



Pergamon

Development of an Orexin-2 Receptor Selective Agonist, [Ala¹¹, D-Leu¹⁵]orexin-B

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Received 29 July 2002; accepted 21 September 2002

Abstract—Investigation of L-alanine and D-amino acid replacement of orexin-B revealed that three L-leucine residues at the positions of 11, 14, and 15 in orexin-B were important to show selectivity for the orexin-2 receptor (OX₂) over the orexin-1 receptor (OX₁). L-Alanine substitution at position 11 and D-leucine substitution at positions 14 and 15 maintained the potency of orexin-B to mobilize [Ca²⁺]_i in CHO cells expressing the OX₂, while their potency for the OX₁ was significantly reduced. In combined substitutions, we identified that [Ala¹¹, D-Leu¹⁵]orexin-B showed a 400-fold selectivity for the OX₂ (EC₅₀ = 0.13 nM) over OX₁ (EC₅₀ = 52 nM). [Ala¹¹, D-Leu¹⁵]orexin-B is a beneficial tool for addressing the functional roles of the OX₂.
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Recently, two novel hypothalamic neuropeptides, orexin-A and orexin-B, have been isolated from rat brain extracts as ligands for orphan G protein coupled receptors, namely, orexin-1 receptor (OX₁) and orexin-2 receptor (OX₂), which are exclusively expressed in the brain.¹ Both peptides are derived from the same precursor protein by proteolytic processing and are mainly presented in the lateral and posterior sections of the hypothalamus, which is known to be the center for feeding regulation.^{1,2} Orexin-A, which is a 33-residue peptide with two intramolecular disulfide bridges, shows similar potency for both the OX₁ and the OX₂, whereas orexin-B, which is a 28-amino acid-linear peptide, showed about a 10-fold selectivity for the OX₂ over the OX₁.¹

Several investigations have shown that orexins are involved in a variety of physiological functions, such as the regulation of locomotor activity, the sleep/awake cycle, blood pressure, metabolic rate, gastric acid secretion, pituitary luteinizing hormone secretion, and feeding regulation.^{3–9} Different expression patterns of OX₁ and OX₂ mRNAs support the diverse functions of orexins.¹⁰ Using potent and selective antagonists, the roles of OX₁ are going to be elucidated.¹² However, the lack of

selective ligands for the OX₂ makes it difficult to address the role of the OX₂.

To develop potent and selective OX₂ ligands, we conducted L-alanine and D-amino acid replacements to emphasize the intrinsic selectivity of orexin-B (Fig. 1).

Peptide Preparation

Orexin-A and -B were purchased (Peptide Institute, Inc, Osaka, Japan). All orexin-B analogue peptides were synthesized on the solid support using a Fmoc/^tBu strategy on PioneerTM peptide synthesizer (Applied Biosystems, Foster City, CA, USA) or ACT357MPS (Advanced ChemTech, Louisville, KY, USA). Fmoc-NovaSyn[®] TGR resin (Novabiochem, Laufelfingen, Switzerland) was used as a starting material. All crude peptides were cleaved from resins with TFA/thioanisole/ethanedithiol/*m*-cresol within 2 h, followed by the addition of cold diethyl ester for solidification. The resulting crude peptides, obtained after centrifugation, were purified with preparative reversed-phase HPLC (Hitachi D-6000 system, Hitachi, San Jose, CA, USA) with a YMC-Pack ODS-AQ column (20 × 250 mm; YMC, Kyoto, Japan). All peptides were characterized by analytical HPLC (HP 1100LC system, Agilent Technologies, Palo Alto, CA, USA) with a YMC-Pack ODS-AQ column (2.0 × 150 mm; YMC, Kyoto, Japan) and by electrospray mass

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spectrometry (Waters Platform ZMD2000, Waters, Millford, MA, USA).

Measurement of Intracellular Calcium Ion Concentrations

CHO-K1 cells expressing each subtype of orexin receptors (OX₁ and OX₂) were maintained by a general method. The intracellular calcium ion concentrations were measured by the method previously reported.³ In brief, CHO-K1 cells stably expressing human OX₁ or OX₂ receptors (OX₁-CHO or OX₂-CHO) were seeded into 96-well plates and incubated with a cytoplasmic calcium indicator, Fluo-3 AM. After free dye was washed away, the intracellular Ca²⁺ mobilization evoked by orexins and related peptides was monitored as the change in cell fluorescence intensity by FLIPR (Molecular Devices, Sunnyvale, CA, USA). The lack of agonistic activities of synthesized peptides in the intact CHO-K1 cells were confirmed to address their specificity for orexin receptors (data not shown).

L-Alanine Scan of Orexin-B

To address the importance of each residue with minimal impact to the three-dimensional structure, alanine-scanned analogues of orexin-B were synthesized. Three alanine residues at positions 17, 22, and 23 in the orexin-B sequence were replaced with glycine. Their potency to mobilize intracellular Ca²⁺ concentrations in OX₁-CHO and OX₂-CHO cells were measured (Table 1). The substitutions at positions 1–9, 12–14, 16, and 17 did not significantly affect the intrinsic potency of orexin-B for both the OX₁ and the OX₂. In contrast, replacement at positions 10, 15, and 18–28 considerably decreased the potency for both receptors. Among these alanine-scanned analogues, [Ala¹¹]orexin-B showed unique profile. The potency of [Ala¹¹]orexin-B was selectively reduced for the OX₁ (EC₅₀ = 8.1 nM), but not the OX₂ (EC₅₀ = 0.066 nM), and resulted in a 120-fold selectivity for the OX₂. Additional substitutions using 12 usual and 10 unusual amino acids at position 11 showed that serine and tryptophan were also efficacious in emphasizing the selectivity of orexin-B to a similar extent of alanine (Table 1).

D-Amino Acid Replacement of Orexin-B

To change the orientation of the amino acid side chain with minimal impact on the overall hydrophobicity and dipole moment, we conducted D-amino acid replacement (Table 2). D-Alanine was used as a substitute for glycine

in the sequence of orexin-B. D-Amino acid substitutions at positions 23–28 showed significant reduction of the potency for both orexin receptors. In addition, D-leucine substitutions at positions 14 and 15 significantly improved the selectivity of orexin-B by 5- and 8-fold, respectively.

Combination Study on the Alanine Scan and D-Amino Acid Replacement

Each positional scan showed that three leucine residues, Leu¹¹, Leu¹⁴, and Leu¹⁵, in the orexin-B sequence were important for the activation of the OX₁, and that modification of these sites was relatively tolerable for the activation of the OX₂. Based on these results, combination replacements of orexin-B were performed to determine whether or not synergistic effects would be observed (Table 3). Among the synthesized peptides, the combination of an L-alanine

Table 1. Biological activity and receptor selectivity of L-alanine or glycine substituted orexin-B analogues^a

Peptide	EC ₅₀ value (nM)		Selectivity OX ₁ /OX ₂
	OX ₁	OX ₂	
Orexin-B	0.83 ± 0.041	0.081 ± 0.0091	10
[Ala ¹]orexin-B	2.3 ± 0.24	0.20 ± 0.055	12
[Ala ²]orexin-B	2.4 ± 0.55	0.22 ± 0.063	11
[Ala ³]orexin-B	2.3 ± 0.81	0.18 ± 0.072	13
[Ala ⁴]orexin-B	3.2 ± 1.2	0.23 ± 0.096	14
[Ala ⁵]orexin-B	3.1 ± 0.68	0.29 ± 0.10	11
[Ala ⁶]orexin-B	1.6 ± 0.21	0.12 ± 0.025	14
[Ala ⁷]orexin-B	1.2 ± 0.13	0.12 ± 0.0083	9.2
[Ala ⁸]orexin-B	2.7 ± 0.20	0.46 ± 0.10	5.9
[Ala ⁹]orexin-B	1.1 ± 0.20	0.20 ± 0.029	5.5
[Ala ¹⁰]orexin-B	9.1 ± 1.6	0.77 ± 0.086	12
[Ala ¹¹]orexin-B	8.1 ± 0.72	0.066 ± 0.011	120
[Ala ¹²]orexin-B	3.4 ± 0.53	0.23 ± 0.045	15
[Ala ¹³]orexin-B	1.7 ± 0.30	0.13 ± 0.026	13
[Ala ¹⁴]orexin-B	0.82 ± 0.18	0.065 ± 0.021	13
[Ala ¹⁵]orexin-B	39 ± 4.6	2.2 ± 0.33	18
[Ala ¹⁶]orexin-B	1.5 ± 0.33	0.13 ± 0.042	12
[Gly ¹⁷]orexin-B	0.41 ± 0.087	0.091 ± 0.035	4.5
[Ala ¹⁸]orexin-B	6.0 ± 2.4	1.6 ± 0.97	3.8
[Ala ¹⁹]orexin-B	14 ± 3.0	1.1 ± 0.43	13
[Ala ²⁰]orexin-B	32 ± 7.4	2.3 ± 1.1	14
[Ala ²¹]orexin-B	5.7 ± 1.7	0.72 ± 0.41	7.9
[Gly ²²]orexin-B	7.5 ± 1.1	0.27 ± 0.083	28
[Gly ²³]orexin-B	22 ± 4.3	0.84 ± 0.13	26
[Ala ²⁴]orexin-B	680 ± 73	290 ± 92	2.3
[Ala ²⁵]orexin-B	> 10,000	390 ± 92	—
[Ala ²⁶]orexin-B	710 ± 91	140 ± 34	5.1
[Ala ²⁷]orexin-B	110 ± 27	9.7 ± 1.8	11
[Ala ²⁸]orexin-B	220 ± 40	15 ± 2.6	15
[Ser ¹¹]orexin-B	150 ± 41	1.2 ± 0.57	120
[Trp ¹¹]orexin-B	4.8 ± 1.4	0.15 ± 0.095	32

^aData represent the mean of more than three independent determinations performed in duplicate.

Peptide	Sequence
orexin-A	<EPLPDCCRQKTCSCRLYELLHGAGNHAAGILTL-NH ₂
orexin-B	RSGPPGLQGRLQRLQLQASGNHAAGILTM-NH ₂

Figure 1. Amino acid sequences of orexins.

Table 2. Biological activity and receptor selectivity of corresponding D-amino acid substituted orexin-B analogues^a

Peptide	EC ₅₀ value (nM)		Selectivity OX ₁ /OX ₂
	OX ₁	OX ₂	
Orexin-B	0.83±0.041	0.081±0.0091	10
[D-Arg ¹]orexin-B	1.6±0.80	0.19±0.15	8.2
[D-Ser ²]orexin-B	1.2±0.60	0.15±0.10	8.3
[D-Ala ³]orexin-B	1.3±0.49	0.16±0.090	8.4
[D-Pro ⁴]orexin-B	0.89±0.35	0.15±0.092	5.8
[D-Pro ⁵]orexin-B	0.85±0.39	0.16±0.12	5.3
[D-Ala ⁶]orexin-B	1.4±0.51	0.20±0.16	7.1
[D-Leu ⁷]orexin-B	1.4±0.37	0.084±0.044	16
[D-Gln ⁸]orexin-B	4.0±1.3	0.32±0.18	13
[D-Ala ⁹]orexin-B	1.4±0.63	0.13±0.089	10
[D-Arg ¹⁰]orexin-B	9.8±3.6	0.87±0.62	11
[D-Leu ¹¹]orexin-B	7.0±0.71	0.60±0.24	12
[D-Gln ¹²]orexin-B	0.77±0.26	0.11±0.078	6.8
[D-Arg ¹³]orexin-B	3.8±1.2	0.41±0.29	9.4
[D-Leu ¹⁴]orexin-B	34±3.0	0.44±0.060	85
[D-Leu ¹⁵]orexin-B	2.7±0.26	0.052±0.0051	52
[D-Gln ¹⁶]orexin-B	5.8±1.6	0.41±0.27	14
[D-Ala ¹⁷]orexin-B	2.6±0.77	0.26±0.19	10
[D-Ser ¹⁸]orexin-B	7.9±0.87	0.46±0.22	17
[D-Ala ¹⁹]orexin-B	0.52±0.12	0.16±0.069	3.2
[D-Asn ²⁰]orexin-B	0.61±0.14	0.26±0.14	2.4
[D-His ²¹]orexin-B	7.3±3.9	1.0±0.75	6.9
[D-Ala ²²]orexin-B	22±4.4	2.6±1.7	8.3
[D-Ala ²³]orexin-B	230±12	14±6.7	16
[D-Ala ²⁴]orexin-B	400±66	67±23	6.0
[D-Ile ²⁵]orexin-B	> 10,000	1400±1200	—
[D-Leu ²⁶]orexin-B	> 10,000	200±170	—
[D-Thr ²⁷]orexin-B	> 10,000	260±52	—
[D-Met ²⁸]orexin-B	> 10,000	290±110	—

^aData represent the mean of more than three independent determinations performed in duplicate

substitution at Leu¹¹ and a D-leucine substitution at Leu¹⁵ showed a good selectivity (400-fold) without significant reduction of the potency for the OX₂.

Recently, the 3-D structure of human [Phe¹]orexin-B was determined using NMR analysis.¹¹ The structure of orexin-B consisted of a random coil (positions 1–7), kink region (positions 20–22), and two α -helices (positions 7–19 and 23–28). These two helices form a bent structure. Our results from L-alanine and D-amino acid substitutions suggest that modification at positions 23–28 significantly reduced the potency of orexin-B for both OX₁ and OX₂. Amino acid residues in the C-terminal portion were well conserved between orexin-A and orexin-B. In addition, C-terminal truncated orexins drastically lost their potency for both orexin receptors (ref 13, unpublished our observations). Therefore, the second helix, consisting of amino acid residues 23–28, might be a common core needed to recognize and activate orexin receptors. Moreover, the changes at position 11 (L-leucine to L-alanine) and at positions 14 and 15 (L-leucine to D-leucine) located in the 1st helix selectively reduce the potency for the OX₁. Thus, the 1st helix might play a key role in determining the selectivity of orexin analogues. In addition, interestingly, the side chains of Leu¹¹ and Leu¹⁵ faced to the same direction in the 1st helix.¹¹ To consider the drastically decreased potency of [Ala¹¹, D-Leu¹⁵]orexin-B for the OX₁, the surface of the 1st helix constructed by Leu¹¹ and Leu¹⁵ might be of interest.

Table 3. Biological activity and receptor selectivity of two-point substituted orexin-B analogues^a

Peptide	EC ₅₀ value (nM)		Selectivity OX ₁ /OX ₂
	OX ₁	OX ₂	
Orexin-B	0.83±0.041	0.081±0.0091	10
[Ala ¹¹ ,D-Leu ¹⁴] orexin-B	270±13	1.9±0.28	140
[Ala ¹¹ ,D-Leu ¹⁵] orexin-B	52±3.1	0.13±0.012	400

^aData represent the mean of more than three independent determinations performed in duplicate

In conclusion, [Ala¹¹, D-Leu¹⁵]orexin-B identified in the course of the L-alanine and D-amino acid replacements of orexin-B is a potent and selective OX₂ agonist. A novel agonist, [Ala¹¹, D-Leu¹⁵]orexin-B should be a promising tool for addressing the roles of OX₂.

Acknowledgements

We thank Shinnosuke Abe and Hirokazu Ohsawa for the analysis of mass spectra. We are also grateful to Ms. K. Marcopul and Ms. J. Lowry, Merck & Co., for her critical reading of this manuscript.

References and Notes

- Sakurai, T.; Amemiya, A.; Ishii, M.; Matsuzaki, I.; Chemelli, R. M.; Tanaka, H.; Williams, S. C.; Richardson, J. A.; Kozlowski, G. P.; Wilson, S.; Arch, J. R. S.; Buckingham, R. E.; Haynes, A. C.; Carr, S. A.; Annan, R. S.; McNulty, D. E.; Liu, W. S.; Terrett, J. A.; Elshourbagy, N. A.; Bergsma, D. J.; Yanagisawa, M. *Cell* **1998**, *92*, 573.
- Nambu, T.; Sakurai, T.; Mizukami, K.; Hosoya, Y.; Yanagisawa, M.; Goto, K. *Brain Res.* **1999**, *827*, 243.
- Okumura, T.; Takeuchi, S.; Motomura, W.; Yamada, H.; Egashira, S.; Asahi, S.; Kanatani, A.; Ihara, M.; Kohgo, Y. *Biochem. Biophys. Res. Commun.* **2001**, *280*, 976.
- Pu, S.; Jain, M. R.; Kalra, P. S.; Karla, S. P. *Regul. Pept.* **1998**, *78*, 133.
- Lin, L.; Faraco, J.; Li, R.; Kadotani, H.; Rogers, W.; Lin, X.; Qiu, X.; de Jong, P. J.; Nishino, S.; Mignot, E. *Cell* **1999**, *98*, 365.
- Chemelli, R. M.; Willie, J. T.; Sinton, C. M.; Elmquist, J. K.; Scammell, T.; Lee, C.; Richardson, J. A.; Williams, S. C.; Xiong, Y.; Kisanuki, Y.; Fitch, T. E.; Nakazato, M.; Hammer, R. E.; Saper, C. B.; Yanagisawa, M. *Cell* **1999**, *98*, 437.
- Ida, T.; Nakahara, K.; Katayama, T.; Murakami, N.; Nakazato, M. *Brain Res.* **1999**, *821*, 526.
- Lubkin, M.; Stricker-Krongrad, A. *Biochem. Biophys. Res. Commun.* **1998**, *253*, 241.
- Samson, W. K.; Gosnell, B.; Chang, J. K.; Resch, Z. T.; Murphy, T. C. *Brain Res.* **1999**, *831*, 248.
- Trivedi, P.; Yu, H.; MacNeil, D. J.; Van der Ploeg, L. H. T.; Guan, X. M. *FEBS Lett.* **1998**, *438*, 71.
- Lee, J. H.; Bang, E.; Chae, K. J.; Kim, J. Y.; Lee, D. W.; Lee, W. *Eur. J. Biochem.* **1999**, *266*, 831.
- Haynes, A. C.; Jackson, B.; Chapman, H.; Tadayyon, M.; Johns, A.; Porter, R. A.; Arch, J. R. *Regul. Pept.* **2000**, *96*, 45.
- Darker, J. G.; Porter, R. A.; Eggleston, D. S.; Smart, D.; Brough, S. J.; Sabido-David, C.; Jerman, J. C. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 737.