1 in the cell media. Inhibition of the vesicular ATPase proton pump with bafilomycin A1, addition of the lysosomotropic weak base NH $_4$ Cl, and dissipation of pH gradients with the ionophore monensin each resulted in a marked decrease of TGF β 1 secretion (Fig. 1C) (bafilomycin A1, 137.5 ± 5.1 pg/mL; 10 mmol/L NH $_4$ Cl, 887.8 ± 21.1 pg/mL; 40 mmol/L NH $_4$ Cl, 373.3 pg/mL; monensin, 86.8 ± 3.3 pg/mL). The mechanism by which these conditions suppressed mature TGF β secretion was associated with inhibition of the proprotein processing,

as shown by a marked reduction in the intensity of the bands migrating at 47.5 kDa, corresponding to pro-TGF β and a decrease in the intensity of the bands migrating at 32.5 kDa, corresponding to the LAP. The control peptide RVRKcmk had no effect on pro-TGF β processing.

Drugs that are lysosomotropic weak bases suppressed TGF β 1 maturation

Chloroquine, hydroxychloroquine, amodiaquine, and azithromycin are weak bases that bear a tropism for the acidic TGN/endosomal compartment. Each of these drugs showed a concentration-dependent suppression of TGF β 1 processing by either alveolar epithelial cells (Fig. 2A) or glioblastoma cells (Fig. 2C). For A549 cells, the values at 50 μ mol/L were (in pg/mL): amodiaquine 105.6 ± 3.3 , control (media alone) 598.3 ± 21.4 ; chloroquine 234.2 ± 5.9 , control 721.9 ± 34.7 ; hydroxychloroquine 351.4 ± 19.1 , control 853.0 ± 58.5 ; azithromycin 635.5 ± 24.5 , control 861.3 ± 37.2 . For T98G cells, the value at 50 μ mol/L chloroquine was 571.4 ± 44.0 pg/mL versus control 1273.3 ± 196.7 pg/mL. As can be seen, the increasing concentrations of chloroquine were associated with an increase in the pro-TGF β band on Western blots (corresponding to the band migrating at 47.5 kDa) from each cell line, indicating that proprotein processing was decreased (Figs. 2B and 2D, one representative result of 4 separate experiments).

Fig. 2. Alkalinizing lysosomotropic drugs affect TGF β 1 proteolytic processing of pro-TGF β protein in A549 cells (A, B) and T98G glioblastoma cells (C, D) at 24 h. The TGF β 1 concentrations in the cell media were measured by ELISA (A, C). Pro-TGF β processing was assessed by Western blot using an antibody that recognizes the latency-associated peptide (B, D). Significant at *, $p < 0.05$ and **, $p < 0.01$ vs. control (media alone), $n = 4$. Blots are representative of at least 4 separate experiments.



Kinetics of chloroquine effect on TGF β 1 production

Incubation of A549 cells with 100 μ mol/L chloroquine decreased the concentration of TGF β 1 detectable in the A549 supernatants as early as 60 min and this effect was sustained at 24 h (Fig. 3). The values (in pg/mL) at 60 min were chloroquine 14.8 ± 1.3 compared with control 28.9 ± 0.7 ; at 24 h, chloroquine 145.2 ± 7.8 compared with control 1121.8 ± 82.1 .

Inhibition of pro-TGF β processing suppressed Smad2 phosphorylation

Active TGF β in the extracellular space binds to its receptor and initiates phosphorylation of Smad proteins, as observed in the A549 lysates illustrated in the Western blot specific for phosphorylated Smad2 protein (Fig. 3C). Inhibition of pro-TGF β processing with RVKRCmk markedly suppressed Smad2 phosphorylation. Similarly, 100 μ mol/L chloroquine induced a decrease in Smad2 phosphorylation in A549 cells.

Chloroquine decreased TGF β in BAL fluid of mice

Mice treated with daily subcutaneous injections of 1 mg chloroquine showed a significant decrease in BAL fluid TGF β levels (Fig. 4). The values shown are chloroquine 315.0 ± 22.5 pg/mg protein and control 430.3 ± 45.3 pg/mg protein. We observed no adverse effects of the chloroquine administration.

Discussion

Proteases of the subtilisin-like proprotein convertase family, such as furin, are involved in TGF β maturation and may

therefore be considered as important targets in many diseases. The furin-like proteinase inhibitor RVKRCmk has been reported to suppress TGF β 1 and TGF β 2 release into the extracellular milieu of glioblastoma cells (Leitlein et al. 2001). Our observations in alveolar epithelial cells indicated that RVKRCmk alone is sufficient to markedly suppress production of bioactive TGF β (by abolishing processing) and consequently Smad2 phosphorylation. The effect was specific to the RVKR sequence, a recognition site for furin-mediated cleavage, since RVKRCmk had no effect. Thus, because furin-like proteinase activity is located mostly in the TGN/endosomal system, we focused our investigations on interventions affecting this cellular compartment.

Brefeldin A, which disrupts the Golgi apparatus and the TGN/endosome compartment, abolished pro-TGF β processing and consequently lowered the levels of bioactive TGF β 1 in the media of treated cells. We then hypothesized that agents affecting the mildly acidic TGN/endosomal compartment may also decrease TGF β 1 maturation by preventing cleavage of the TGF β proprotein. This was confirmed by using 3 different approaches to alkalinize the TGN/endosomal pH: (i) a vesicular ATPase proton pump inhibitor, bafilomycin A1, (ii) a lysosomotropic weak base, ammonium chloride, and (iii) an ionophore that dissipates vesicular pH, monensin. Our results indicated that alkalinization of the mildly acidic TGN/endosomal compartment can be an effective approach to control TGF β processing in mammalian cells.

Recently, Perkett et al. have reported that bronchial epithelial cells expressing the Δ F508 mutant of cystic fibrosis transmembrane conductance regulator (CFTR) produce increased amounts of TGF β and that this abnormal production

Fig. 3. Time course of chloroquine-mediated inhibition of TGF β 1 accumulation in the extracellular milieu and effect of chloroquine on Smad2 phosphorylation. A549 cells were incubated with media alone (control) or 100 μ mol/L chloroquine for (A) 0–120 min and (B) 6–24 h. (C) Western blots of cells incubated 24 h in the presence of 10 μ mol/L RVKRemk or 0–100 μ mol/L chloroquine, washed, and lysed. Specific antibodies against phosphorylated Smad2 protein were used to determine the state of Smad2 phosphorylation. Significant at *, $p < 0.05$ and **, $p < 0.01$ vs. control, $n = 4$ per condition.

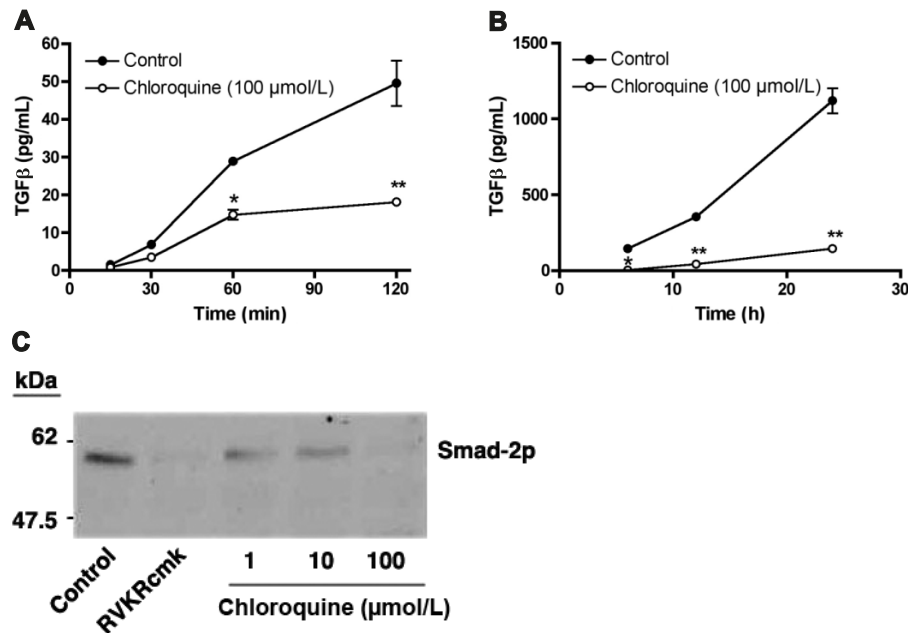
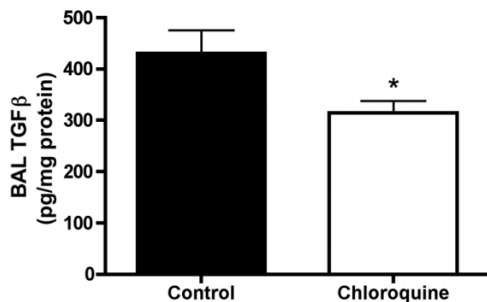


Fig. 4. TGF β in bronchoalveolar lavage fluid (BAL) from the lungs of mice treated with daily subcutaneous injections of 0.9% saline (control) or 1 mg chloroquine for 7 days. Levels of TGF β in BAL were determined by ELISA. Asterisk (*) indicates significant difference ($p < 0.05$) from control, $n = 11$ per group.



can be blocked by chloroquine (Perkett et al. 2006). The current study provides evidence that chloroquine decreases pro-TGF β post-translational processing. Several drugs are lysosomotropic weak bases. These include chloroquine, amodiaquine, hydroxychloroquine, and azithromycin, all of which have a long history of use in humans and a known safety profile. Each of these drugs showed a concentration-dependent inhibition of pro-TGF β protein processing and TGF β 1 secretion in A549 and T98G cells. Our results indicated that the effects of chloroquine are not specific to cells expressing Δ F508 CFTR, and we speculate that this class of drugs may have the potential for broader application in other conditions associated with furin-mediated proprotein cleavage.

The suppressive effect of chloroquine on pro-TGF β processing was observed within the first 2 h after treatment. The

effect of the lysosomotropic drugs was not through a generalized decrease in protein secretion because increased pro-TGF β protein was detected by Western blot in the cell supernatants. Although our work did not allow us to identify the mechanism by which lysosomotropic weak bases prevent pro-TGF β processing and TGF β 1 secretion, one possibility is that an increase of the TGN/endosomal pH may affect the capacity of furin to complete its autocatalytic activation. Furin is synthesized as an inactive proprotein, and the first step in its activation consists of an autocatalytic prodomain excision from the mature protein in the ER, a prerequisite for the efficient furin egress from this compartment. After the initial cleavage, the prodomain remains tightly associated with the convertase, preventing protein activity. When this stable furin–prodomain complex reaches the TGN, the increase in H $^{+}$ and (or) Ca $^{2+}$ concentration triggers a secondary cleavage, resulting in the dissociation of the prosegment and, consequently, the activation of the enzyme (Anderson et al. 1997; Bassi et al. 2005; Leduc et al. 1992). These steps in furin activation occur within 2 h (Anderson et al. 1997). Almost complete inhibition of furin cleavage has been reported with chloroquine in the Madin–Darby bovine kidney (MDBK) cell line (Vey et al. 1994). Although other mechanisms may be involved, the rapid inhibition of TGF β 1 secretion we observed is consistent with a pH-dependent interference of furin activation. The dependence of protein maturation on endosomal pH is not specific to pro-TGF β , as other TGN proteins such as TGN38 and foreign viral proteins undergo pH-dependent processing with furin-like proteases (Chapman and Munro 1994; Li et al. 2008; Yu et al. 2008).

Chloroquine, and other lysosomotropic agents, are concentrated in tissues. In animals, concentrations of chloro-

quine 10–700 times the plasma concentration may be detected in several tissues, including the lung and brain (Adelusi and Salako 1982). In the current study, TGF β levels in mice at the alveolar epithelial surface, as determined by BAL, were decreased after 1 week of subcutaneous administration of chloroquine at a dose that appeared to be well tolerated. The decrease was more modest in vivo than in vitro, most likely due to the lower tissue concentrations of the drug. Further studies are needed to determine whether similar results can be achieved in humans.

Inhibition of furin markedly decreases astrocytoma proliferation and invasiveness in vitro and thus has been identified as a promising therapeutic target for glioblastoma multiforme, a severe form of malignant astrocytic tumor (Mercapide et al. 2002; Wick et al. 2006). One indication that clinically meaningful inhibition of furin may be achievable by chloroquine in vivo comes from a clinical trial of chloroquine as an adjuvant therapy in patients with glioblastoma (Sotelo et al. 2006). The median survival of patients treated for glioblastoma with surgery, conventional chemotherapy, and radiation therapy was 11 months, whereas it increased to 24 months in patients who were given a daily dose of 150 mg chloroquine. The authors state that the results may be attributable to the antimutagenic properties of chloroquine (Briceno et al. 2003; Sotelo et al. 2006). We suggest that this remarkable improvement in survival may also be related, at least in part, to decreased pro-TGF β processing.

TGF β has been clearly shown to be sufficient to induce all of the histologic features of the IPF lung (Xu et al. 2003). Current therapies for adults with IPF are largely ineffective, possibly because none are known to affect TGF β synthesis or cell signaling (Gauldie 2002). In a recent clinical trial, however, patients with IPF showed modest improvement in some lung function parameters and had fewer respiratory exacerbations while taking the drug pirfenidone for 9 months (Azuma et al. 2005). Pirfenidone has recently been shown to inhibit furin in a malignant glioma cell line (Burghardt et al. 2007). The inhibition of pro-TGF β maturation we report in the current study was observed with chloroquine and amodiaquine concentrations 100-fold lower than those of pirfenidone.

The decrease in TGF β processing we report is most likely due to a class effect of the tested drugs, and in this context, other alkalinizing lysosomotropic drugs may also be of interest. In addition, alkalinizing lysosomotropic compounds may have the potential to treat patients with several diseases, including influenza (Ooi et al. 2006), in which furin-related processing may be important.

In summary, we have observed that the lysosomotropic drugs chloroquine, hydroxychloroquine, amodiaquine, and azithromycin suppress production of bioactive TGF β in lung alveolar epithelial cells and glioblastoma cells. Systemic chloroquine administration also decreases levels of TGF β in BAL fluid of mice. The mechanism would appear to be related to inhibition of pro-TGF β protein maturation and likely occurs through suppression of furin-like proteinase activity in the TGN/endosomal compartment. There is a pressing need for effective therapies in several fatal diseases characterized by an excess of TGF β activity, such as IPF and glioblastoma multiforme. Because of their availabil-

ity, low-cost, and well-defined safety profile, alkalinizing lysosomotropic drugs warrant consideration for clinical studies of TGF β -related diseases.

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